MANUAL FOR THE IDENTIFICATION OF GRASS STEMBORERS AND ASSOCIATED PARASITOIDS IN KENYA



Alberto T. Barrion, Robert S. Copeland and Zeyaur R. Khan

MANUAL FOR THE **IDENTIFICATION OF GRASS** STEMBORERS AND ASSOCIATE PARASITOIDS IN KENYA

Alberto T. Barrion, Robert S. Copeland and Zeyaur R. Khan icipe - African Insect Science for Food and Health









ACKNOWLEDGEMENTS

Financial assistance for the production of this handbook was provided by the United Nations Environment Programme (UNEP)/Global Environment Facility (GEF) through the project GFL/2711-01-4345, "Conservation of Gramineae and Associated Arthropods for Sustainable Agricultural

Development in Africa".

We are indebted to Dr F. N. Muyekho and the GEF/Grass technicians, Silas Ouko, Raphael Odhiambo and Michael Watiti, based at the *icipe*-Thomas Odhiambo Campus, Mbita field station, Kenya, for the field collection, laboratory rearing and curatorial processing of the specimens used in this manual. Eshmael Kidiavai and Peter Ollimo performed similar services in Machakos and Busia districts, respectively. Special gratitude is given to Mr Gordon Ochieng, Mbita Point, Kenya for the grass stemborer illustrations and to Mr Ruel Medina, Santa Rosa, Laguna, Philippines for drawing the diopsid, mordellid and languriid stemborers and their associated insects (parasitoids and hyper-parasitoids). Mrs Jessamyn Adorada, University of the Philippines at Los Baños, Philippines drew the *Hypamazso* beetles and their parasitoids.

We thank Pascal Moyal for confirming identifications or providing new ones for the Noctuidae we reared. Jay Shaffer helped us in a similar way with the peoriine pyralids. We are especially grateful to Miss Aimee Lynn A. Barrion for taking digital photographs of some of the insects and of the illustrations. Ms Karie Darrow of the Department of Entomology, National Museum of Natural History, Smithsonian Institution, Washington, DC, took time out from her busy schedule to photograph some of the adult moths and their genitalia. Dr Wanja Kinuthia, Department of Invertebrate Zoology, National Museums of Kenya, Nairobi provided access to

the insect collections and facilitated the loan of specimens.

Manual for the Identification of Grass Stemborers and Associated Parasitoids in Kenya

by

Alberto T. Barrion, Robert S. Copeland and Zeyaur R. Khan

ISBN: 92 9064 189 4

Copyright © 2007 International Centre of Insect Physiology and Ecology All rights reserved.

> Published and printed by: icipe Science Press P. O. Box 72913-00200 Nairobi, Kenya Tel: +254 (20) 8632000 Fax: +254 (20) 8632001/2 isp@icipe.org www.icipe.org

Editorial Assistance: D. Osogo Cover Design and Layout: I. Ogendo

CONTENTS

Foreword
List of Platesv
List of Figuresv
INTRODUCTION
Composition of the Manual
THE INSECT ORDERS REARED FROM WILD AND CULTIVATED GRASSES
Key to the insect orders associated with Gramineae in Kenya, based on adult characters
Key to the insect orders associated with Gramineae in Kenya, based on larval characters
Key to the insect orders associated with Gramineae in Kenya, based on pupal characters
THE STEMBORERS
Lepidoptera: Morphology of a Lepidopteran Stemborer
Key to the families of lepidopteran stemborers, based on adult characters
Key to the families of lepidopteran stemborers, based on larval characters
Family Cossidae
Family Gelechiidae
Family Noctuidae
Key to the adults of noctuid stemborers
Key to the larvae of common noctuid stemborers
Key to the pupae of common noctuid stemborers
Family Pyralidae
Key to the adults of pyralid stemborers
Key to the larvae of common pyralid stemborers1
Key to the pupae of common pyralid stemborers1
Family Tortricidae1
Coleoptera: Morphology of a Coleopteran Grass Stemborer
Key to the families of coleopteran stemborers, based on adult characters
Family Anthribidae 13
Key to the adults of anthribid stemborers
Family Cerambycidae
Key to the adults of cerambycid stemborers14
Key to the larvae and pupae of cerambycid stemborers14
Family Curculionidae15
Key to the adults of curculionid stemborers
Family Languriidae
Key to the adults of languriid stemborers
Family Mordellidae
Key to the adults of mordellid stemborers18

Family Tenebrionidae	19
Key to the larvae of tenebrionid stemborers	
Diptera: Morphology of a Dipteran Stemborer	20
Key to the families of dipteran stemborers	20
Family Chloropidae	20
Key to the larvae and pupae of chloropid stemborers	20
Family Diopsidae	21
Key to the adults of diopsid stemborers	21
Family Muscidae	21
Family Tephritidae Key to the adults of <i>Bistrispinaria</i> from wild grasses in Kenya (after Hancock, 1999)	22
THE PARASITOIDS	
General morphology of a parasitic hymenopteran	24
Key to the parasitoids associated with stemborers	24
Plates	
Figures	58
Appendix—List of Stemborers and Parasitoids Reared from Grasses	206
Glossary	209
Selected References	212

FOREWORD

This manual illustrates many of the stemborers and associated insects of grasses in Kenya, and provides keys for their identification. It can be used together with *icipe's* booklet *A Primer on Grass Identification and Their Uses in Kenya*, which provides diagnostic and pictorial information

on grass species that abound in and around agricultural lands in Kenya.

It is of paramount importance to identify stemborers and their natural enemies associated with grasses to implement ways of using native grasses to manage insect pests in farmers' fields. Wild grasses may act to repel or physically block pests of cereal crops, limiting access to crop plants. Grasses may also serve as a preferred habitat for beneficial insects and spiders that parasitise or prey on pest species. Proper management of wild grasses around crop fields can provide a natural resource of beneficial species of parasitoids and predators, increasing the chances of self-sustainable agriculture development among resource-poor farmers throughout Kenya. In this manual, the focus is on the pests and their parasitoids. As the technologies for self-regulatory pest management and sustainable agriculture grow more sophisticated and specific, correct identification of pest insects and their enemies is very important. However, there is presently no single manual available that offers users a tool to identify insects to at least the generic level. This manual addresses that need.

The primary target users of this manual are field or laboratory technicians and researchers. Technical language has been minimised as far as possible. However, many species of stemborers on grasses look similar, and identification based solely on external morphological features can be frustratingly difficult. This is particularly true for moth species of the genera *Chilo, Sesamia* and *Busseola*, especially when adult specimens are rubbed. Therefore, photographic images of the genitalia of most stem-boring moths have been included to provide help in identification to the species level. The illustrations and photos should facilitate the identification of stemborers and their associated insects reared from tall grasses. These data will help to determine the grass species that are important for conservation and maintenance of insect biodiversity.

I hope the manual will be a useful tool in identifying the stemborers on grasses and their natural enemies associated with the wild and cultivated grasses of Kenya, and that it also provides information useful for supporting self-regulatory pest management for sustainable

agriculture and the conservation of biological diversity.

C. Borgemeister
Director General,
icipe – African Insect Science for Food and Health

1 8

LIST OF PLATES

Plate 1. Chilo partellus, C. thyrsis, Eldana saccharina male genitalia and aedeagus	
Plate 2. Chilo incertus and Eldana saccharina female genitalia	
Plate 3. Chilo partellus, C. thyrsis female genitalia	
Plate 4. Peoriinae taxon A male genitalia and aedeagus	32
Plate 5. Peoriinae taxon A female genitalia with closeup of ovipositor	33
Plate 6. Maliarpha concinnella male genitalia, aedeagus	34
Plate 7. Ematheudes species: E. sp. nov., E. straminella male genitalia and aedeagus	35
Plate 8. Ematheudes species: Male costal spines and female genitalia of E. sp. nov. and	
E. straminella	36
Plate 9. Peoriinae taxon B male genitalia and aedeagus	37
Plate 10. Saluria lentistrigella and Sciomesa piscator male genitalia and aedeagus	38
Plate 11: Sesamia species: S. calamistis, S. nonagrioides botanephaga, S. penniseti,	00
S. poephaga, S. sp. nov. male antennae	39
Plate 12. Sesamia sp. nov. male genitalia and aedeagus; female genitalia	40
Plate 13. Sesamia calamistis: male genitalia and aedeagus	4 1
Plate 15. Sesamia nonagrioides botanephaga male genitalia and aedeagus	
Plate 16. Sesamia penniseti male genitalia	
Plate 17. Sesamia poephaga male genitalia and aedeagus	45
Plate 18. Sciomesa cf. sp. nov. male genitalia and aedeagus	
Plate 19. Busseola species: B. fusca, B. phaia, B. obliquifascia male genitalia and aedeagus	47
Plate 20. Busseola species: B. fusca, B. phaia, B. obliquifascia female genitalia	48
Plate 21. Manga nubifera male genitalia	
Plate 22. Manga nubifera male aedeagus; female genitalia	50
Plate 23. Eldana saccharina, Chilo partellus, C. thyrsis adult moths	51
Plate 24. Peoriinae taxon A, Peoriinae taxon B, Maliarpha concinnella adult moths	52
Plate 25. Ematheudes sp. nov., E. straminella, Saluria lentistrigella adult moths	
Plate 26. Sesamia species: S. calamistis, S. nonagrioides hotanephaga, S. nenniseti.	
	54
5. voevliaga, 5. sp. nov. adult moths	
S. poephaga, S. sp. nov. adult moths	55
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57 58
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57 58 58
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57 58 58 59
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57 58 59 60
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 59 60 61
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 59 60 61 62
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 59 60 61 62
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 59 60 61 62 62 63
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 58 58 59 60 61 62 62 63 64
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 58 58 59 60 62 62 63 64 64
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 58 58 59 60 62 62 63 64 64 65
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 69 61 62 62 63 64 64 65 66
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 59 61 62 62 63 64 65 65 66
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 57 58 58 69 61 62 62 63 64 65 66 66
Plate 27. Manga nubifera, Sciomesa piscator adult moths	55 56 57 58 59 61 62 62 63 64 65 65 66 67 68
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths. Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae. LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa. Figure 4 a-d. Stegasta sp. adult Figure 4 e-g. Stegasta sp. male genitalia, uncus and aedeagus Figure 5 a-d. Eldana saccharina Walker adult Figure 5 b-n. Eldana saccharina male genitalia and its aedeagus and female genitalia. Figure 5 o-r. Eldana saccharina pupa. Figure 6 ca-e. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male at rest. Figure 6 m-r. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 m-r. Chilo partellus larva Figure 6 m-r. Chilo partellus larva Figure 6 m-r. Chilo partellus larva Figure 6 m-r. Chilo partellus pupa	55 57 58 58 59 61 62 63 64 65 65 66 67 68 68
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths. Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae. LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa. Figure 4 a-d. Stegasta sp. adult Figure 4 e-g. Stegasta sp. male genitalia, uncus and aedeagus Figure 5 a-d. Eldana saccharina Walker adult Figure 5 b-n. Eldana saccharina male genitalia and its aedeagus and female genitalia. Figure 5 o-r. Eldana saccharina pupa. Figure 6 a-e. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 m-r. Chilo partellus larva Figure 6 s-u. Chilo partellus pupa Figure 6 s-u. Chilo partellus pupa Figure 7 a-d. Chilo orichalcociliellus Strand adult	55 56 57 58 59 61 62 63 64 65 65 66 67 68 68 69 70
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths. Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae. LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa Figure 4 a-d. Stegasta sp. adult Figure 4 e-g. Stegasta sp. male genitalia, uncus and aedeagus Figure 4 h-l. Stegasta sp. male genitalia, uncus and aedeagus Figure 5 a-d. Eldana saccharina Walker adult Figure 5 e-g. Eldana saccharina larva Figure 5 b-n. Eldana saccharina larva Figure 6 n-c. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male at rest Figure 6 f-i. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 s-u. Chilo partellus larva Figure 6 s-u. Chilo partellus pupa Figure 7 a-d. Chilo orichalcociliellus Strand adult Figure 7 e-g. Chilo orichalcociliellus Strand adult Figure 7 e-g. Chilo orichalcociliellus genitalia of male and aedeagus, and female genitalia	55 56 57 58 59 61 62 63 64 65 65 68 68 69 70
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths. Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae. LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa Figure 4 a-d. Stegasta sp. adult Figure 4 e-g. Stegasta sp. male genitalia, uncus and aedeagus. Figure 4 h-l. Stegasta sp. pupa Figure 5 a-d. Eldana saccharina Walker adult Figure 5 e-g. Eldana saccharina male genitalia and its aedeagus and female genitalia. Figure 5 h-n. Eldana saccharina larva. Figure 6 h-n. Eldana saccharina pupa Figure 6 f-i. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male at rest. Figure 6 f-i. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 m-r. Chilo partellus barva Figure 6 s-u. Chilo partellus pupa Figure 7 a-d. Chilo partellus pupa Figure 8 a-e. Chilo partellus pupa	55 56 57 58 59 60 61 62 63 64 65 65 68 68 68 69 71 71
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa Figure 4 a-d. Stegasta sp. male genitalia, uncus and aedeagus Figure 4 b-l. Stegasta sp. pupa Figure 5 a-d. Eldana saccharina Walker adult Figure 5 a-d. Eldana saccharina male genitalia and its aedeagus and female genitalia Figure 5 h-n. Eldana saccharina larva Figure 6 a-e. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male at rest Figure 6 m-r. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 m-r. Chilo partellus larva Figure 7 a-d. Chilo orichalcociliellus Strand adult Figure 7 a-d. Chilo orichalcociliellus Strand adult Figure 8 a-e. Chilo thyrsis Bleszynski adult. Figure 8 f-g. Chilo thyrsis male genitalia and aedeagus Figure 8 f-g. Chilo thyrsis male genitalia and aedeagus	55 56 58 58 69 62 62 63 64 65 65 66 67 68 69 71 71 71
Plate 27. Manga nubifera, Sciomesa piscator adult moths. Plate 28. Busseola species: B. fusca, B. phaia, B. obliquifascia adult moths. Plate 29. Busseola fusca, Chilo partellus, Sesamia calamistis eggs and larvae. LIST OF FIGURES Figure 1 a-c. Adult moth Figure 1 d-f. Legs I-III, tympanal organ in family Pyralidae and in Noctuidae Figure 1 g-i. Genitalia of male and its aedeagus and female genitalia. Figure 2 a-d. Lepidopteran larva Figure 3 a-c. Lepidopteran pupa Figure 4 a-d. Stegasta sp. adult Figure 4 e-g. Stegasta sp. male genitalia, uncus and aedeagus. Figure 4 h-l. Stegasta sp. pupa Figure 5 a-d. Eldana saccharina Walker adult Figure 5 e-g. Eldana saccharina male genitalia and its aedeagus and female genitalia. Figure 5 h-n. Eldana saccharina larva. Figure 6 h-n. Eldana saccharina pupa Figure 6 f-i. Chilo partellus (Swinhoe) female Figure 6 f-i. Chilo partellus male at rest. Figure 6 f-i. Chilo partellus male genitalia and aedeagus, and female genitalia Figure 6 m-r. Chilo partellus barva Figure 6 s-u. Chilo partellus pupa Figure 7 a-d. Chilo partellus pupa Figure 8 a-e. Chilo partellus pupa	55 58 58 58 61 62 62 63 64 65 65 67 68 67 71 71 72 73 73

Figure 9 a. Thaumatotibia leucotreta (Meyrick) adult	
Figure 9 b-f. Thaumatotibia leucotreta (Meyrick) pupa	75
Figure 10 a. Sesamia nonagrioides botanephaga Tams & Bowden female genitalia	76
Figure 10 b-f. Sesamia nonagrioides botanephaga pupa	76
Figure 11 a–f. Sesamia calamistis Hampson male	77
Figure 11 i-m. Sesamia calamistis larva	70
Figure 11 n-q. Sesamia calamistis pupa	70
Figure 12 a-f. Manga nubifera (Hampson) adult	80
Figure 12 a-i Manga nuhifera (Hampson) pupa	81
Figure 12 g-i. Manga nubifera (Hampson) pupa	81
Figure 13 f–j. Busseola fusca larva	82
Figure 13 k-m. Busseola fusca pupa	83
Figure 14 a. Phragmataecia boisduvalii Herrich-Shäffer adult	83
Figure 14 b-i. Phragmataecia boisduvalii larva	84
Figure 14 j-l. Phragmataecia boisduvalii pupa	85
Figure 15 a-h. Morphology of a beetle	
Figure 15 i-o. Coleopteran larva	87
Figure 15 p-s. Coleopteran pupa	88
Figure 16 a-c. Phloebius sp. A	89
Figure 17 a-c. Phloebius sp. B	89
Figure 18 a-g. ?Odioporus sp.	
Figure 19 a-f. Lixus sp. A	91
Figure 19 g-i. Lixus sp. A male	92
Figure 20 a-f. Lixus near germaini Hustache	93
Figure 20 g-1. Lixus near germaini Hustache male	94
Figure 21 a-c. Tanymecus near dilaticollis Gyllenhal	95
Figure 22 a-f. Tanymecus sp. A	96
Figure 22 g-j. Tanymecus sp. A male	
Figure 22 k-o. Tanymecus sp. A female	98
Figure 23 a-i. ?Tanymecus sp	
Figure 24 a-g. Unknown genus sp. A	100
Figure 24 h-m. Unknown genus sp. A male	101
Figure 24 n-v. Unknown genus sp. A female	102
Figure 24 w-z2. Unknown genus sp. A larva	
Figure 24 z3-z4. Unknown genus sp. A pupa	104
Figure 25 a–g. Unknown genus sp. B	105
Figure 25 h-p. Unknown genus sp. B abdominal venter, sternites, and male and	
female genitalia	106
Figure 26 a–i. Unknown genus sp. C	
Figure 27 a–e. Unknown genus sp. D	
Figure 28 a–j. Unknown genus sp. E	
Figure 29 a-i. Hypamazso pauli (Fairmaire) female and male	
Figure 29 j-m. Hypamazso pauli male genitalia, siphon, aedeagus and sternite IX	
Figure 29 n-s. Hypamazso pauli larva	.112
Figure 29 t-w. Hypamazso pauli pupa	
Figure 30 a-i. Hypamazso pauli adult wing patterns and form of elytral hairs	
Figure 31 a-i. Hypamazso sp. B adult	
Figure 31 j-o. Hypamazso sp. B larva	
Figure 32 a–g. Hypamazso sp. C adult	
Figure 32 h-k. Hypamazso sp. C larva	
Figure 33 a-d. Obeneopsis sp.	.118
Figure 33 e-l. Obeneopsis sp. female abdominal venter, sternites, and male and	110
female genitalia	119
Figure 33 m-t. Obeneopsis sp. larva	
Figure 33 u–y. Obeneopsis sp. pupa	121
Figure 34 a-d. Hyllisia near vittata Fåhraeus	
rigure 35 a-f. Stenalia near occidentalis Pic	
Figure 35 g-k. Stenalia near occidentalis larva	
Figure 35 I-q. Stenalia near occidentalis pupa	
GOUTE Sh 3_t Standing CD H	126

	Figure 37 a-g. Stenalia sp. C	126
1	Figure 38 a-e. Stenalia sp. D	127
	Figure 39 a-e. Stenalia sp. E	127
	Figure 40 a-i. Paramarygmus sp. larva	120
	Figure 40 a-1. Futumut ygmus sp. tatva	120
	Figure 41 a-g. Zophodes sp. larva	
	Figure 42 a-f. Stenolanguria caudata	
	Figure 42 g–j. Stenolanguria caudata larva	
	Figure 42 k-n. Stenolanguria caudata pupa	
	Figure 43 a-f. Stenolanguria sp.	133
	Figure 43 g-l. Stenolanguria sp. abdominal venter, hindwing, female genitalia,	
	spermatheca and right elytron	. 134
	Figure 44 a-h. Barbaropus near olseni Pic	135
	Figure 44 i-m. Barbaropus near olseni Pic larva	126
	Figure 45 a–h. Barbaropus sp. C.	
	rigure 45 a-n. <i>Durvuropus</i> sp. C	13/
	Figure 46 a-h. Barbaropus sp. B	138
	Figure 47 a-c. Barbaropus sp. D	139
	Figure 48 a-h. Barbaropus sp. E	140
	Figure 49 a-i. Promecolanguria rufocephala	141
	Figure 50 a. Dorsal view of a hypothetical fly	142
	Figure 50 b-d. Lateral view of a hypothetical fly	143
	Figure 51 a. Dorsal view of Diopsina sp.	
	Figure 51 b-d. Lateral view of Diopsina sp.	
	Figure 52 a-d. Diopsis near lindneri Feijen and Diopsis sp. B	145
	Figure 53 a-d. Mepachymerus sp. larva and pupa	146
	Figure 54 a-d. Elachiptereicus abessynicus Becker larva and pupa	146
	Figure 55 a-d. Pachylophus sp. larva and pupa	146
	Figure 56 a-b. Bistrispinaria fortis (Speiser) adult	147
	Figure 57 a-h. Atherigona soccata Rondani adult	147
	Figure 58 a-d. Dorsal view of a parasitic hymenopteran	
	Figure 58 e-f. Lateral view of a parasitic hymenopteran	149
	Figure 59 a-b. Dorsal view of Siphona sp. A	150
	Figure 59 c-e. Lateral view of Siphona sp. A	
	Figure 60 a-f. Siphona sp. B	152
	Figure 61 a-b. Dorsal view of Lydella near sesamiae Mesnil	
	Figure 61 c-g. Lateral view of Lydella near sesamiae	. 154
	Figure 62 a. Dorsal view of Descampsina sesamiae Mesnil	. 155
	Figure 62 b-e. Lateral view of Descampsina sesamiae	. 156
	Figure 63 a-g. Leskia sp.	
	Figure 64 a-c. Scelio sp.	
	Figure 65 a-c. Inostemma sp.	
	Figure 66 a. Dorsal view of Brachymeria kassalensis (Kirby)	
	Figure 66 b-c. Lateral view of Brachymeria kassalensis	
	Figure 67 a-d. Hockeria sp.	. 162
	Figure 68 a-j. Gonatocerus sp.	
	Figure 69 a-c. Tetrastichus sp.	. 164
	Figure 70 a-c. Pediobius homoeus (Waterston)	. 165
	Figure 71 a–c. Pediobius furvus (Gahan)	.166
	Figure 72 a-c. Sycophila sp.	167
	Figure 73 a-c. Eurytoma oryzivora Delvare	169
	Figure 74 a-c. Eurytoma sp. A	
	Figure 75 a-c. ?Eurytoma sp. C.	
	Figure 76 a-d, Eurytoma sp. D	
	Figure 77 a-c. Macroneura sp.	
	Figure 78 a-c. Norbanus sp.	
	Figure 79 a. Goniozus indicus Ashmead	
	Figure 80 a-c. Triaspis sp.	
	Figure 81 a–c. Cotesia flavipes Cameron	
	Figure 82 a-c. Cotesia sesamiae Cameron	
	Figure 83 a-c. Cotesia near sesamiae Cameron	
	Figure 84 a-d. Dolichogenidea polaszeki Walker	
	Figure 85 a-d. Dolichogenidea sp. C.	180

Figure 86 a-c. Dolichogenidea sp. A	181
Figure 87 a-c. Dolichogenidea sp. B	182
Figure 88 a-c. Phanerotoma sp.	183
Figure 89 a-b. Chelonus curvimaculatus Cameron	184
Figure 90 a-h. Rhaconotus sp. A	185
Figure 91 a-c. Rhaconotus sp. B	186
Figure 92 a-d. Rhaconotus sp. C	
Figure 93 a-h. Habrobracon sp. A	188
Figure 94 a-h. Habrobracon sp. B	189
Figure 95 a-d. Habrobracon sp. C	190
Figure 96 a-d. Bracon sp.	191
Figure 97 a-c. Stenobracon rufus Szepligèti	192
Figure 98 a-h. Stenobracon sp.	193
Figure 99 a. Syzeuctus sp.	194
Figure 100 a-d. Pristomerus bullis Fitton	
Figure 101 a-d. Pristomerus sp. A	
Figure 102 a-e. Pristomerus sp. B	197
Figure 103 a-e. Pristomerus sp. C	198
Figure 104 a-d. Dentichasmias busseolae Heinrich	199
Figure 105 a-d. Venturia sp. A	
Figure 106 a-c. Venturia near jordanae Fitton	201
Figure 107 a-d, Venturia sp. B	
Figure 108 a-d. Venturia sp. C	
Figure 109 a-d. Holcopimpla sp. A	204
Figure 110 a-d. Holcopimpla concolor (Brullé)	205
41)	

INTRODUCTION

The Global Environment Facility (GEF) with implementation support from the United Nations Environment Programme (UNEP) funded the project GFL/2711-01-4345 focusing on "Conservation of Gramineae and Associated Arthropods for Sustainable Agricultural Development in Africa". The goal of the project was to document the diversity of wild grasses (Gramineae) and associated insects in different agroecosystems and their adjacent natural habitats, and to exploit the relationships between certain grasses and insects to integrate self-regulatory and sustainable pest management strategies into traditional farming methods. The primary objective was to identify and implement conservation and management measures that would prevent the loss of biodiversity of wild grasses and their associated insects, and to conserve these rich genetic resources in and around agroecosystems in Ethiopia, Kenya and Mali.

A rich community and diverse taxonomic array of stemborers and their associated insects—parasitoids and hyperparasitoids—were reared during the project. The great diversity of stemborers and associated insects present on grasses presented two serious challenges: (1) successfully rearing stemborers from small grass stems and (2) identifying the stemborers and their associated parasitoids, at least to the genus level, depending on availability of species descriptions, keys and taxonomic experts. Experience showed that the small stems of wild grasses were not suitable for rearing field-collected larvae in the laboratory. Grass stems shrivelled easily, killing the young larvae inside. After considerable experimentation, a standard method for handling and rearing borer larvae was developed and has been made available with the publication of the handbook "How to Handle Grass Stemborer Larvae", available in a revised edition in French as "Comment mener à terme un élevage de foreurs?". These methods contributed to the successful rearing of stemborers and their parasitoids. The present work aims to provide an identification tool to enable field workers and researchers to identify stemborers and their associated parasitoids reared from grasses.

Composition of the Manual

The manual comprises keys to various insect taxa that bore in grasses or that parasitise the borers. Some hyperparasitoids (parasitisers of primary parasitoids) are also included. The keys are supplemented by figures of most of the species and, in the case of lepidopteran stemborers, photographic plates of adults and their genitalia. An examination of genitalia is the gold standard for identification of stem-boring moths. Adult wing patterns and colours are highly variable and scales easily rubbed. Characteristics of genitalia are highly conserved with minimal intraspecific variability. To our knowledge, no regional guide exists that includes an extensive photographic treatment of both adult moths and their genitalia. A glossary is also provided. All words that appear in the glossary are in bold in the main text.

Sixty-eight (68) species of stemborers are keyed and most of these are illustrated with line drawings and photograph plates (moths). An effort was made to provide illustrations of all developmental stages (except the egg) for each species, whenever stage-specific specimens were available. The borers were represented by the beetles (order Coleoptera) with 6 families, 14 genera and 33 species; the moths (order Lepidoptera) with 5 families, 13 genera, and 25 species and the flies (order Diptera) with 4 families, 7 genera and 10 species. Fifty-three (53) species of parasitic insects were reared from stemborers. These were of two orders, the wasps (Hymenoptera) with 11 families, 26 genera and 48 species, and the flies with 1 family, 4 genera and 5 species.

Many of the identifications are to the level of genus only. This was due to the absence of published taxonomic work on several groups of insects and because some of the insects we reared represent new species. In these cases, morphospecies are designated within genera.

Figures illustrating the general morphology of the four orders—Lepidoptera (Figures 1a–3c), Coleoptera (Figures 15 a–s), Diptera (Figures 50 a–d) and Hymenoptera (Figures 58 a–f) are referred to at the beginning of the treatment of each order. These illustrations should help the novice identify grass stemborers and associated parasitoids.

THE INSECT ORDERS REARED FROM WILD AND CULTIVATED GRASSES

Key to the insect orders associated with Gramineae in Kenya, based on adult characters

•	duit characters
1	Two pairs of membranous wings present (rarely wingless); mouthparts either with strong mandibles or modified into a coiled proboscis
1	One pair of membranous wings present, the other pair (forewing or hindwing) modified; mouthparts mandibulate (for chewing) or suctorial (for sponging or lapping)
2	Forewings and hindwings covered dorsally and ventrally with overlapping scales; mouthparts modified into a coiled proboscisLepidoptera (p. 3)
2	Wings without scales, hindwings with fewer veins and cells than forewings (rarely wingless); mouthparts with strongly developed mandibles for chewing
3	Forewings modified into hardened and rigid elytra, usually covering the entire abdomen and the membranous hindwings; mouthparts mandibulate
3	Forewings membranous, hindwings reduced to club-like halteres; mouthparts suctorial Diptera (p. 20, 24)
	ey to the insect orders associated with Gramineae in Kenya, based on arval characters
	52 5-91 (MICO-000-NO-0000)
1	Prolegs present on two or more segments of the abdomen, usually on abdominal segments III to VI and anal segment; crochets well developedLepidoptera
1'	Prolegs absent on abdominal segments III to VI; crochets absent
2	Head well developed and visible dorsally; thoracic legs present; ampullae and transverse plicae present dorsally
2′	Head hardly visible dorsally or totally absent; thoracic legs absent; ampullae and transverse plicae absent
3	Body commonly U-shaped, wider at midlength and bluntly rounded on both ends; mouthparts reduced to a pair of opposable sharply-pointed mandibles or sclerotised plates fused to the head segment
3′	Body maggot-like or vermiform, anterior end pointed, posterior end blunt; hook-like mouthparts visible
	ey to the insect orders associated with Gramineae in Kenya, based on upal characters
1	Appendages hidden (coarctate pupa) or, if visible, the appendages fused to the body wall and incapable of moving (obtect pupa)
1'	Appendages not adhering to the body wall, and capable of moving (exarate pupa) 3
2	Obtect type; appendages visible but fused to each other and to the body wall forming a cover plate; antennae, wing pads and galea visible; spiracles distinct on the mesothorax and some abdominal segments; antennae adjacent to the inner margins of wings; cremaster usually present; body elongate with pointed posterior end; genital and anal openings distinct
2′	Coarctate type; appendages not visible on the outside; caudal and thoracic spiracles present as scars; antennae not visible; cremaster absent; body barrel-shaped to elongate, often with paired spiracles posteriorly; genital and anal openings not visible
3.	Body constricted between thorax and gaster; ocellar triangle distinct; wing rudiments not elytra-like; legs against the lateral side of body; prothorax not concealing the head; legs with femur and tibia almost parallel to each other
3′	Body without marked constriction between thorax and gaster; ocellar triangle not visible; wing pads elytra-like; legs against the ventral side of body; prothorax concealing the head; legs strongly elbowed at posterior end of femur

THE STEMBORERS

LEPIDOPTERA: MORPHOLOGY OF A LEPIDOPTERAN STEMBORER (Figures 1 a-i, 2 a-d, 3 a-c)

11 4	48 6	C * *	- £ 1 1	A 4	-4	Language and	adult charac	4
KPV TO	The I	ramilies	ar ieni	aanteran	stemnorers.	nased or	ı adılır charac	Tere

ey to the tamilies of lepidopteran stemborers, based on adult chara Small to medium-sized moths (8.5-16 mm long) with slender bodies; head and thoracic dorsum without thick hair tuft; forewings straw-coloured to dull brown and pinkish-red; if dull-coloured, forewings bear brown dots or silvery markings; labial palps prominently Robust-bodied moths (11-26 mm long); head and thorax covered with thick hairtuft; forewings with variable colour pattern ranging from light beige with a longitudinal brown stripe, grey-brown with median and terminal blackish-grey bands, or whitish and mottled with light grey spots; labial palps short and hardly visible......4 Terminal margin of forewings obliquely truncate or rounded; forewing vein R, and R, not stalked, M, and M, approximated at base; hindwing vein CuP present; basal lobe and basal fringes of long hairs absent; head with or without a corneous point; labial palp long and snout-like, if short, terminal segment pointed downwards; ocelli and chaetosemata present; tympanal organ usually present (absent in Tortricidae); hind tibiae without long hairy scales ______3 Terminal margin of forewings acute or pointed; forewing vein R, and R, stalked, M, and M, wide apart; hindwing vein CuP absent or poorly developed, basal lobe distinct and with long fringe of hairs; head rounded, without a corneous point; labial palp upcurved with a short terminal segment; ocelli and chaetosemata absent; tympanal organ wanting; hind Labial palps porrect and beak-like; forewings vary from straw-coloured with brown dots and silvery markings, to light beige, to pinkish red; costa not strongly arched, vein CuP absent, R, and R, stalked, chorda absent; hindwing vein Sc + R, fused with Rs beyond discal cell, CuP complete, Rs and M, stalked near the discal cell; tympanal organ present at base of abdomen Pvralidae (p. 9) Labial palps ascendingly recurved with the small terminal segment pointed downward; forewings light brown mottled with yellow brown to dark brown scales, costa strongly arched; vein CuP present near the wing margin, R3 and R4 separated, chorda present; hindwing vein Sc + R, separated from Rs beyond discal cell, CuP incomplete not reaching margin, Rs and M, close to each other basally; tympanal organ absent from base of abdomen Tortricidae (p. 11) Tympanal organ located in the metathorax; forewing vein M not forked inside the discal cell, areole present, chorda absent; hindwing vein Sc + R, shortly fused with Rs beyond Tympanal organ absent; forewing vein M forked forming a small cell inside the discal cell, areole absent, chorda present; hindwing vein Sc + R₁ separated from Rs beyond discal cell, Key to the families of lepidopteran stemborers, based on larval characters 1' Head semiprognathous; primary setae usually on strongly pigmented pinacula; crochets on prolegs arranged in a complete circle or penellipse; anal shield on A10 indistinct; body Head hypognathous to semihypognathous; primary setae not on strongly pigmented pinacula; crochets on prolegs uniordinal, arranged in a linear pattern; anal shield on A10

Prothoracic shield (T1) humped and rugose with a posterior band of spicules or small peg-

FAMILY COSSIDAE

General features: Cossids are medium-sized to large-bodied moths commonly found in the tropics and subtropics. In general, adults have elongated and slender forewings much shorter than the hindwings, and often mottled with brown, grey or cream-white shades. Head with elevated and slender scales lining the vertex and a mass of converging scales along the frons. Chaetosemata and ocelli absent, proboscis very short and naked to reduced, labial palp moderately short and upturned, epiphysis often present. Antennae bipectinate in male and usually filiform in female. Forewings elongate with an areole, pterostigma absent, retinaculum on Sc or Sc and costa, strongly forked M cell, chorda present, R₃ stalked with R₄ to termen and CuP present. Hindwings broad and rounded, vein M forked, CuP reduced and 2A veins present.

Male genitalia: Uncus usually broad and well developed, gnathi linked by the scobinate and bilobed bulla but may be weak, separate, long and slender with sclerotised bands, or totally absent as in *Phragmataecia*; valves with a basal spine and fused with the juxta, a scobinate structure associated with juxta basally and sacculus ventrally, aedeagus may be a simple rod or broad, short, and apically cleft into dorsal and ventral longitudinally corrugate 'jaws'.

Female genitalia: Elongate and slender ovipositor lobes, segment 8 and the apodemes distinct.

Larva: Commonly wood and stemborers with broad head longer than wide and with large mandibles. Prothorax with a distinct shield that is very rough centrally and very smooth along the caudal margin. Pinacula present, heavily sclerotised, setae very small to reduced. Prolegs present with crochets in complete circle.

Pupa: Long, cylindrical, distinctly sclerotised. Head with spines. Abdominal segments III–VII in males and III–VI in females movable. Segments II–VII of males and II–VI in females each with two transverse rows of spines, segments VII–VIII in female and VIII in male with one row only. Cremaster absent. During moulting, pupae are extruded from tunnel.

Biological information: The larva of Phragmataecia boisduvalii Herrich-Shäffer was dissected only from the stems of Echinochloa pyramidalis (Lam.) Hitchc. & Chase and Phragmites karka (Retz.) Steud.

Phragmataecia boisduvalii Herrich-Shäffer (Figures 14 a-l)

Diagnosis: Adult robust with thick tuft of hairs on the thorax. Forewings white, heavily mottled with grey spots towards the apical half, basal half generally greyish. Head light brown with frontal tufts brown to pale reddish-brown. Basal 2/3 of antenna bipectinate, distal segments serrate except simple tip. Pectinate segments around 25 with middle segments having longest arms. Labial palpi short and projected forward. Hindwings creamy white to light yellow-brown without marks. Underside of both wings (fore and hind) striated, striae on forewings only visible in the upper side. Frenulum a single stout spine. Retinaculum distinct and well developed. Legs, except tarsi, light brown and fringed with hairy scales. Venation of forewing shows a long areole with nearly a third of its length outside the cell angle. R₁ emanates from the basal one-third of the areole, R₂ from close to tip of areole, R₃ shortly stalked and anastomosed with base of vein R₄ + R₅. M₁ comes well below the upper angle of cell, large median cell with upper and lower branches terminating between veins M₁-M₂ and the distinctly basally separated M₂-M₃. Hindwing with M₁ far below Rs, large median cell with its branches ending nearly between M₁-M₂ and Cu_{1A}-M₃. Veins M₂ and M₃ widely separated.

Male genitalia with highly sclerotised uncus with bluntly rounded distal end, subtriangular tegumen, vinculum relatively wide medially, saccus short, juxta subglobose without clear median lobes but lateral lobes slender, aedeagus developed with no distinct

base and relatively large vesica thickened on one side only.

Mature larva 25–31 mm long and 5–6.5 mm wide. Body cylindrical, robust, dull white. Prothoracic shield broad with rough central area bordered with yellow markings. Head wider than long. Coronal suture pale to absent. Frons widened at midhalf. Epicranial notch absent. Adfrontals distinctly broad towards the mouth. Prothoracic spiracle large and oblongate, about twice the size of abdominal segment I–VII spiracles. On thorax, subdorsal seta 2 (SD2) almost caudad to subdorsal seta 1 (SD1) on the prothorax (T1); lateral (L) group trisetose, present in all thoracic segments, lateral seta 1 (L1) closer to L3 than to L2 in the mesothorax (T2) and metathorax (T3). On the abdomen, SD1 large and above spiracle on abdominal (A) segments A1–A7, SD2 very small, located anterior of spiracle on A1–A7. Spiracles of segment VIII ovoid, visible dorsally. Caudal end cleft medially. Last abdominal segment (A10) shield with four setae on each side. Prolegs on A3–A6 and A10 short.

Pupa dark reddish-brown, not more than 31 mm long, cylindrical with tapering head and short wing pads. Abdominal segments II–VIII with transverse row of spines, segment III–VIII each with a long row of anterior spines reaching and passing in front of the spiracles and a short row posteriorly. Segments IX–X with strong spines dorsally, laterally and ventrally.

Biological information: P. boisduvalii has been reported in Kenya to breed on tall grasses. The pupa is host to a pteromalid wasp, Norbanus sp. (Hymenoptera: Pteromalidae) that parasitises grass stemborers (e.g. Chilo partellus) that attack maize. Rearing records showed that second to third instar larvae took four months to pupate using moderately mature 8.75 cm long maize stalks. Adult emerged from the pupa in 42 days.

FAMILY GELECHIDAE

General features: Small moths with relatively broad wings; antennae simple, scape rarely with an anterior comb of stiff hairs or pecten; head smooth with scales closely pressed to the body; proboscis (tongue) thickly covered with hairs basally; labial palps slender and stout, recurved or upturned, segment II tufted underneath, segment III slender and pointed; maxillary palps usually 4-segmented; ocelli present or absent; chaetosemata absent; epiphysis present; tibia III bears long hairlike scales; forewing lacking pterostigma, vein R4 and R5 stalked, R5 to costa, CuP absent; retinaculum of adult females a row of curved specialised scales on vein R; hindwing usually trapezoidal with distinct apex, termen sinuate or emarginate, veins R5 and M1 approximate at base or stalked, CuA sometimes with a basal pecten, CuP usually absent; abdomen rarely with spines dorsally; male genitalia with a broad or slender uncus; aedeagus bulblike, strong and loosely connected to a membrane, cornuti often present; valves rodlike, long and slender; gnathos well developed, hooklike or elongate and pointed; sternite VIII divided into distal lobes covering the genitalia in some cases; female genitalia with variable ostium ranging from membranous and ill-defined to strongly sclerotised, or with elaborate foamlike structure and with projecting ostial plate; bursa copulatrix with 1–2 signum/signa of various designs.

The larva usually lacks secondary setae on prolegs; sometimes with an anal fork; crochets

vary from uni- to biordinal, arranged in a circle or mesal penellipse.

Pupae with labial palps and fore femora hidden; cremaster sometimes present; maxillary palps always present.

Biological information: Larvae have different feeding habits, mining or boring in leaves, stems, seeds, fruits and tubers. Generally, larvae feed in concealed habitats (e.g. rolled leaves or within shoots). Pupae in silken threads or cocoon in or near larval shelter; not protruded on eclosion.

Stegasta sp.

(Figures 4 a-g [adult]; 4 h-l [pupa])

Diagnosis: Small brownish-yellow moth, 15 mm long, forewing span 30 mm. Forewings sparsely mottled with dark brown scales, basally uniformly chocolate brown, apex pointed and costal margin moderately convex. Hindwings clothed costally and basally with very

long tuft of grey hairs almost reaching the wing tip. Labial palpi chocolate brown except basal segment covered with white scales, upcurved, pointed tip well beyond the vertex and twice as long as diameter of eye. Head rounded, ventral margins lined with long cream-white hairs extending to midlength of eyes. Antenna simple, scape without long hairs. Legs I and II covered with brown scales and tibia III with white scales. Abdomen dull brown.

Full grown larva about 17 mm long, yellowish-white with brownish median area of head, blackish-brown to reddish-brown mandibles, pale brown marks on the meso- and metanotum and sides of the body. Legs yellowish-brown, prolegs with circular crochets.

Pupa about 18 mm long, yellowish-brown with a roughened head. Venter of segments V and VI with a pair of scale-like tubercles, each tubercle lined with three short white hairs in a transverse row and a very long white hair anterolaterally. Posterior end with a pair of dark reddish-brown tooth-like projections. Cremaster downcurved and projected anteriorly toward segment VIII.

Biological information: Stegasta sp. is the only gelechiid we reared. It was reared from Sorghum bicolor (L.). When reared on maize stems the larval period was 16 days (L3-L5) and pupation lasted for 8 days.

FAMILY NOCTUIDAE

General features: Small to large heavily-bodied, dull-coloured moths easily distinguished from other moths by the postspiracular location of the counter-tympanal hood and the presence of a thick tuft of scales on the thoracic dorsum. Ocelli present, antennae usually filiform but may be dentate or pectinate, labial palps porrect or curved upwards, maxillary palps with a single segment, epiphysis present, tibial spurs 0-2-4, spines sometimes present on the tibiae and tarsi. Forewings fairly narrow, hindwings distinctly broadened. Areole present in forewing, vein M, much closer to M3 than to M1, cubitus 4-branched, 1A+2A forming a basal branch. Hindwing with Sc + R, separated at base but fused shortly with Rs near base of discal cell, vein M, present or absent. If present it is weak and borne closer to M, than to M, (trifine group) or strong, arising closer to M_1 than to M_1 (quadrifine group).

Noctuids represent a very diverse group with many subfamilies. Only the trifine subfamily Amphipyrinae will be dealt with, as the grass stemborers—Sesamia Guenee, Busseola Thurau and Manga Bowden—belong to this group. Members of this subfamily have stout adults, thorax crested with hairlike scale coverings or uncrested and clothed with hairs only, trifine hindwing venation without vein M2, hairy and lashed eyes, and spines on tibia.

Male genitalia with valve entire, sacculus separate from cucullus, manica spinose to membranous and cornutus bulbed or unbulbed. Valvae usually with a clavus (a structure on the dorsal margin of the sacculus). Base of abdomen provided with paired hair-pencils.

Female genitalia with or without signa in the bursa copulatrix. Ostial segment with or

without a membranous pad before the ostium.

Larvae of most species usually with only primary setae but others may show well developed secondary setae present on verrucae. Head rarely with secondary setae. Crochets uniordinal, well developed on **prolegs** of abdominal segments III–VI and X.

Pupae with smooth to rough head, prominently circular spiracles and punctures along the dorsal anterior portion or nearly the entire area of abdominal segments II-VIII. Segment VI-VII with a pair of round swellings in some species. Cremaster present.

Biological information: Noctuids are generally nocturnal moths. Larvae demonstrate diverse feeding habits represented by fruit-piercers, defoliators and stem cutters, stem and fruit borers. A few species are predactious. Stem-boring females lay eggs in groups of 30-150 on the inner surface of the leaf sheaths. The incubation period is 5-7 days. Newly hatched larvae live aggregately between the stem and the leaf sheath and disperse only after about a day. Young leaves in the whorl provide food, shelter and protection to the dispersing larvae from predators and parasitoids.

Key to the adults of noctuid stemborers

- Forewings light beige; thorax clothed with hairs only; hairlike scales absent

2	Antennal segments serrate or bipectinate (males)
2′	Antennal segments simple (females)
3	Antennal segments not pectinate, weakly serrate, saccular process of male genitalia lacking spines, costal spine absent
3′	Antennal segments pectinate or bipectinate, saccular process always spinose, costal spine present
4	Antennal segments strongly bipectinate, pectinations mostly longer than width of antennal shaft
4′	Pectinations of antennal segments never longer than width of antennal shaft
5	Juxta of male genitalia as wide or nearly as wide as long. Spine on costal process variable, sometimes tapering, more often nearly truncate at apex, usually with small tooth at one end, saccular process robustly spined
5′	Juxta of male genitalia much longer than wide, costal spine weakly and asymmetrically
1	bifid, saccular process not robustly spined
6	Costal spine clearly bifid, strong and more or less symmetrical
6'	Costal spine spatulate, sharp, with irregular, fine teeth
7	Base of the ductus bursae strongly sclerotised, either in longitudinal strips or with the sclerotised area cut off obliquely and appearing as a dark triangle
7′	Base of the ductus bursae sclerotised and appearing invaginated; sclerotisation not extending into the ductus
8	Sclerotisation at base of ductus bursae cut off obliquely, appearing as a dark triangle, sclerotised lateral lobes of lamella vaginalis absent
8′	Sclerotisation appearing as longitudinal strips, extending into ductus, sclerotised lateral lobes of lamella vaginalis present
9	Antennae bipectinate (males)
9'	Antennae more or less simple, not bipectinate (females and some males)
10	Antennal pectinations very short, appearing nearly serrate, juxta with a pronounced, ventral finger-like process, aedeagus with two robust blade-like cornuti
10'	Antennal pectinations obvious, juxta without ventral process, cornuti not as described11
11	Aedeagus with paired lateral dentate processes and terminal cornutus, clasper of valve large and heavily sclerotised Busscola fusca (Fuller) (Plates 19, 28 and 29; Figures 13 a-e)
11'	Aedeagus with paired lateral dentate processes but lacking terminal cornutus, clasper of valve small and lobelike
12	Males
12′	Females15
13	Sacculus with heavily toothed and well-developed clasper, aedeagus with a sclerotised band ending, when everted, in a variable number of small spines (usually three or four) Manga nubifera (Hampson) (Plates 21, 22 and 27; Figures 12 a-f)
13′	Sacculus lacking clasper, aedeagus not as above
14	Aedeagus with heavily sclerotised, long, slightly curved and pointed cornutus, about one-half length of aedeagus

14	'Aedeagus with cornuti confined to one end, one a group of longish recurved spines and the other more or less wheel-like with a serrate margin
15	Sclerotisation of ductus uniform and not patterned
15	Sclerotisation of ductus with a distinctly beaded pattern
16	Corpus bursae with two obvious signa, ductus inserted obliquely between sclerotised latera lobes of lamella vaginalis
16	Corpus bursae with one obvious signum (the dorsal one very faint, if present); sclerotised lateral lobes absent
17	Sclerotisation of ductus starting at base and continuing asymmetrically into the ductus ending obliquely; the two signa ellipsoid, each with one or two ridges
17'	Sclerotisation of ductus weak or absent from bottom 1/3 of ductus, extensive and prominent in middle, often appearing expanded and sac- or appendix-like; signa variable, usually two sometimes only one apparent signum; the signa oblong or ellipsoid, each usually with a more or less central dark spot, lacking ridges
	Busseola obliquifascia (Hampson) (Plates 20 and 28)
Ke	y to the larvae of common noctuid stemborers
1	Larva with a brown head, brownish-yellow body with pale pink dorsal markings; elevated chitinised plate with setae broad and relatively darker; angle between dorsal setae (D2 and D1) and subdorsal seta (SD1) always greater than 140° angle; prothoracic shield pale yellow brown; caudal plate yellow brown; body about 30 mm long and 3.5 mm wide
1'	Larva with dark brown head, body usually buff to purple brown with elevated chitinised plate bearing setae narrow and pale yellow brown; angle between dorsal setae (D2 and D1) and subdorsal seta (SD1) narrow, often less than 120" angle; prothoracic shield lighter than head; caudal plate same colour as the prothoracic shield
K.	y to the pupae of common noctuid stemborers
1	Anterior region (head) with rough stony-like pattern; cremaster indistinctly separated from the last abdominal segment
1′	
2	Cremaster with four short but robust spines, two inner and two outer all slightly projected posteriorly to upcurved; midanteroventral part of the head with rough stony appearance, dorsal portion smooth; abdominal segments V–VI each with a pair of minute swellings Sesamia calamistis Hampson (Figure 11 n–q)
2'	Without the above combinations of characters
3	Cremaster with four spines, two strong dorsolaterals and two small ventrals; anteroventral tip of head entirely rough; abdominal segments V–VI with pronounced pair of swellings, connected to each other by concave punctures
3'	Cremaster reduced to a heavily sclerotised dark reddish-brown process without spines; entire head region dorsally and laterally rough and stony; abdominal segments VI–VII roughly punctate anteriorly and finely punctate posteriorly, VIII roughly punctate anteriorly
4	Cremaster with two short strongly diverging spines arising from a common base
4′	Cremaster with two short diverging spines widely separated basally and arising from a half-rounded plate

FAMILY PYRALIDAE

General features: Small to moderately-sized moths representing some of the most economically important species associated with agricultural crops. Majority dull-coloured and yellowishbrown. Pyralids are known as snout moths due to the long, porrect and beaklike labial palpi with three segments. Head bears smooth scales, some species have anterior tuft. Proboscis thickly covered with scales at base, but may be reduced or vestigial. Ocelli and chaetosemata present or absent. Compound eyes large and distinct, or reduced. Maxillary palpi usually 4segmented, some species with two or three segments only but always scaled. Antennae variable, from simple and filiform in both sexes, to slightly thickened and serrate or bipectinate in males. Epiphysis present, spurs usually 0-2-4 and rarely 0-2-2. Wings variable in venation and provide useful characters in species identification. Forewing elongate or triangular with cubitus showing four branches, Cu, greatly reduced to a tubular vein in terminal part of the wing or entirely absent, chorda absent and vein M stem in cell with R, and R, stalked or running together, M, approximated to M₃ at base, 1A + 2A commonly with a large fork, CuP usually absent. Hindwing usually broad, with 1-3 frenular bristles in female, vein Sc + R_1 always free from base to a little beyond angle, close to or shortly fused with Rs beyond discal cell, M2 approximated to M3 basally, CuP often present, large anal area with two anal veins, CuA with basal pecten in some species, coremata often present, and tympanal organ located at base of abdomen. Abdomen with tympanic bullae located in the ventral part of the basal segment close to the midline. When present, a longitudinally-oriented median chitinous plate (praecinctorum) hangs in the cavity between the abdomen and thorax in front of the bullae. The plate is sometimes absent. When present, it may be well developed, transversely flattened and prominently bilobed, or poorly divided distally, reduced to a lobe, or simply composed of distal tufts of scales.

Male genitalia with a strong, hooklike gnathos, simple to complex valve, well developed anellus, and a prominent juxta. The simple tegumen, which may be broad or narrow, connects laterally to the U- or V-shaped vinculum. Saccus poorly developed or absent. Coremata usually

visible on 7th-8th abdominal segments and strongly developed in some species.

Female genitalia: Ovipositor shape variable, ranging from simple and short to long and drawn out with blade-like or serrate lobes. Ostium variable, from simple and membranous to sclerotised and very complex; signum/signa present, either single, double or multiple, varying from simple and thornlike, star-shaped, serrate, platelike, scobinate, to a group of spines or complex bands.

Biological information: Larvae exhibit a variety of behaviours. Many species feed while concealed (shoot, stem and fruit borers) while others live under folded or rolled leaves. The pupae are either naked or covered with silken threads and may be found inside the stem or concealed inside rolled leaves. In this manual, the focus is only on the stemborers reared from grasses; those belonging to the subfamilies Crambinae, Galleriinae, Peoriinae and Phycitinae.

Key to the adults of pyralid stemborers

2′	Forewings yellow-brown to reddish-pink, without metallic scales, apex rounded, costal band distinct; ocelli small to vestigial; labial palp 2–3x as long as the diameter of eye 6
3	Forewings without metallic scales
3'	Forewings with metallic scales 5
4	Male genitalia with juxta plate symmetrical with large central part, lateral arms truncate apically and not extending past costa, costa with pronounced submedian projection, aedeagus with a strong basal projection, ventral arm present. Female genitalia with ostial pouch heavily sclerotised, delicately wrinkled throughout, and with a pronounced invaginated notch, signum present, lamellate, with a median ridge
4'	Male unknown. Female genitalia with ostial pouch wrinkled at opening, heavily sclerotised, without invaginated notch, signum absent
5	Male genitalia with valve broad and moderately acute to rounded at apex; vinculum robustly U- or V-shaped; left arm of juxta plate shorter than right arm, or the arms subequal in length, apices of both arms broad and with bristles, aedeagus without a subapical digitate process. Female genitalia with seventh sternum not heavily sclerotised, subtriangular and thickly lined with minute spikes; two semi-triangular spikes on each side of the slightly sunken or enclosed ostial pouch; ductus bursae long and slender, gradually widening towards the bursa copulatrix; genital opening small; single long, scobinate signum
5'	Male genitalia with valve moderately narrow forming a very distinct bluntly rounded apex; vinculum slender and narrowly U-shaped; left arm of juxta plate spine-like apically, much shorter than the bent right arm; aedeagus with a distinct subapical digitate process; female genitalia with seventh sternum heavily sclerotised, subquadrate and lined with variably developed spikes on the entire plate, ostial pouch deeply enclosed; ductus bursae long and slender, parallel-sided, and without distinct widening towards the bursa copulatrix; genital opening large; single long, scobinate signum
6	Forewings pinkish, copper-red, or with red areas, without black spots7
6'	Forewings pale to dull yellow-brown, usually with black spots or bands, occasionally with extensive diffuse black area
7	Upper surface of forewing more or less uniformly copper-red or pinkish-red, sometimes with lighter areas posteriorly, but never with a pale costal band; aedeagus with two cornuti, one large, opaque and dagger-like, the other short, subelliptical, and with a serrate outer margin; posterior half of female ovipositor unsclerotised, pad-like, uniformly bristly
7'	Upper surface of forewing with definite pale costal band, darker band posterior to it, and rest of surface reddish; cornutus of aedeagus not dagger-like, but covered with denticles
8	Medial process of gnathos of male genitalia distinctly tongue-like, wing without extensive areas of black scales
8'	Medial process of gnathos not tongue-like, wing with extensive areas of black scales 10
9	Costal spine of male genitalia attached broadly to valve at its midlength; apex of valve acute; aedeagus distinctly swollen towards one end. Female genitalia without pronounced sclerotisation of the ductus bursae
9'	Costal spine of male genitalia weakly attached to valve near its base; apex of valve blunt, aedeagus not distinctly swollen at one end, nearly parallel sided, oblong. Female genitalia with ductus bursae strongly sclerotised along its entire length
10	Forewing with dark bands alternating with light bands, pattern evident; male genitalia with large centrally-located transtilla appearing as mirror image 'J'-shaped structures
10'	Forewing with black scales diffuse, darker at base, without recognisable pattern; saccular process of male genitalia ending in lobe bearing two hook-like processes

Key to the larvae of common pyralid stemborers

- 1' Crochets usually arranged in an oblong pattern, sometimes circular, complete and biordinal; body segments often without brown sclerotised platesPeoriinae
- 2 Lateral sclerotised plate located in front of prothoracic spiracle; body greyish-black with golden yellow hairs; each abdominal segment with M-shaped dotted marks, head blackishbrown; prothoracic shield dark brown Eldana saccharina (Walker) (Figure 5 h-n)

Key to the pupae of common pyralid stemborers

FAMILY TORTRICIDAE

General features: Adults are small moths with broadly rectangular forewings, usually brown, green or grey in ground colour. A number of species live on agricultural and horticultural crops but are common also in forest habitats. While some species are pests of cultivated crops, others are beneficial providing biological control of certain grasses viz. genus Bactra. Adults with ocelli and chaetosemata, proboscis without scales, labial palps beaklike and usually porrect, maxillary palp 1–4 segmented, 0-2-4 tibial spurs, antenna filiform. In males, forewings often with a costal fold and retinaculum on vein Sc, pterostigma usually absent, chorda present and with vein M stem sometimes present in cell, CuA with cubital pecten basally, CuP present distally, 1A + 2A on a distinct fork. Female with three frenula, abdomen without dorsal spines.

Male genitalia: Aedeagus generally short and curved, loosely connected to a membrane or attached to a sclerotised juxta, vesica with internal spinelike cornuti or barbs, vinculum V- or U-shaped with poorly developed saccus, valves simple with sacculus and costa distinct and cucullus well bounded, gnathos prominently developed to weak and vestigial, uncus shape varying from long and bonnetlike or narrow to weakly or deeply notched.

Female genitalia: Signum shape varies from two hornlike structures, two triangular hollow plates, single or double circle of rasplike projections, or pocketlike to prominent dentate patches masking large area of the bursa.

The larvae are common leaf feeders, rolling and joining the leaves and shoots but some may mine and tunnel through stems, fruits and inflorescences. They have a distinct **prothoracic** shield with sclerotised pinacula, crochets circular in uni-, bi- or triordinal pattern, and the anal fork with straight prongs above the anus. Tortricid larvae are easily recognised by the presence of a lateral (L) group of trisetose setae on the prothorax, and by having L1 and L2 setae adjacent on abdominal segments I–VIII and usually borne on the same pinaculum.

The pupa remains in the larval chamber and protrudes only during moulting. Head with spines, posterior end with cremaster and hooked setae, abdomen with two rows of dorsal spines arranged transversely on segments III–VII.

The grass stemborers reared from grasses belong to the subfamily Lethreutinae. They are characterised by the presence of one annulus (a ringlike segment) on the antennal flagellum, strigulae along the costa of the forewing and aedeagus united with the anellus and juxta. Two genera, Bactra and Thaumatotibia, have been recorded from grasses in Kenya. However, only the latter genus was found during the study. Unlike Thaumatotibia, the genus Bactra represents a group of true grass stemborers.

Thaumatotibia leucotreta (Meyrick) (Figure 9 a–f)

Diagnosis: Brown-coloured with dark reddish-brown concave band in the middle of the forewing touching the costa. Margin of costa with alternating rows of creamy-white and reddish-brown bands. The head is covered with dark brown to brown scales projected towards the curvature of the eyes. Ocelli distinct and small, chaetosemata present. Labial palps 1.5x as long as diameter of eye, laterally covered with white scales except for brownish, downcurved apical segment. Crown with a converging set of creamy-white and reddish-brown, small paddlelike scales with forked tips. Patagium covered with chocolate-brown appressed scales. Tegula thickly covered with broad paddlelike creamy-white, and black and reddish-brown scales provided with 2–7 serrations. At resting position the two broad bands are connected to each other by a thin transverse stripe anteriorly, thoracic and dorsal side of the forewings with puffed out hairlike scales. Legs with chocolate-brown, slender and needlelike scales on tibia III, inner tibial spur of leg III very long, almost as long as basal segment of tarsus III.

Larva pink to pale yellow with a reddish tinge. Mature larva about 15 mm long and 2.5 mm wide. Light brown head wider than long. Ocelli 3 and 4 very close to each other. Body tubercles often unsclerotised or with pale pigmentation, spines with sharp and needlelike tips. Spiracles circular. Anal comb distinct. Tubercle of ventral seta not touching coxa on II and III.

Pupa yellow-brown to reddish-brown, about 8–10 mm long and 2–2.5 mm wide. **Ventral** part of **face** without depression or cavities, front distinctly elevated. Abdominal segment II rough, strongly punctured **dorsal**ly, with two rows of spines. Spines on segments VIII and IX prominently developed. Segment X with spines and two pairs of hooked **setae**. Leg III very close to but not touching wing tips.

Biological information: This species is a common generalist pest of commercial and wild fruits. It is well recognised as a pest of fruits and cotton bolls in Africa (Schmutterer, 1969). In a recent study in Kenya it was reared from fruits of 79 host species, representing 33 plant families (RSC, unpubl. data). Not technically a stemborer, *T. leucotreta* is included here because during our survey of the wild and cultivated grasses of Kenya, it was found attacking maize cobs.

COLEOPTERA: MORPHOLOGY OF A COLEOPTERAN GRASS STEMBORER (Figure 15 a-s)

Key to the families of coleopteran stemborers, based on adult characters

FAMILY ANTHRIBIDAE

General features: Anthribids are minute to small (0.4–12 mm long) beetles with broad noses. They vary in shape from moderately elongate to ovoid and from slightly to strongly convex. Body usually clothed with white decumbent scales instead of setae, often with an obvious pattern. Beak very short to relatively long but stout and flattened. Labrum distinct and separated from rostrum, the maxillary palpi four-segmented. The 11-segmented antennae are straight and, in males, prominently long, resembling those of the Cerambycidae. Club indistinct to 3-segmented. Mandibles strongly developed and relatively acute. Pronotum with a basal or prebasal transverse ridge. Elytra often striate. Legs without apical tibial spurs, prothoracic coxae clearly separated and tarsi pseudotetramerous with fourth segment hidden. Abdomen with visible sternites I–IV usually connate.

Larva glossy white, C-shaped, typically resembling the curculionids. Diagnostic characters are the two-segmented thoracic legs lacking a terminal clawlike segment or **tubercle**, head distinctly separated from prothorax, **epicranial stem** long, frontoclypeal **suture** prominent showing distinct **clypeus**. **Ocelli** absent. Antenna large, conical with one segment. Mandibles with three teeth consisting of two marginals and a sharper **apical** tooth. Abdominal segments I–VII each with two **dorsal transverse plicae**, segment X without ovoid lobes separated by a **longitudinal** furrow. **Spiracles** commonly **annular** with thoracic **spiracles** on mesothorax or between the prothorax and mesothorax.

Pupa exarate with the legs held transversely parallel to each other, body oblongate with rounded head and tapering posterior end. Head prominent with large black eyes and short white hairs. Abdomen without prominent marking, lateral margins with short white hairs.

Biological information: Only one genus of anthribid was collected and successfully reared from grasses in Kenya. The two species are tentatively placed in the genus *Phloebius*. Previous to this the genus *Sintor* was reported to bore, feed and live in stems.

Key to the adults of anthribid stemborers

FAMILY CERAMBYCIDAE

General features: Adults with cylindrical, elongate and relatively flattened bodies with extremely long, usually 11-segmented antennae, at least half as long as the body and often much longer. Because of the noticeably long antennae, these insects are called longhorn beetles. Antennae are inserted on prominences facilitating movement of antennae towards the rear on top of the body. Eyes commonly emarginated or deeply notched on the inner margins, the antennae arising in front of or within the clefts. Prothorax without lateral carina, but often with lateral spines, and mesonotum sometimes with stridulatory file. All legs with tibial spurs, the 5-segmented tarsi appearing 4-segmented with the bilobed third segment concealing the small fourth segment. Tarsal claws simple.

Larvae commonly bore in stems or branches of trees and grasses. Many species are destructive to fruit trees, ornamental plants, bamboos and forest trees. Larval body whitish, long and cylindrical, without legs, mildly sclerotised with fine setae or small spines. Head deeply grooved, retracted into the enlarged prothorax, possessing large and robust mandibles and short antennae. Legs small, wide apart, absent in some species. Abdomen with fleshy ampullae, blisterlike structures present on the first six or seven abdominal segments, that aid in larval movement

Pupa typically exarate, body elongate with a narrow to wide cleft on the head formed by the bases of antennae lining the anterolateral one-half of the body. Head hypognathous, varying in shape from short and subtriangular to oblongate. Mandibles distinctly developed. Dorsum of abdomen with spines or oblong, blister-like structures.

Key to the adults of cerambycid stemborers

1	Body flat with antenna a little shorter than the body; eyes small, located close to the midlength of the head; head longer than wide2
1,	Body cylindrical with antenna prominently longer than the body; eyes large, located in the anterolateral corner of the head; head as long as wide4
2	Pronotum with three white longitudinal bands or ridges; tips of elytral striae 2, 3 and 4 separated distally, not reaching wing tip, striae 2 and 4 not Y-shaped
	(Note: This species shows considerable variation in shape of the elytra as well as the pattern, number of striae, and form of hairs present on the elytra [Figure 30 a-i]. It may represent a species complex).
2′	Pronotum with only the median longitudinal ridge distinct, sublateral white bands indistinct; tips of elytral striae 2 and 4 merged subdistally leaving stria 3 unconnected and forming a Y-shaped pattern, base of the Y merged with stria 6 before the wing tip
3	Scape without a ventral tubercle or swelling
3'	Scape with a sub-basoventral tubercle or swelling
4	Pronotum and elytra uniformly yellow; last abdominal sternite with a cleft in the posterior margin
4'	Pronotum and elytra with alternating grey and cream white longitudinal stripes; posterior margin of the last abdominal sternite slightly elevated medially
Ke	ey to the larvae and pupae of cerambycid stemborers
1	Larva
1'	Pupa
2	Ampulla or blisterlike structure on abdominal tergite VIII entire and lip-like, similar to those in IV–VII; epicranial stem long and entire; anal posterior margin strongly rounded
2'	Ampulla on tergite VIII broken in the middle; epicranial stem present or absent; anal posterior margin differently shaped
3	Prothoracic shield white and slightly sclerotised; ampulla on tergite VIII lip-like, with constriction in the middle; head relatively wider below midlength

FAMILY CURCULIONIDAE

General features: Adults with head distinctly produced in front of the eyes producing an elongate rostrum or snout that houses the folded, geniculate antennae. Antennal scape very long. Club compact. Body compact, usually heavily sclerotised, robust and strongly convex, lined with scales or bristles. Mouthparts modified, lacking labrum, maxilla reduced to short but robust palps, and gular sutures fused. Legs with five-segmented tarsi. Penultimate segment very small and hidden at the base of lobed third segment. Trochanters small and triangular, femora clavate or dilated towards apical one-half or grooved to house the tibiae. Abdomen with five visible sternites.

Larvae usually C-shaped, white. Body slightly sclerotised with brown patches. Head heavily sclerotised, hypognathous. Epicranial suture long. Clypeus distinctly developed, labrum with rounded, sinuous or emarginate anterior margin, mandibles robust. Single-segmented antennae minute, membranous with pronounced conical or plate-like sensory structure. Maxillary palpi two-segmented, labial palpi with a single segment. Larvae without thoracic legs but sometimes bearing ventrolateral swellings called pedal lobes with stiff setae. Dorsum of abdomen with three or four transverse folds in each segment, lateral sides without partitioning.

Pupae generally whitish in colour and exarate in form.

Key to the adults of curculionid stemborers

- 1' Scape inserted far away from the eye usually at midlength of the rostrum or between midlength and anterior tip of rostrum; body colour variable; rostrum short and stout; pronotum distinctly wider posteriorly; posterior end of abdomen usually not exposed 2

- 4' Midpronotum with broad median and lateral bands, not running parallel to each other; venter of abdomen lined with short fine hairs; male genitalia without sclerotised central area, ring segment absentLixus near germaini Hustache (Figure 20 a-l)
- 5' Elytra with a pale white V-shaped band of white hairs below midlength extending, with interruptions, laterally and incurved to the base of the wings, or V-shaped band absent and elytra heavily punctate _______6

- 7' Entirely brown without white scales forming a distinct pattern on the pronotum and elytra, elongated scales present in each elytral puncture
 9

- 10' Coxa III globose; ventral tibial spur in leg III acutely pointed and perpendicular to the femur; apodeme slightly sclerotised and straight towards apices; mandibular teeth 2-2

 Unknown genus sp. E (Figure 28 a-j)

Description of Curculionidae unknown genus larva and pupa (Figure 24 w-z4)

Diagnosis: Larva moderately crescent-shaped, curvature more pronounced towards the **posterior** abdominal area. Head **hypognathous**, bearing few hairs, longer (0.93 \pm 0.06 mm, n = 17) than wide (0.79 \pm 0.05 mm, n = 17), with a head length (HL): head width (HW)

ratio of 1.16 ± 0.07 . Epicranial stem long with a prominent endocarina. Frons broadly triangular with four transverse rows (2-2-2-4) of setae close to the endocarina and six setae lining the sclerotised frontoclypeal margin, corner setae about twice as long as the four submedian setae. Mandibles strong, almost triangular with two submedian setae and two apical teeth. Epipharynx with 14 setae in two transverse rows; pale, long and U-shaped labral rods. Maxillae of two segments, with lateral scars or light sclerotised margins. Outer flange with eight setae, three apical and five basal. Abdomen with distinct dumb-bell-like plicae. Prolegs absent but golden erect hairs present. Larvae of all five of the species of this genus are similar and very difficult to separate from each other.

Young pupa oblongate, exarate with a globose head and pointed abdominal tip. Anterior half of the globose head with seven setae and two each posterolaterally. Wing pads show 4–5 oblique striae. Abdominal tergites each with two pairs of subdorsal setae. Last abdominal segment heavily sclerotised. Older pupa compact, with broad to

rectangular pronotal plate lined marginally with fine hairs.

Biological information: The genus has been reared from three species of native grasses in Kenya; Cymbopogon nardus (L.) Rendle, Sorghum versicolor Anderss and Themeda triandra Forssk.

FAMILY LANGURIIDAE

General features: Adults with elongate, subcylindrical, flattened and narrow bodies, pronotum and elytra often different in colour pattern. Elytra commonly black, prothorax often red, some species with yellow elytra or posterior one-half of elytra black. Head large, recessed into the prothorax which reaches the hind eye margin, hiding the vertex. Antennae 11-segmented with 3–5 segmented club. Pronotum elongate or quadrate, with smooth lateral margins, always with paired furrows at base and with a transverse groove. Elytra elongate with apices truncate, or rounded, or pointed and separated with a groove in between. Dorsal surface of elytra may be shiny, glabrous or hairy. Legs simple with 5-5-5 tarsal formula. Procoxal cavities open behind, mesocoxal cavities closed externally by sterna. Abdomen with four visible sternites.

Larva often white, elongate, with ocelli, 3-segmented antennae, and bidentate mandibles. Head rounded to relatively long with short frontal arms and indistinct frontoclypeal suture. Maxillary palpi 3-segmented, labial palps 2-segmented. Thoracic legs present, each with a distinct terminal claw. Abdomen nearly parallel-sided, tergite IX bearing 1–2 upturned spines or urogomphi or a setiferous tubercle, X located posteroventrally and either rounded or cleft.

The pupa is exarate with pointed wing tips, large eyes on a rounded head and broad pronotum strongly rounded anteriorly and truncated posteriorly. Dorsum of abdonten with or without a subposterior transverse row of spines. Tergite IX with 1 or 2 urogomphi.

Biological information: Languriids are basically herbivores and detritivores. The larvae have been reported to feed on plant stems. For instance, Anadastus parvulus (Wiedemann) is a recorded pest of Italian millet, Setaria italica and the reddish-brown languriid, A. filiformis (F.) is a rice stemborer in Indonesia. With respect to languriids boring in the stems of tall grasses in Kenya, information is scanty. Eight species belonging to three genera—Barbaropus, Promecolanguria and Stenolanguria—were collected and reared on cut maize stems. These are true grass stemborers reported for the first time in Kenya. Setaria incrassata (Hochst.) Hack was the host of Barbaropus and Hyparrhenia rufa (Nees) Stapf was the host of Stenolanguria caudata Kraatz.

Key to the adults of languriid stemborers

- 4 Pronotum smooth and finely punctate, posterior longitudinal groove small and shallow ...

 Barbaropus near olseni Pic (Figure 44 a-m)

 4' Pronotum generally roughly punctate, posterior longitudinal groove short to long and

FAMILY MORDELLIDAE

General features: Commonly called tumbling flower beetles, adults are small (2–12 mm long) and characterised by the strongly wedge-shaped or humpbacked appearance. Body laterally flattened, pointed posteriorly with the terminal tergite sclerotised, and projected as a swordlike process beyond posterior margins of elytra. Head wider than long, transverse, and bent down covering the prosternum, pro- and mid-coxae. Lateral carina on pronotum complete. Legs with tarsal formula of 5-5-4, hind coxae very broad, tibiae and tarsi with oblique rows of comb-like spines, tarsal claws serrate or pectinate.

Larva white to glossy whitish-yellow, elongated, with a rounded head, long epicranial stem, robust mandibles and short legs without claws. Tergite IX with a pair of minute tubercles or median terminal spine. Pupa exarate, elongated with a large prothoracic shield extended to the upper margins of the eyes. Head subtriangular laterally, slightly bean-shaped frontally. Terminal segment of abdomen long with a posteromedian spine.

Biological information: Mordellids are generally decomposers feeding on decaying wood but some species have been reported to feed and mine in grass stems and other herbaceous plants (Booth et al., 1990). In Kenya, species belonging to the genus Stenalia were reared from stems of Hyparrhenia cymbaria (L.) Stapf, H. rufa (Nees) Stapf., Setaria incrassata (Hochst.) Hack., S. sphacealata (Schumach.) Moss and Panicum maximum Jacq.

Key to the adults of mordellid stemborers

- 3 Head black with a median longitudinal line; elytra yellow with thin black bands only on the inner and outer lateral margins; leg III spines 3:3:1:1 Stenalia sp. B (Figure 36 a-f)

- 4 Body entirely black with pale white hairs on the wings; oblique spines on leg III 3:1:1:0....

 Stenalia sp. D (Figure 38 a-e)

Larva of *Stenalia* nr. *occidentalis* Pic (Figure 35 g-k)

Description: Larva 4–6.8 mm long by 1.4–1.8 mm wide, shiny white and lightly sclerotised except the dark pigmented mouthparts and urogomphi at the tip of abdominal tergite IX; elongate to cylindrical with lateral abdominal swellings. Head abruptly curved downward with epicranial stem, endocarina and frontal arms absent, 3-segmented antennae very small, frontoclypeal suture distinct with anterolateral bulges near the antennae, ocelli visible. Mandibles symmetrical without mola, maxillae with small rounded mala. Prothoracic shield large, slightly sclerotised with the subtruncated anterior margin distinctly wider than the concave posterior margin. Legs very short with no distinct segments, coxae widely apart. Abdominal tergites I–VII each with a pair of posteriorly converging ovoid markings, tergite IX light brown, covered with hairs, four small spines arranged mid-dorsally in a longitudinal row, forked spine at the tip. Abdominal segment X slightly rounded posteroventrally. Spiracles annular, those on abdominal segment VIII the largest.

Pupa of Stenalia nr. occidentalis Pic (Figure 35 l-q)

Description: Pupa whitish-yellow to yellowish-brown, c. 6.5 mm long and 1.4 mm wide, exarate and slender, head bluntly rounded, almost entirely covered by the transverse and anteriorly rounded pronotum. Eyes large, bean-shaped, touching the lateral sides of the pronotum. Frons with a shallow transverse scar between the eyes, not reaching eye margins. Appendages broadly attached to the sides of the body. Abdominal segments, except terminal one, with a dorsal pair of parallel longitudinal lines; lateral edges bulging posterolaterally with a group of hairs on top of the bulge. Dorsal tip of last abdominal segment with a hooked spine.

FAMILY TENEBRIONIDAE

General features: The tenebrionids, or darkling beetles, are primarily scavengers, some feeding on plant materials like roots while others are serious pests of stored grain products. Generally, they are black or brown with a variety of shapes. The antennae are moniliform or incrassate with hidden bases. Eyes are distinctly emarginate and the lateral carina on the pronotum is simple and complete. Leg tarsal formula is 5-5-4.

Larva yellow brown to dark reddish-brown, prominently elongate and subcylindrical, heavily sclerotised with darkened head, reddish-brown mandibles and black urogomphi. Head with short to indistinct epicranial stem, mandibles concave and developed, ocelli absent, maxillae with rounded mala, and labium with 2-segmented palpi. Legs relatively similar in sizes with a sharply pointed claw. Abdomen parallel-sided with scars dorsad or posterior to the spiracles, anterior margins of each segment with or without punctures, tergite IX heavily sclerotised and punctate, dorsally flat with a pointed process or obliquely concave.

Biological information: Although tenebrionids are not known to bore stems of grasses, two genera—Paramarygmus and Zophodes—have been recently found inside the stems of Pennisetum polystachion (L.) Schult. in Busia District, Kenya. Both were mining the root system and feeding inside on the plant tissues producing deadhearts on young plants or whiteheads on flowering ones. Unfortunately, no pupae and adults were available during the preparation of this manual.

Key to the larvae of tenebrionid stemborers

- Abdominal tergites VIII and IX heavily punctate, tergite IX strongly concave dorsally and oblique laterally; frontal arm fully developed and broadly V-shaped; vertical scars present on posterior one-third of segments I–VII, absent on VIII Paramarygmus sp. (Figure 40 a-i)
- 1' Abdominal tergite VIII punctate only anteriorly; tergite IX entire, cylindrical and rounded posteriorly with a median tooth; frontal arm poorly developed; vertical scars longitudinal on segments I-VIII but not reaching posterior margin.......Zophodes sp. (Figure 41 a-g)

DIPTERA: MORPHOLOGY OF A DIPTERAN STEMBORER (Figure 50 a-d)

Key to the families of dipteran stemborers

- 1 Head with a pair of strongly diverging long stalks bearing the widely separated eyes and antennae; scutellum with a pair of long and diverging strong spines Diopsidae (p. 21)
- Ocellar triangle large and strongly developed; wing without anal cell, cell CuP and vein CuA +1A absent; cell M fused with the discal cell; costa with a break before the end of R₁... Chloropidae (p. 20)

FAMILY CHLOROPIDAE

General features: Adult chloropids are easily recognised by the strongly sclerotised, enlarged and well developed ocellar triangle, the absence of strong fronto-orbital setae, and convergent to cruciate postvertical setae. Costa of forewings with a single break near the apex of vein R_1 , subcosta not completely developed except at base, anal vein indistinct, discal cell and second basal cell running together, vein $M_3 + M_4$ usually with a flexure (weak or distinct) close to middle of discal cell, anal cell absent, alula prominently developed. Legs without strong setae. Abdominal tergites 1 and 2 confluent forming syntergite 1 + 2, longer than other tergites.

Larva usually white, varying from elongate and flat with narrowing anterior and posterior ends to oblong and cylindrical with acute head and rounded posterior end. In some species, larva with forked posterior tip, while in others the tip is rounded with small spiracular knobs.

Pupa coarctate with reddish-brown to dark brown sclerotisation.

Biological information: Three genera of chloropids—Elachiptereicus, Mepachymerus and Pachylophus—have been reared from tall grasses in Kenya. Elachiptereicus was commonly isolated from the yellowing and drying shoots of Rottboellia cochinchinensis (Lour.) W.D. Glayton. Panicum maximum Jacq., Setaria sphacealata (Schumach.) and Echinochloa pyramidalis (Lam.) Hitchc. & Chase were the hosts of Mepachymerus. Pachylophus sp. was reared from P. maximum.

Key to the larvae and pupae of chloropid stemborers

- 5 Anterior spiracles located dorsally and directed upwards and forwards; length about 5–6.5 mm and 1.5 mm wide...... Elachiptereicus abessynicus (Figure 54 b)
- 5' Anterior spiracles located ventrally; length 6.5-8 mm............... Pachylophus sp. (Figure 55 b)

FAMILY DIOPSIDAE

General features: Adults with long stalks arising laterally from each side of the head. Eyes and antennae located at or near the end of the stalks. Body slender. Wings with or without spots or bands. Scutellum with two strong diverging spines with or without apical bristle. Legs slender, femora I enlarged, with two rows of longitudinal ventral spines.

Larva dull white, flattened and narrowing on both ends. The two genera of diopsid grass stemborers—Diopsis and Diopsina—found on cogon grass, Imperata cylindrica L. in Kenya are

similar in shape and form.

Pupa dark yellowish-brown, c. 6.5 mm long and 1.5 mm wide with segment VI the longest; posterior end with a pair of widely separated porrect spines. Body segments uniformly lined with fine transverse striae.

Biological information: Diopsids are economically important in African agriculture. A number of species are pests, attacking maize, rice and wild Oryza barthii. The genus Diopsis has been reported as pest of both maize and rice. Diopsina sp., on the other hand, are usually found on wild grasses. Two genera of diopsids—Diopsina (one species) and Diopsis (two species)—were reared from Imperata cylindrica L. in Busia District, Kenya.

Key to the adults of diopsid stemborers

FAMILY MUSCIDAE

General features: Muscids are represented by a diverse group of flies making up several subfamilies. The features enumerated here are restricted to the muscid grass stemborers. Adults

with an angular head, face long and bare, antennal flagellomere long, arista bare. A single pair of reclinate orbital setae are present; hind tibia without a posterodorsal seta in apical one-half and wings bare except the costa. Palpi strongly developed. Thoracic setae very small, reduced in size. Wings with vein R₄₊₅ and M parallel at wing margin, A1+CuA2 reaching beyond halfway from crossvein CuA2 to wing margin. Male genitalia with hypopygium with surstyli developed normally, cercal plate may be highly modified, aedeagus with juxta reduced to a simple membranous sac. In the female, the ovipositor is long with cylindrical cerci set free from the membrane in nearly its entire length, tergite VIII not divided longitudinally, sternite VIII reduced to a pair of small hind marginal discs, three spermathecae present.

Larva elongate, typically narrow and tapered anteriorly but rounded to blunt posteriorly, cephalopharyngeal skeleton distinct with developed mandibles; abdominal venter with pseudopads; a pair of posterior spiracles present. A unique feature of stem-boring Muscidae is

the presence of oral bars adapted for cutting tough plant tissues.

Pupa barrel-shaped, elongate with both ends rounded and the **posterior** end with **spiracles**. Mature pupa always reddish-brown.

Biological information: Among the muscids, members of the genus Atherigona, subgenus Atherigona are recognised as true stemborers and are sometimes pests of cultivated grasses such as rice, maize, sorghum and millet (Davies and Seshu Reddy, 1981; Kalshoven, 1981).

Atherigona soccata Rondani (Figure 57 a-h)

Description: Males with dark frontal vitta, palp yellow with basal half brown; scutum without dark vittae, femur I and tibia I yellow; fore tarsus brown, except apical two-thirds of tarsomeres yellow. Wings with a weak tinge at tip of subcosta. Abdominal tergites 1 + 2 and 5 without spots or marks, tergite 3 with a pair of elongate to triangular spots at about one-half of tergal length, tergite IV with a pair of oval spots at one-third of tergal length.

Male genitalia: Tip of subtriangular trifoliate process with two strong lateral spines and two pairs in the middle; hypopygial prominence apically blunt in side view and concavely separated in anterior view.

Female with forelegs dark brown, femora yellow on basal two-fifth and tibia yellow on basal one-half or less; tergite VIII strongly W-shaped anteriorly and tergite VII separated and triangular in shape with a black longitudinal median band along anterior one half.

Mature larva c. 6–8 mm long, yellow to yellowish-white with black mouthparts, narrow

anteriorly and blunt to rounded posteriorly.

Pupa elongate, dark brown to reddish-brown; c. 4-5 mm long.

Biological information: Flies lay elongated white eggs singly in the morning and afternoon on the base of young seedlings. In two days the larva hatches and penetrates immediately into the shoots of young plants, later damaging the growing point. The maggot damage is called deadheart. In c. 2 weeks, larvae bore through the wall of the shoots. Pupation occurs in the surface layer of the soil. Adult emerges in 8–12 days and in well-irrigated areas where host plants are available year round many generations are developed. Maize and Panicum maximum are the most common host plants of A. soccata in Kenya.

FAMILY TEPHRITIDAE

General features: Adults are easy to recognise. Flies with spotted or banded forewings and a subcostal vein that bends sharply forward towards the costa, the bend at or near a right angle, before it totally fades. In the female, abdominal segment VI is well developed into a long process.

Larva commonly white, elongate with blunt **posterior** end and tapering **anterior** end. The abdominal venter has swellings or pads for movement of the larva.

Pupa elongate and oval.

Biological information: Tephritids are mostly fruit- and flower-feeders but some genera have been reared from the stems of wild grass species. The grass boring habit is rare in the Afro-

tropical region, being known only in the genus Bistrispinaria. In Busia District in Kenya, we reared Bistrispinaria magniceps (Bezzi) from Panicum maximum and Bistripinaria fortis (Speiser) from Pennisetum polystachion.

Key to the adults of *Bistrispinaria* from wild grasses in Kenya (after Hancock, 1999)

Note: We include three *Bistrispinaria* species in this key. Although we reared only *B. magniceps* and *B. fortis, Bistrispinaria woodi* was collected, though not reared, at one of our sampling sites (Suba District) in western Kenya.

until.

23

THE PARASITOIDS

General morphology of a parasitic Hymenoptera (Figure 58 a–f)

Key to the parasitoids associated with stemborers

to the parasitorus associated with stemsorers
Forewings developed, hindwings reduced to a pair of halteres; mouthparts forming a sucking proboscis (Diptera: Family Tachinidae)
Both pairs of wings present (or entirely absent); mouthparts mandibulate Hymenoptera 6
Proboscis very long and slender, abruptly bent (geniculate)
Proboscis short, not elongate and geniculate
Terminal segment of proboscis distinctly shorter than the penultimate segment; dorsum of abdomen with a grey longitudinal band, tip of abdomen black
Terminal segment of proboscis longer than the penultimate segment; abdomen yellowish
Head strongly triangular in profile, face deeply excavated; arista prominently thickened in its entire length; humeral callus with two setae; four dorsocentral setae present; wings with brownish bands along all veins
Not entirely as above
Sternopleuron with two setae
Sternopleuron with three setae
Forewing without closed cells, or wings (rarely) absent
Forewing with closed cells
Lateral sides of pronotum reaching the tegulae; forewings with or without stigmal vein; antenna with 8–10 flagellomeres
Lateral sides of pronotum not reaching the tegulae at base of forewing9
Forewing with stigmal and postmarginal vein; antenna with 9–10 flagellomeres; marginal vein thickened to form a pseudostigma; tergite I normal (Family Scelionidae)
Forewing without stigmal and postmarginal vein; antenna with 8 flagellomeres; marginal
vein not thickened to form a pseudostigma; tergite I long, curved and extended to the thorax (Family Platygasteridae)
Femur III enlarged and swollen, ventral margin with a tooth or row of teeth (Family Chalcididae)
Femur III normal, not enlarged, ventral margin without tooth11
Wings present, gena hairy, lightly and finely punctate, a longitudinal groove present below ocelli; scape long and slender, as long as pedicel and three funicular segments; mandibles slightly retracted to hidden; wings with longitudinal brown tinges
Wings absent. Gena coarsely punctate, longitudinal groove below eyes indistinct; scape slightly short and stout in basal one-half, length less than pedicel and two funicular segments; mandibles large, distinctly protruding below Hockeria sp. (Figure 67 a–d)
Frons with H-shaped impressed marks; antennal toruli much closer to eyes than to each other; forewing without postmarginal vein, stigmal vein reduced to a short process usually arising far before midanterior margin, marginal vein not elongate and venation not beyond basal third of wing; blade of hindwing not reaching base, modified to a linear stalk formed by the submarginal vein; tarsi 5-segmented; petiole short, broader than long; antenna 11-segmented with 8 funicular segments and unsegmented club (Family Mymaridae)

11'	Frons without H-shaped lines; antennal toruli usually closer to each other than to eye; other features not as above
12	Legs with 4-segmented tarsi (Family Eulophidae)
12'	Legs with 5-segmented tarsi
13	Scutellum with two distinctly parallel Iongitudinal grooves; mesonotum with a median groove; propodeum mesad of spiracle with an inverted Y-shaped carina; submarginal vein with one or two dorsal bristles; tibial spur on leg III very short and small
13'	
13	propodeum with two median carina; submarginal vein with more than three bristles; tibial spur noticeably long in leg III
14	Hind tibial spur reaching well beyond apex of hind tarsus; antennal scape 4x length of its greatest width; eyes with indistinct hairs; ocelli forming a distinctly acute triangle; frontoclypeus about 2x dorsal breadth of eyes; scutellum strongly elongate-reticulate laterally, median band of moderate width distinctly finely reticulate; spiracle elongate-oval; head dark blue green to blackish blue
14'	Hind tibial spur hardly reaching the apex of hind tarsus; antennal scape distinctly curved, about 5x length of its greatest width; eyes sparsely but distinctly hairy; ocelli forming an obtuse triangle; frontoclypeus more than 2x dorsal breadth of eyes; scutellum with weaker uniform sculpture, narrow medially but median band glossy and smooth; spiracle circular; head bluish-black
15	Pronotum subrectangular and prominent; notaular line incomplete; body non-metallic, usually black; mesopleura large; propodeum sloping; median longitudinal groove present; gaster convex (Family Eurytomidae)
15'	Without the above combination of characters
16	Forewings infuscated on and below the stigma ; petiole distinctly long; tergite IV narrow; ground colour yellow
16'	Forewings without brown infuscation; petiole short; male with five funicular segments; tergite IV the longest; ground colour mostly black
17	Legs uniformly yellow including coxae III; head, thorax and abdomen black with deep, rough punctations clothed with white hairs on the head and thorax; hairs in the face above the mouth distinctly converging; mesonotum with a deep notaular line; propodeum with a narrow median groove and light transverse striae; funicular segments black
17'	Not as above18
	Middle of face with a slightly elevated, hairless triangular plate, upper half with marginal white hairs converging towards each other; gena with fine reticulations; scape yellow; femora and tibiae black with yellow apices, tarsi yellow except black terminal segment Eurytoma sp. A (Figure 74 a-c)
18′	Middle of face without a triangular, hairless plate; gena usually roughly punctate; scape generally black; femora may be fully black and tibiae dark reddish-brown medially and yellow on both ends or variable
	Lower face close to the gena bulging moderately, facial hairs directed downwards; mandibles reddish-brown except black teeth and base; tibiae medially blackish-brown to dark reddish-brown with yellow ends; antennae brownish-yellow with black pedicelEurytoma sp. B
	Lower face not bulging; facial hairs converging or totally absent; mandibles black or yellow with black teeth; tibiae uniformly yellow or black; antennae black or dark reddish-brown with yellow scape
	Head and thorax smooth, without white hairs or punctation ; mesonotum without prominent lobes, fine transverse striations distinct; femur and tibia black
	Head and thorax roughly punctate and with white hairs; mesonotum distinctly lobed and roughly punctate ; legs yellow except dark brown to black outer basal one half of femora Eurytoma sp. D (Figure 76 a-d)

21	close to hind end of mesopleuron; metanotum longer than propodeum; mesonotum with lateral carina; pronotum with a forked spine (Family Eupelmidae)
21′	Wings present. Mesopleuron not large and with a femoral groove; coxa II not inserted close to hind end of mesopleuron; mesonotum without lateral carina; pronotum without spine (Family Pteromalidae)
22	Hindwing without closed cells; body elongate, dorso-ventrally flattened; head elongate and flat, longer than wide and directed forwards; forewing with only the basal and subbasal cells present (Family Bethylidae)
22'	Hindwing with closed cells; body not dorso-ventrally flattened; forewing with many closed cells
23	Forewing with only one recurrent vein (Family Braconidae)
23'	Forewing with two recurrent veins (Family Ichneumonidae)
24	Second intercubitus (2rs-m; Figure 58e) of the forewing absent
24'	Second intercubitus of the forewing very distinct
25	Notaular lines very prominent; abdominal tergite I very small, anterior and median transverse grooves of gaster very distinct, posterior pale or indistinct; mid posterior groove slightly swollen forming a small hump; occipital carina very prominent; middle of scutellum smooth and globosely elevated; antenna 21-segmented, brownish except yellow scape , pedicel and funicular segments F1 and F2; segment F1 long, as long as combined length of F2 + F3; legs yellowish-brown
25'	Notaular lines absent; tergite I forming a distinctly squarish or subtriangular rugose plate; funicular segment F2 as long as F1; legs mostly yellow with brownish coxae
26	Ovipositor sheath always shorter than half the length of tibia III, seldom protruding beyond abdominal tip; abdominal tergite I narrow anteriorly and broad posteriorly
26′	Ovipositor sheath longer than half length of tibia III, prominently protruding beyond tip of abdomen; abdominal tergite I nearly parallel-sided(Genus Dolichogenidea) .29
27	Coxa III usually light brown to brownish-yellow; aedeagus of male truncate at apex
27'	Coxa III usually black or dark reddish-brown; aedeagus of male rounded at apex
28	Coxa-III black; rough area in tergite II wide, not distinctly raised
28′	Coxa III brownish-red; rough area in tergite II narrow and distinctly elevated
29	Tarsus III black; ventrolateral segments II-V of abdomen yellow
29'	Tarsus III yellow; ventrolateral segments II-V of abdomen dark brown to black31
30	Apical one-fifth of tibia III reddish-brown; scutellum with very sparse punctation
30;	Apical one-half of tibia III dark reddish-brown; scutellum smooth and shiny
	Coxa III brown except yellow posteroventral half; middle of propodeum without wide groove, anterior two-thirds moderately rough and punctate; tibia III entirely yellow
31′	Coxa III with light brown apical one-fifth; middle of propodeum with a wide groove, anterior lobe distinctly punctate
32	Abdomen forming a strong carapace, or with an inverted, canoe-like shape with at most three visible tergites
32′	Abdomen not forming a carapace; with more than three visible tergites

33	Ground colour yellow, except stigma and lateral and posterior sides of scutellum brown; propodeum relatively smooth; abdomen with three visible tergites
33	Ground colour black except a broad yellow subanterior transverse band; propodeum rough, with squarish and circular punctation; abdomen without visible transverse lines
34	Abdominal tergites entirely longitudinally sculptured with fine ridges
34	Abdominal tergites not entirely longitudinally sculptured
35	Tip of abdomen, lateroventral margins of meso- and metathorax, posterodorsal part of pronotum, and anterior part of scutellum black
35	Body uniformly orange-yellow
36	Second intercubitus (2rs-m) pale; total length of antenna about as long as body
36′	Second intercubitus (2rs-m) distinct; antenna much longer than body
37	Femur III with a ventral spine; abdominal tergites 2 and 3 uniformly sculptured; wings hyaline without bands
37′	
38	Body orange-red, except pronotum, scutellum, most of propodeum, mid-dorsum of coxa III, distal dorsal part of tibia III and ocellar triangle black Habrobracon sp. A (Figure 93 a–h)
38'	Body orange-red except blackened ocellar triangle and vertex
39	Ocellar triangle on a narrow black spot extending at the back of the head towards the occiput
39'	Ocellar triangle on a broad black band on top of the head
40	Body uniformly yellow to light yellowish-brown; wings hyaline; ovipositor about one-half body length; propodeum with an inverted Y-shaped median carina; tergite I rough with distinct lateral ridges along anterior one-half, converging posteriorly
40'	Body orange to light orange-red, wings with grey-brown longitudinal bands; antenna and ovipositor very long, the latter distinctly longer than abdomen; propodeum smooth and shining; tergite I without elevated ridges laterally
41	Wings dark brown with the proximal half of stigma yellow and a narrow longitudinal whitish band bisecting the middle of first cubital cell and the recurrent vein; ocellar triangle on a narrow black spot; tergite I elevated with smooth lateral margins and strongly rounded anterior end; scape black; body length 7.5–11 mm
41′	Wings with brown tinge as above except for a broader light white shade in the first cubital and outer half of first discoidal; ocellar triangle on a large black spot; tergite I not strongly raised, lateral margins heavily punctate and with a nearly truncated anterior end; scape mostly yellow; body length 7.5 mm
42	Tip of forewing with a brown tinge, areolet distinctly stalked; submetapleural carina forming a strong lobe right behind the middle coxa; propodeum smooth without longitudinal carina; tergite I about twice as long as wide; subgenital plate of female conspicuously large when viewed laterally; body generally reddish-brown except black lower portion of mesopleuron, coxae II and III and femur III
42′	Forewing hyaline without any brown markings; other characters not as above 43
	Femur III with a large ventral spine and other minute spines subposteroventrally 44

43′	Femur III without ventral spine
44	Area superiomedia narrowing posteriorly with lateral ridges fading, forming an opened and striated posterior end; propodeum dark brown in apical one-third, reddish-brown medially and yellow posterior tip; mesonotum with fine transverse striae becoming circular medially and rough posteriorly; abdomen dark reddish-brown
44'	Area superiomedia closed posteriorly; propodeum entirely black or with combinations of yellow and black; mesonotum relatively rough
45	Propodeum black anteriorly, lateral sides and posterior one-half yellow; mesonotum with an M or W-shaped yellow band
45'	Mesonotum either with a U- or V-shaped yellow band, relatively finely punctate, or with a median rectangular black band; scutellum smooth or punctate
46	Mesonotum with U- or V-shaped yellow band; propodeum light brown; scutellum punctate
46'	Mesonotum without U- or V-shaped yellow band, but with antero-medial rectangular black band; propodeum black; scutellum smooth
47	Antennal segments 14–18 white, 1–11 and 19–31 brown and 12–13 yellow; tergites V–VI black, I–IV reddish-brown, VII black along basal one-half and VIII yellowish-white
47'	Antenna without white segments; tergite pattern entirely different
48	Tergite I prominently long and slender, 4x longer than wide at its widest point; clypeus not clearly separated from the face; vein 2Rs of areolet 2x longer than 1Rs; radiella, cubitella, discoidella and brachiella absent from hindwing, although radiella shows a short arm; abdomen compressed laterally, tergites longer than wide (Genus Venturia)
48′	Tergite I broadly attached to the propodeum, as wide as long; clypeus distinctly separated by the frontoclypeal suture from the face; vein 2Rs of areolet as long as 1Rs; radiella, cubitella, discoidella and brachiella all complete in the hindwing; abdomen flattened dorsally, tergites as wide as long (Genus Holcopimpla)
49	Coxa I yellow; femur III reddish-brown; tergite I black, reddish-brown posterolaterally Venturia sp. A (Figure 105 a-d)
49′	Coxa I all or mostly black; femur III brown to blackish-brown; tergite I black or with reddish-brown posterior one-third
50	Coxa 1 and tergite I uniformly black
	Coxa black with yellow distal end; tergite I black with reddish-brown distal one-third Venturia near jordanae Fitton (Figure 106 a-c)
	Femur III and coxae I–III black; area superomedia narrow, constricted medially and roughly punctate posteriorly
	Femur III brown; coxa I black, II and III black with yellow apical ends; area superomedia wide with lateral ridges parallel to each other at midpoint, widening below, forming parallel ridges again posteriorly, and covered with white silvery hairs
	Areolet trapezoidal, 2Rs 1.25x longer than 1Rs; forewing length 5.4 mm; head width: height 1.1:1.0; brachial cell 2.2x longer than wide; body length c. 8–10 mm
1	Areolet almost triangular, 2Rs only 1.1x longer than 1Rs; forewing length 7 mm; head width:
	height 1.4:1.0; brachial cell 2x longer than wide; body length c. 10.5 mm

PLATES

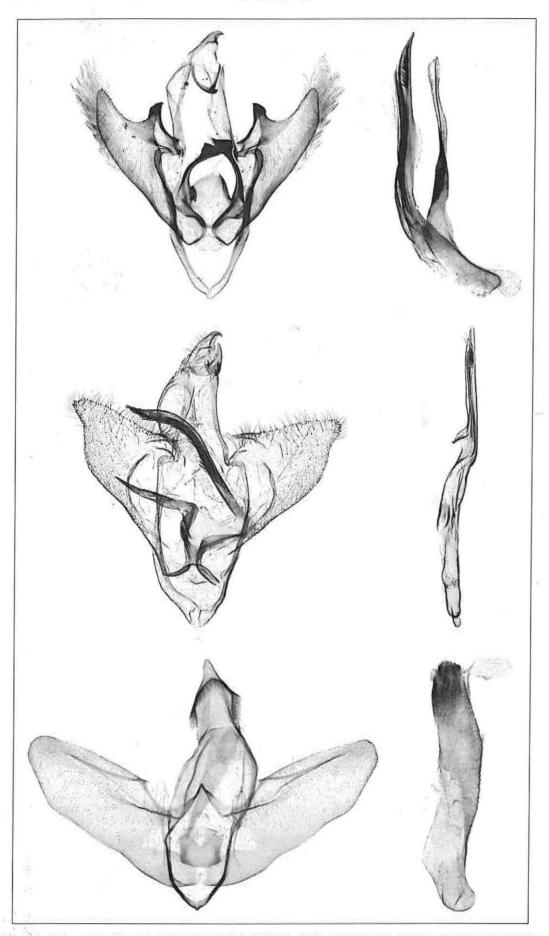


Plate 1. Male genitalia and aedeagus: Top to bottom, Chilo partellus; C. thyrsis; Eldana saccharina

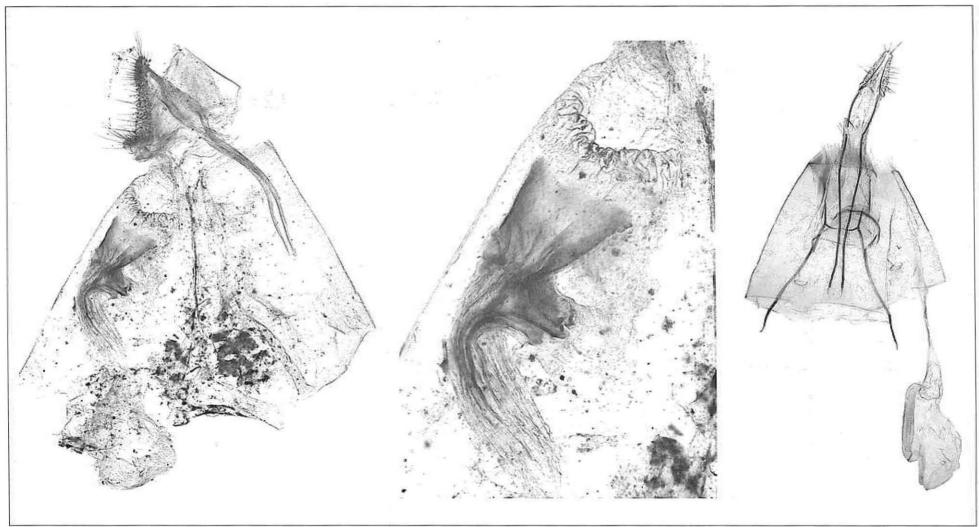
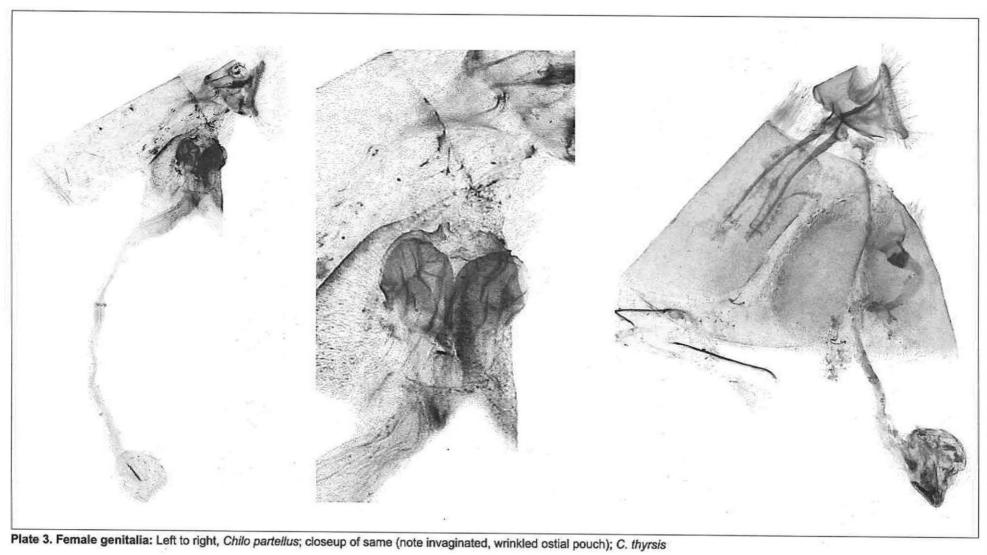


Plate 2. Female genitalia: Left to right, Chilo incertus; close up of C. incertus showing heavily sclerotised ostial pouch; Eldana saccharina (note pronounced appendix in corpus bursae)



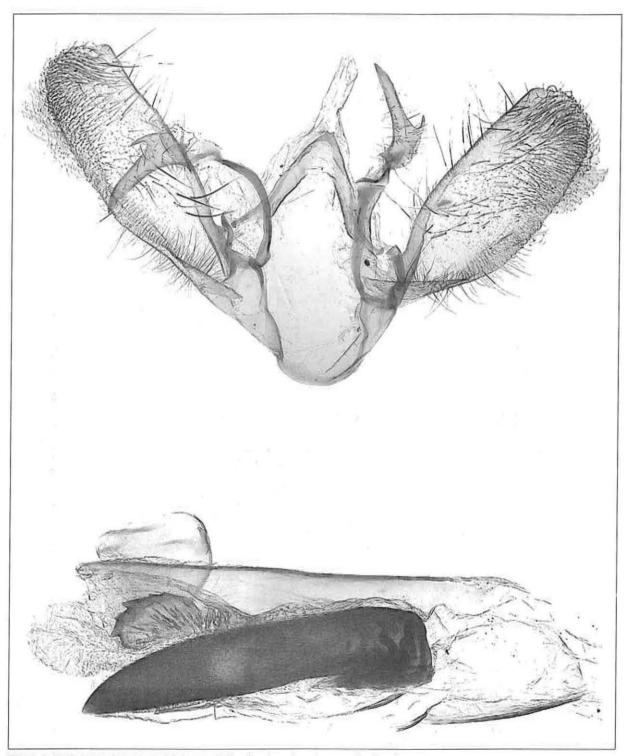


Plate 4. Peoriinae Taxon A: Male genitalia: (top) and aedeagus (bottom)

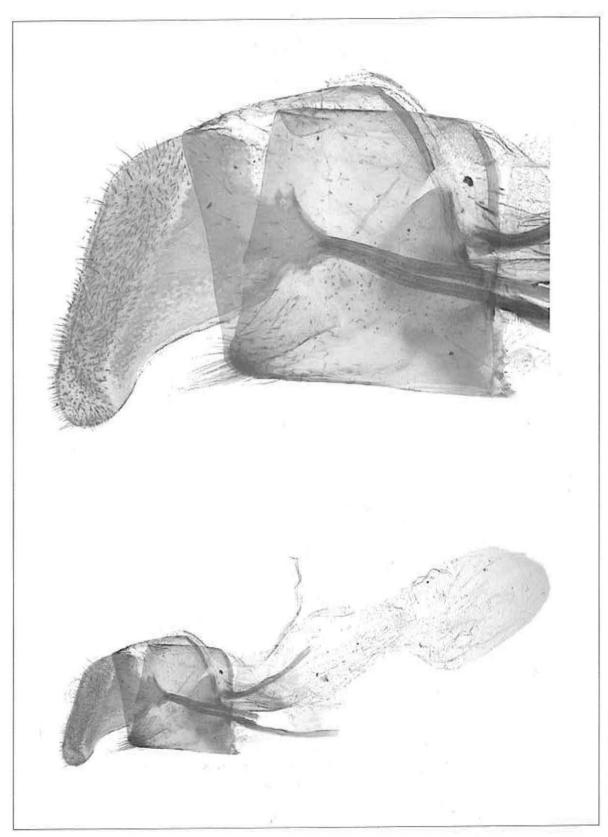


Plate 5. Peoriinae Taxon A: Female genitalia (left) with closeup of ovipositor (right)

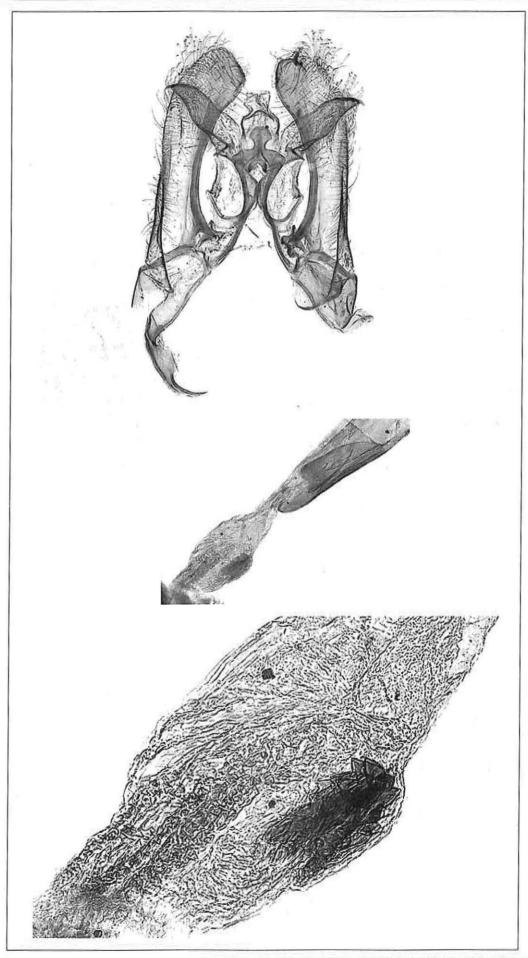


Plate 6. Maliarpha concinnella: Top to bottom, male genitalia; aedeagus, closeup of aedeagus showing cornutus with teeth

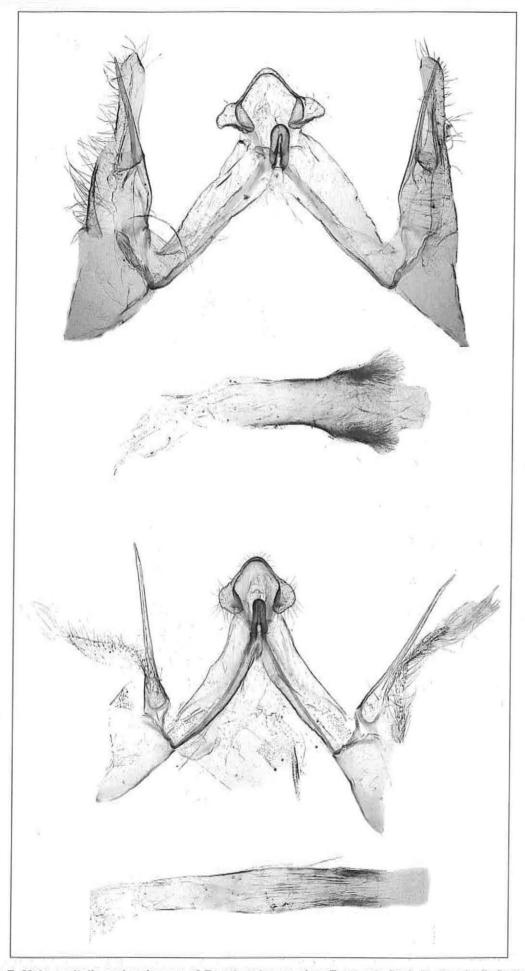


Plate 7. Male genitalia and aedeagus of Ematheudes species: E. sp. nov. (top); E. straminella (bottom)

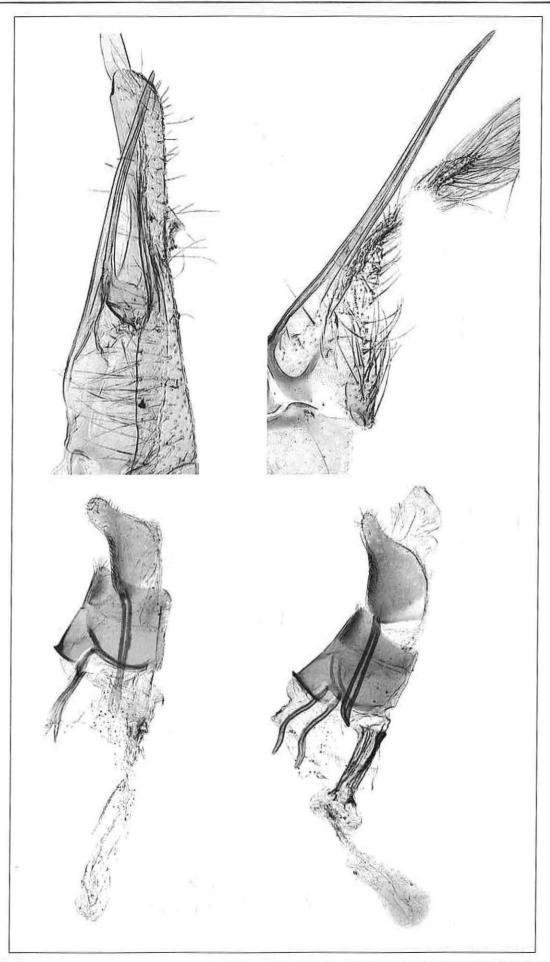


Plate 8. Ematheudes species: Top row, male costal spines of E. sp. nov. (left) and E. straminella (right); bottom row, female genitalia of E. sp. nov (left) and E. straminella (right)

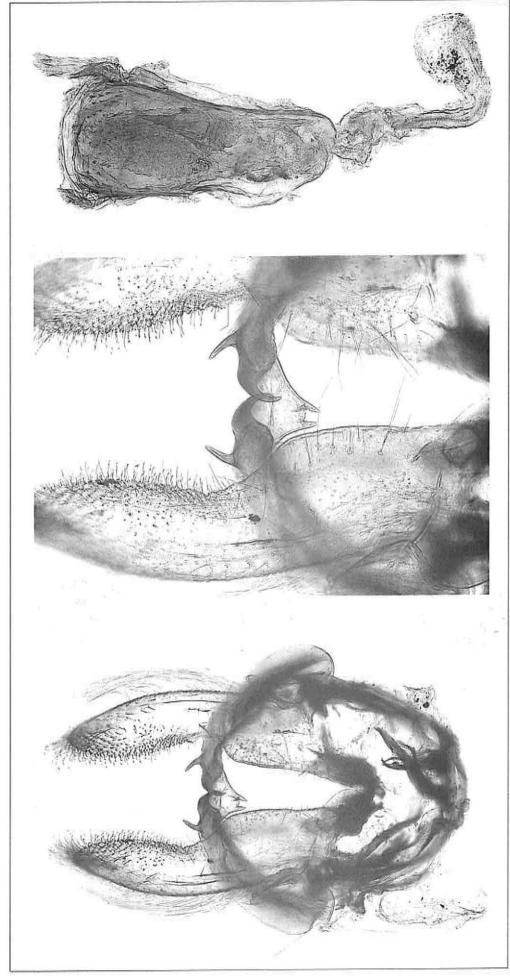


Plate 9. Peorilnae Taxon B: Left to right, male genitalia; closeup of same showing hooked processes; aedeagus

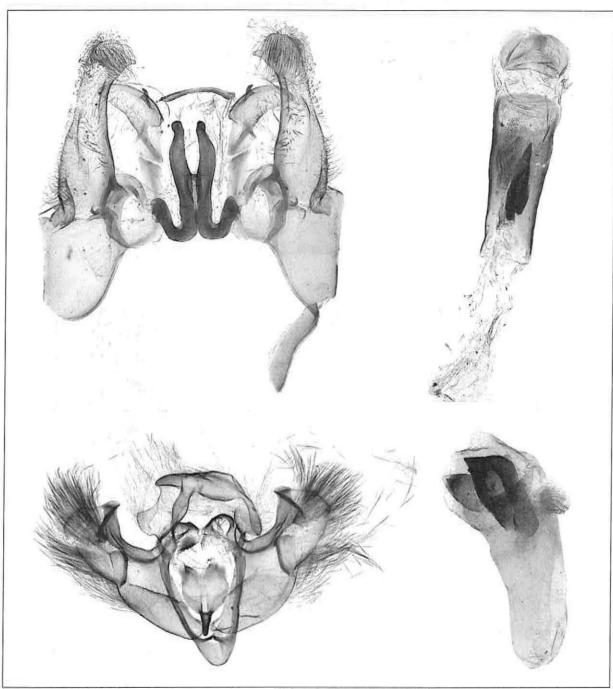


Plate 10. Male genitalia and aedeagus: Saluria lentistrigella (top); Sciomesa piscator (bottom)

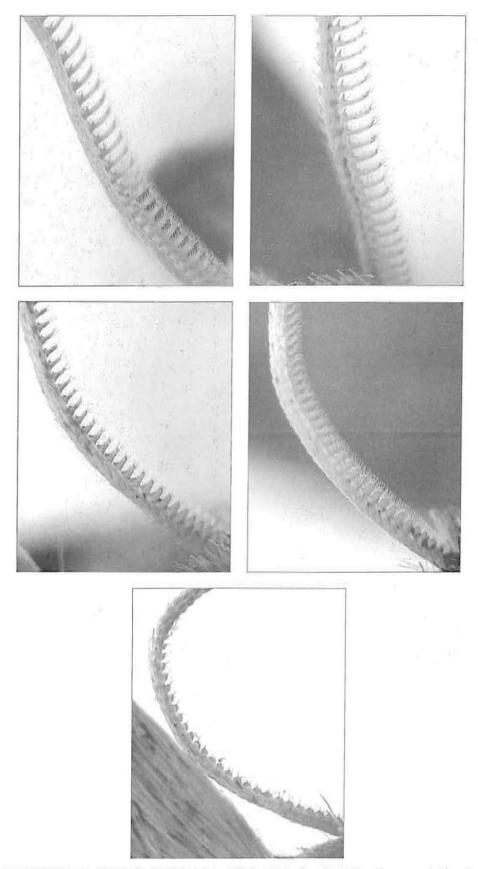


Plate 11. Male antennae of Sesamia species: top, left to right, S. calamistis, S. nonagrioides botanephaga; middle, left to right, S. penniseti, S. poephaga; bottom, S. sp. nov.

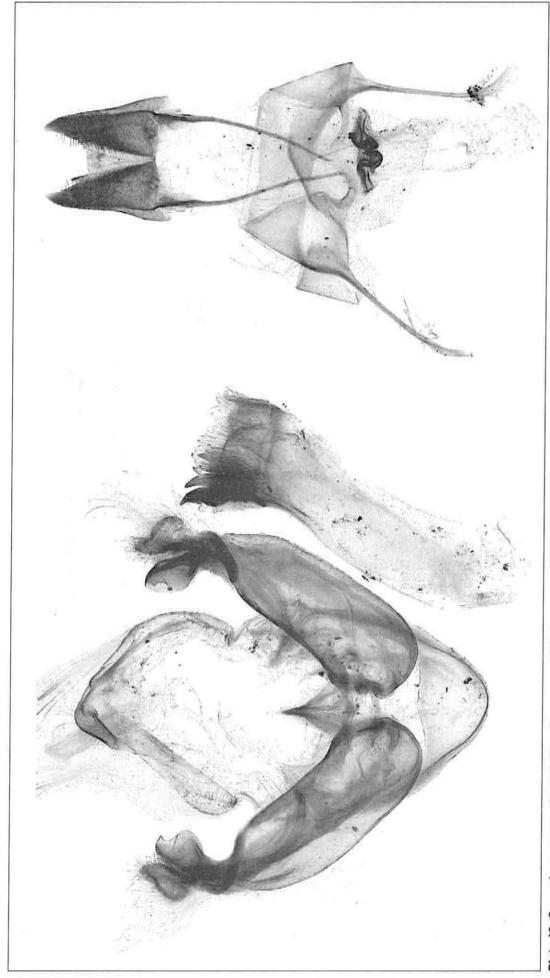


Plate 12. Sesamia sp. nov: male genitalia and aedeagus (left); female genitalia (right)

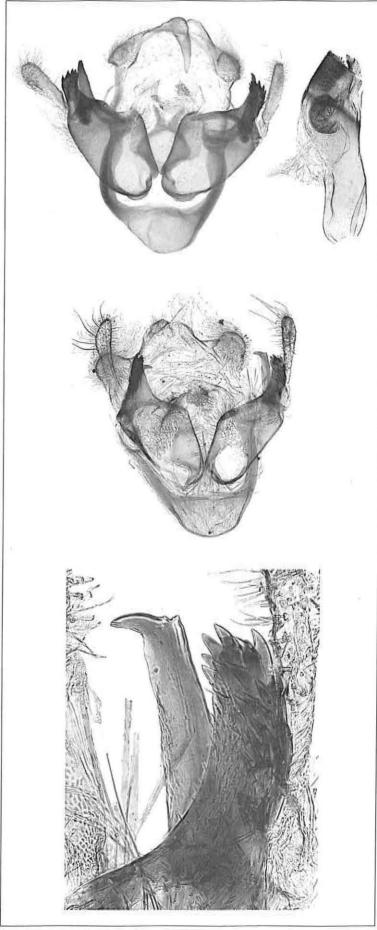


Plate 13. Sesamia calamistis: male genitalia showing centrally placed flask-shaped juxta (top left) and aedeagus (top right); male genitalia showing more common, nearly truncate variant of costal spine (centre); closeup of costal spine and robustly-toothed saccular process (bottom)

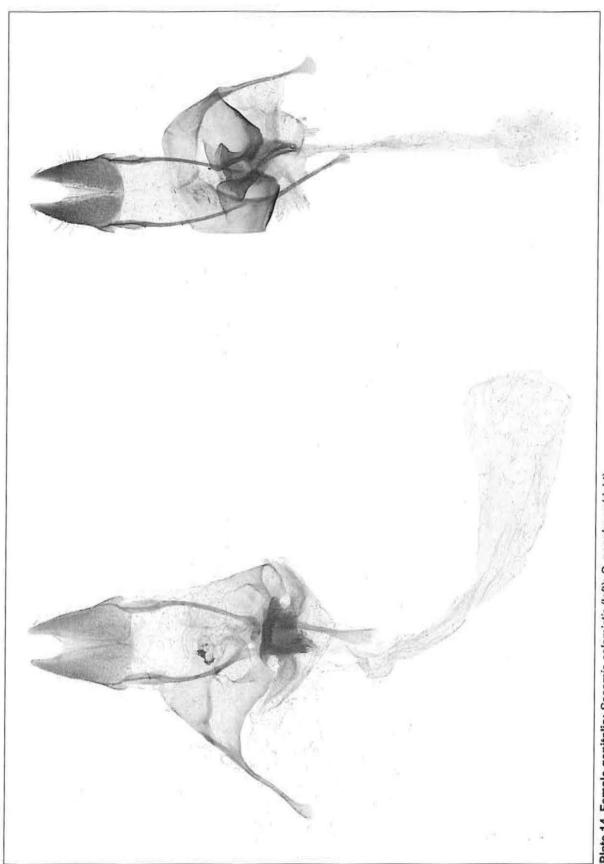


Plate 14. Female genitalia: Şesamia calamistis (left): S. poephaga (right)

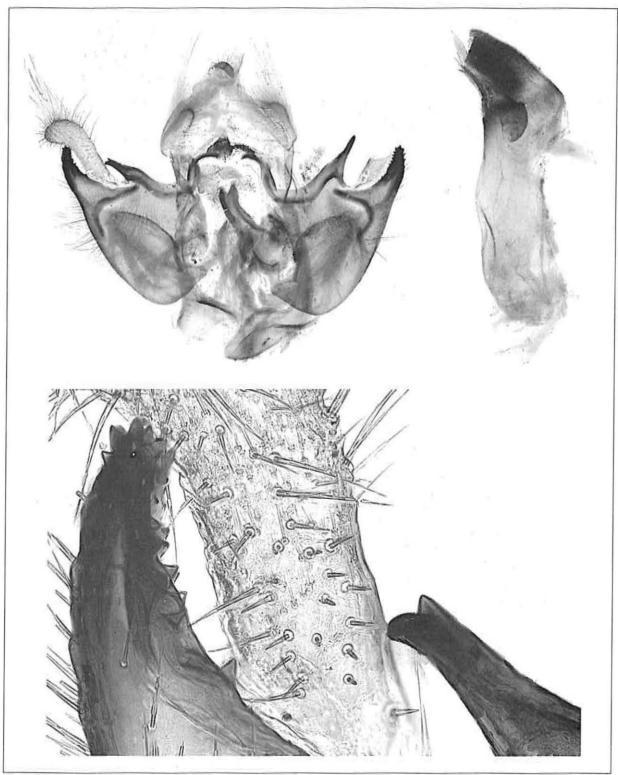


Plate 15. Sesamia nonagrioides botanephaga: male genitalia showing centrally-placed juxta with extended neck (top left); aedeagus (top right) and closeup of costal spine and moderately-toothed saccular process (bottom)

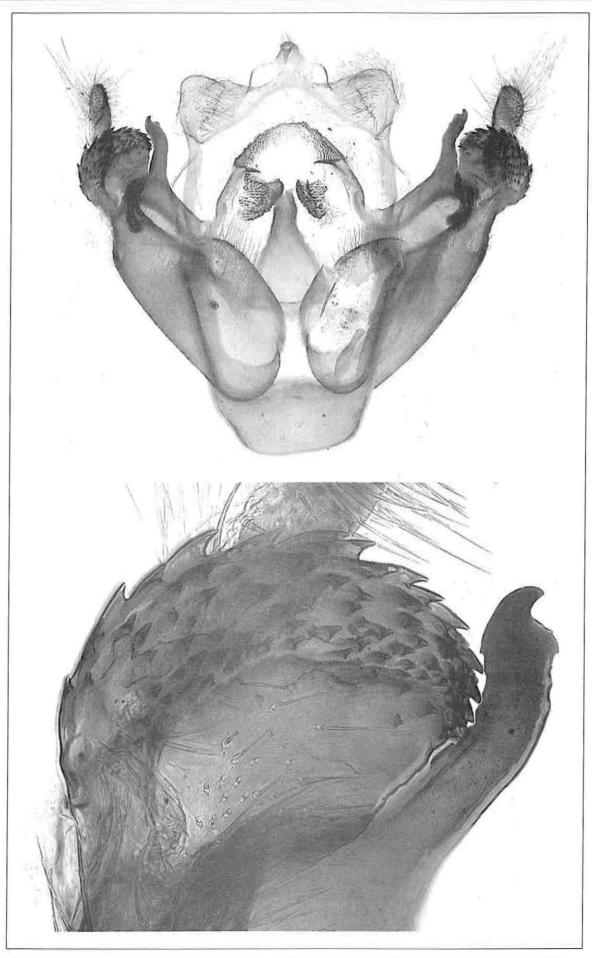


Plate 16. Sesamia penniseti: male genitalia (top); closeup of robustly-spined saccular process and spatulate, sharp costal spine with irregular, fine teeth (bottom)

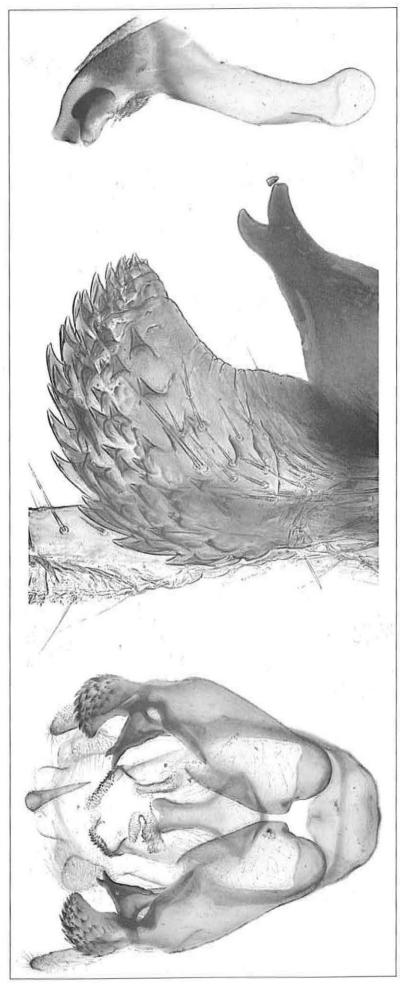


Plate 17. Sesamia poephaga: male genitalia (left); closeup of robustly-spined saccular process and bifid costal spine (centre); aedeagus (right)

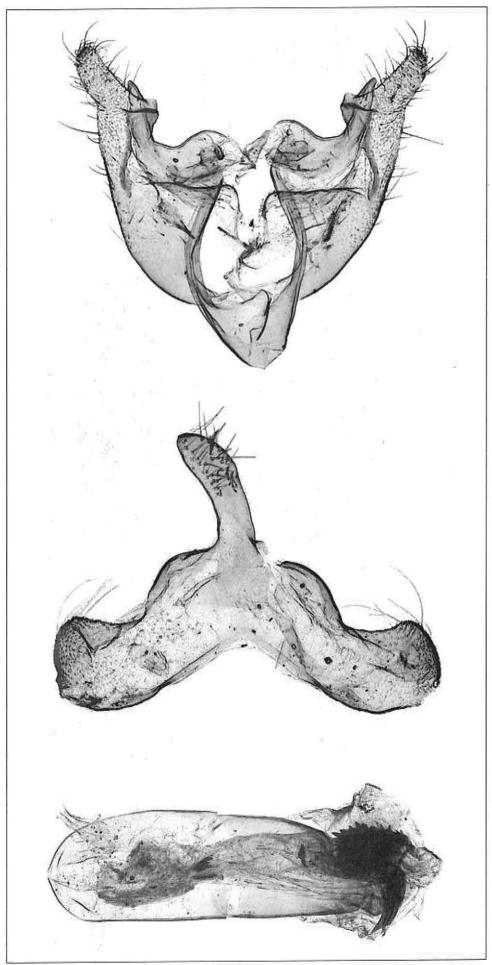


Plate 18. Sciomesa cf. sp. nov. top to bottom, male genitalia; closeup of uncus; aedeagus

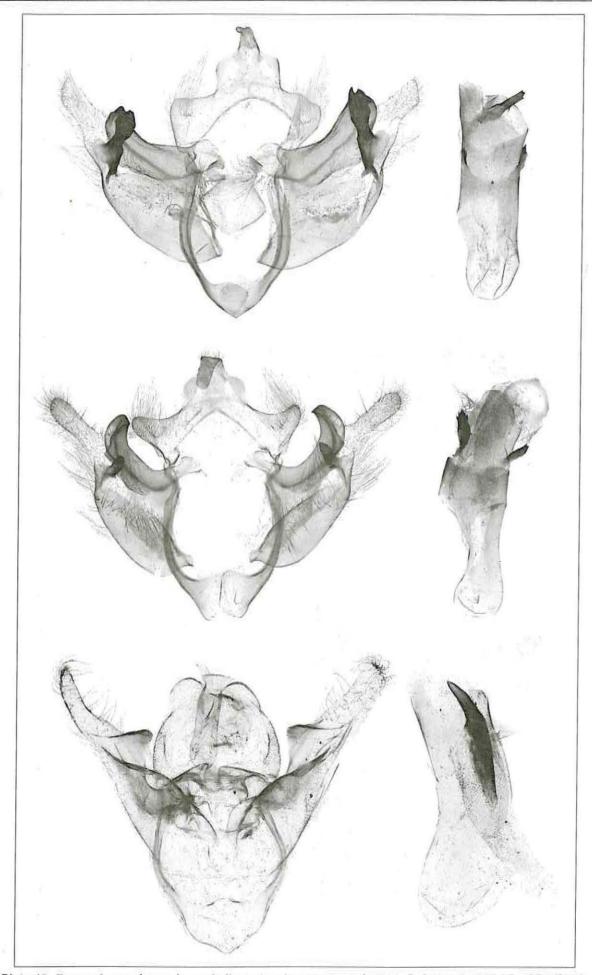


Plate 19. Busseola species male genitalia and aedeagus: top to bottom, B. fusca; B. phaia; B. obliquifascia

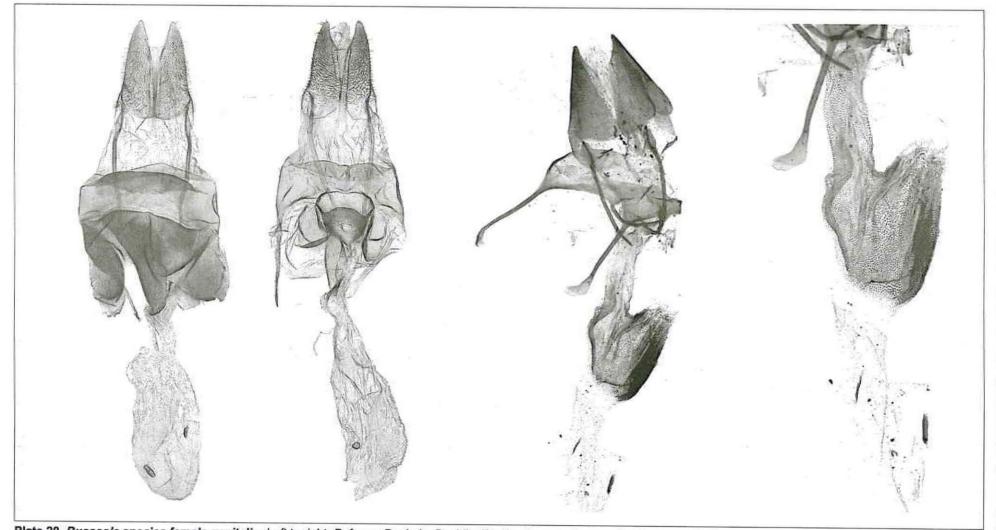


Plate 20. Busseola species female genitalia: Left to right, B. fusca, B. phaia, B. obliquifascia; closeup of same showing appendix-like structure in corpus bursae and two linear signa

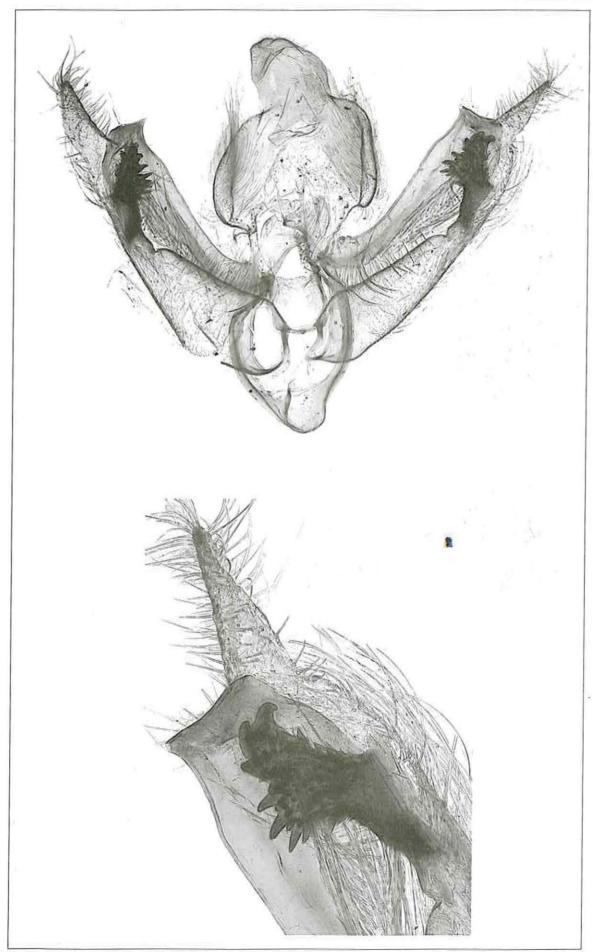


Plate 21. Manga nubifera male genitalia: male genitalia (top); closeup of same showing flange-like costal spine and heavily toothed saccular process (bottom)



Plate 22. Manga nubifera: left to right, male aedeagus; female genitalia; closeup of same showing asymmetrical sclerotisation of ductus and two ellipsoid, ridged signa

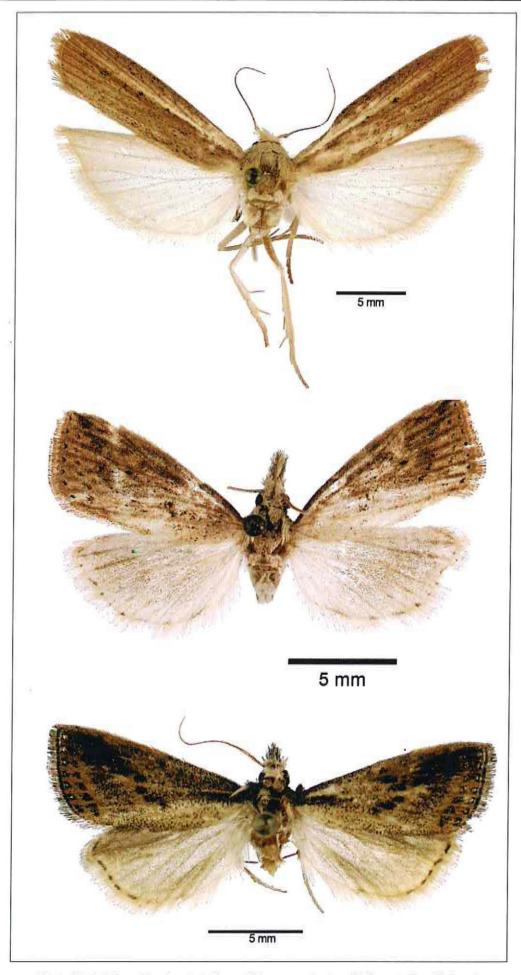


Plate 23. Adult moths: top to bottom, Eldana saccharina; Chilo partellus; C. thyrsis

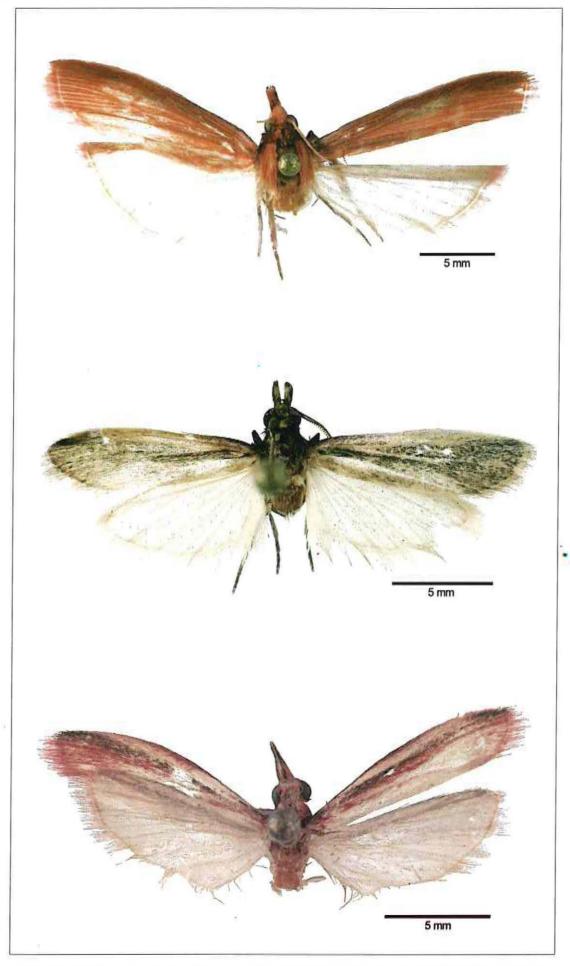


Plate 24. Adult moths: top to bottom, Peoriinae taxon A; Peoriinae taxon B; Maliarpha concinnella

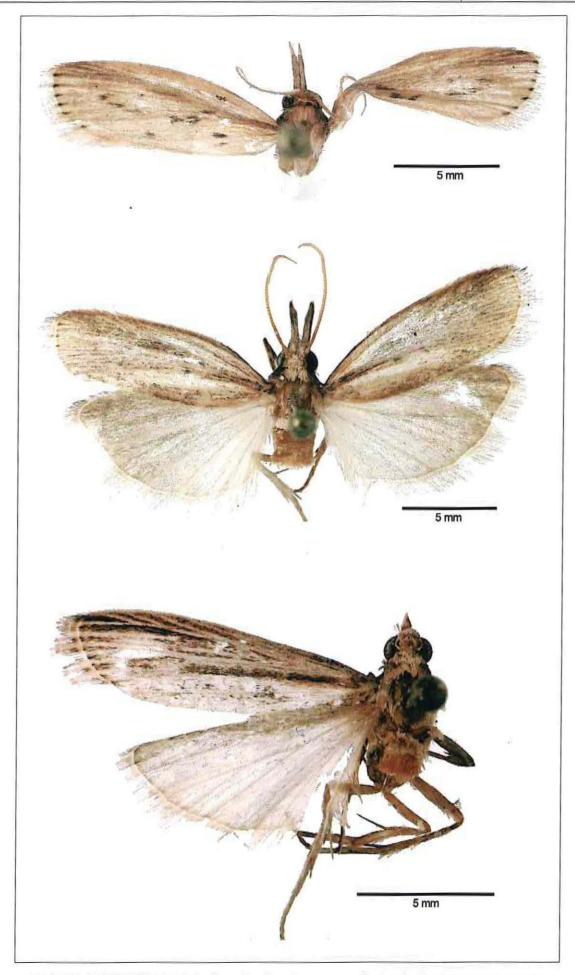


Plate 25. Adult moths: top to bottom, Ematheudes sp. nov.; E. straminella; Saluria lentistrigella

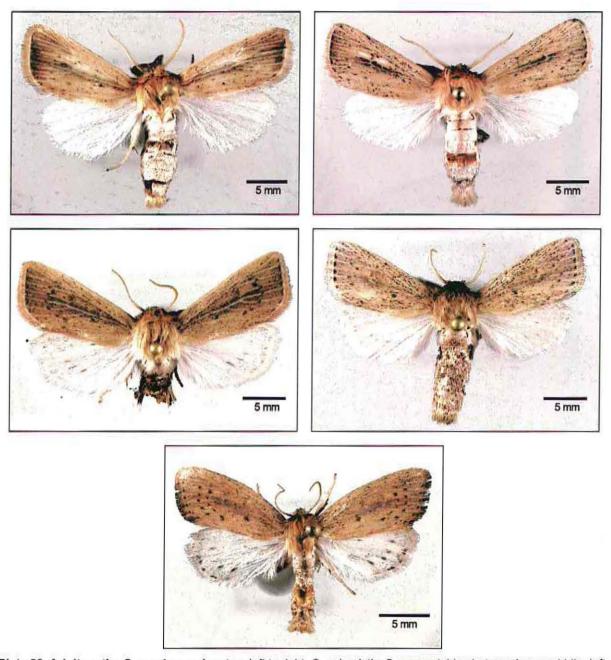


Plate 26. Adult moths Sesamia species: top, left to right, S. calamistis; S. nonagrioides botanephaga; middle, left to right, S. penniseti, S. poephaga; bottom, S. sp. nov.

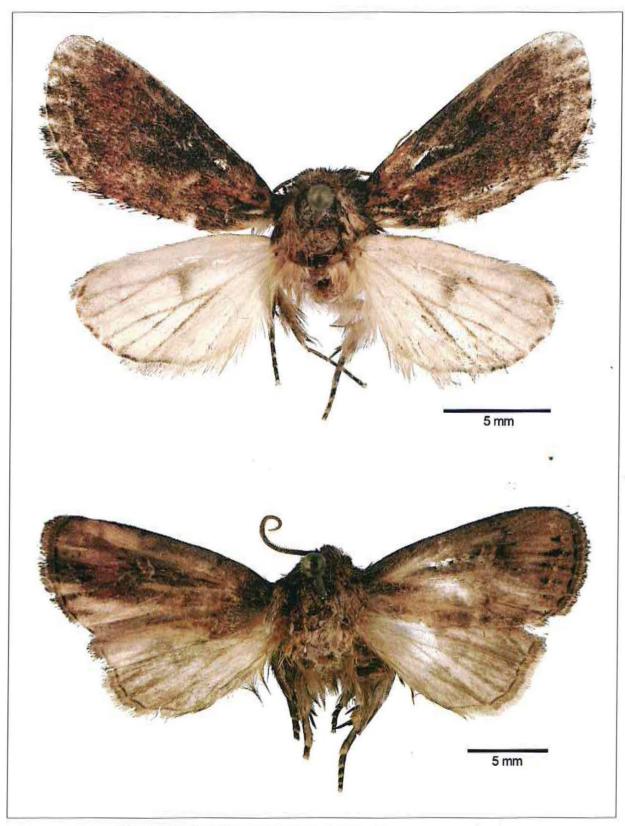


Plate 27. Adult moths: top to bottom, Manga nubifera; Sciomesa piscator

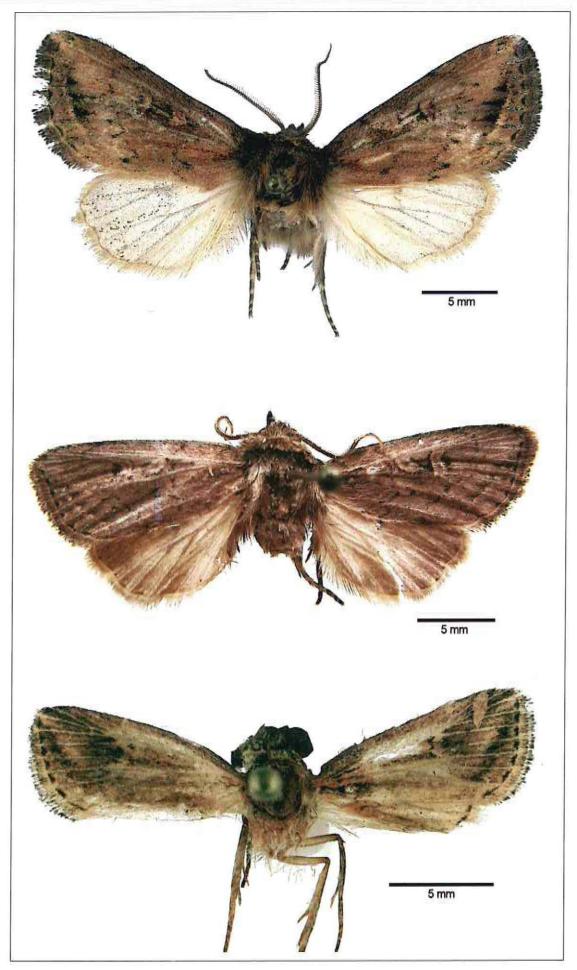
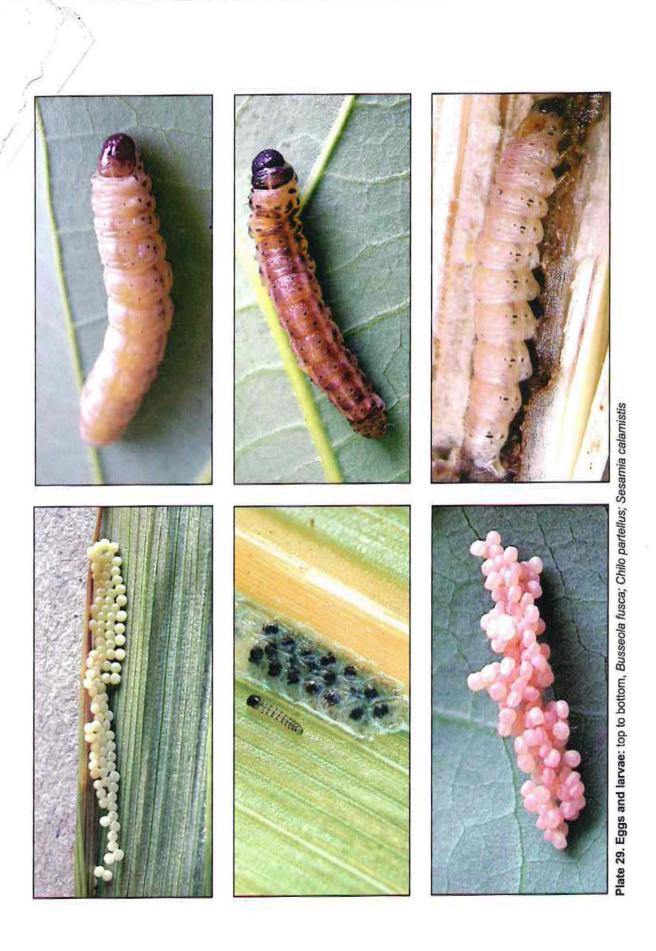


Plate 28. Adult moths, Busseola species: top to bottom, B. fusca; B. phaia; B. obliquifascia



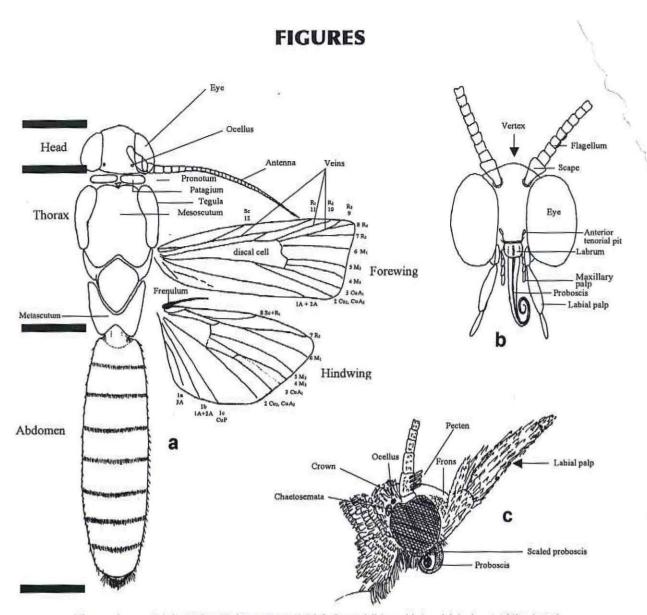


Figure 1 a-c. Adult moth with legs removed (a), frontal (b) and lateral (c) views of the head

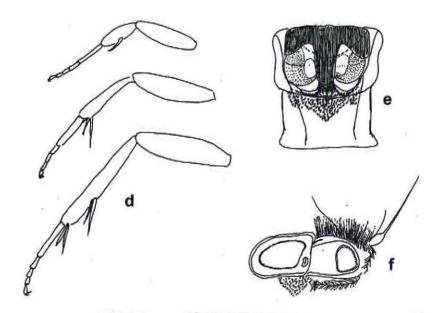


Figure 1 d-f. Legs I-III (d), tympanal organ in family Pyralidae (e) and in Noctuidae (f)

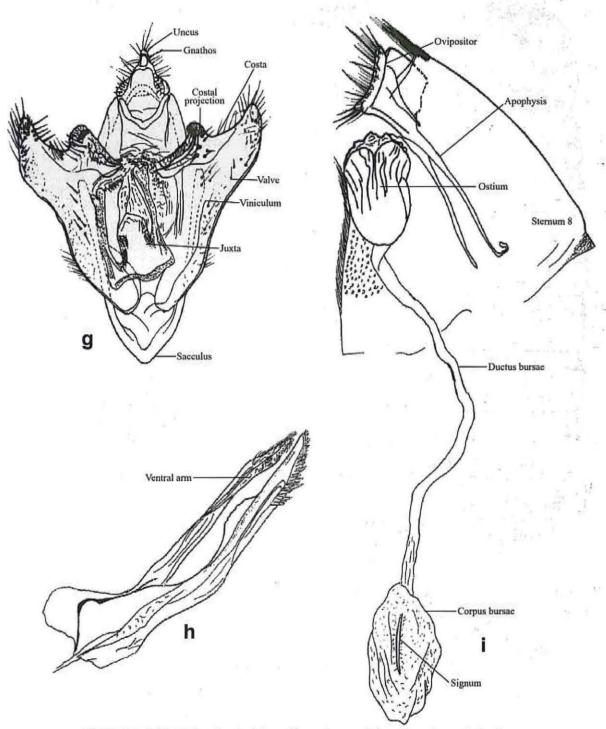


Figure 1 g-i. Genitalia of male (g) and its aedeagus (h) and female genitalia (i)

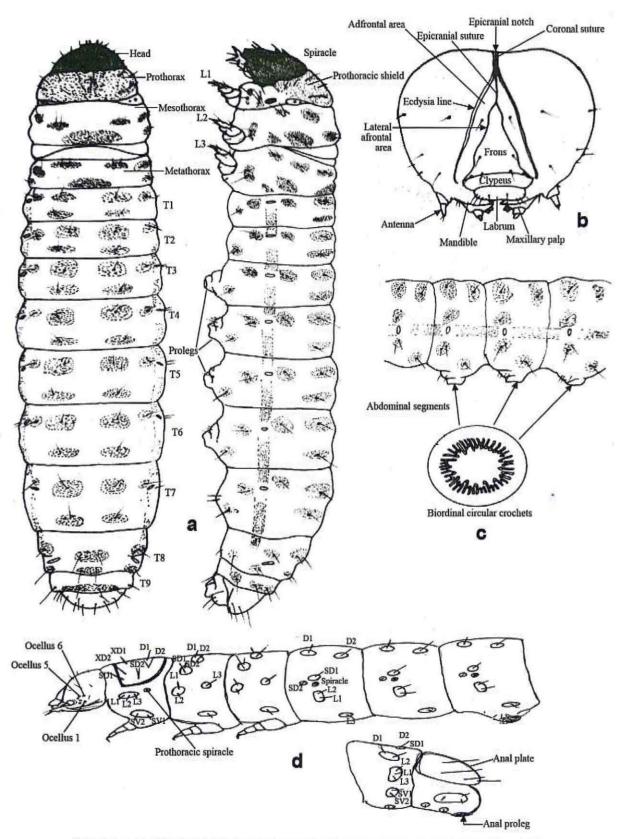


Figure 2 a-d. Dorsal and lateral (a) views of lepidopteran larva. Frontal view of head (b), abdominal prolegs and crochets (c), and larval body setae (d)

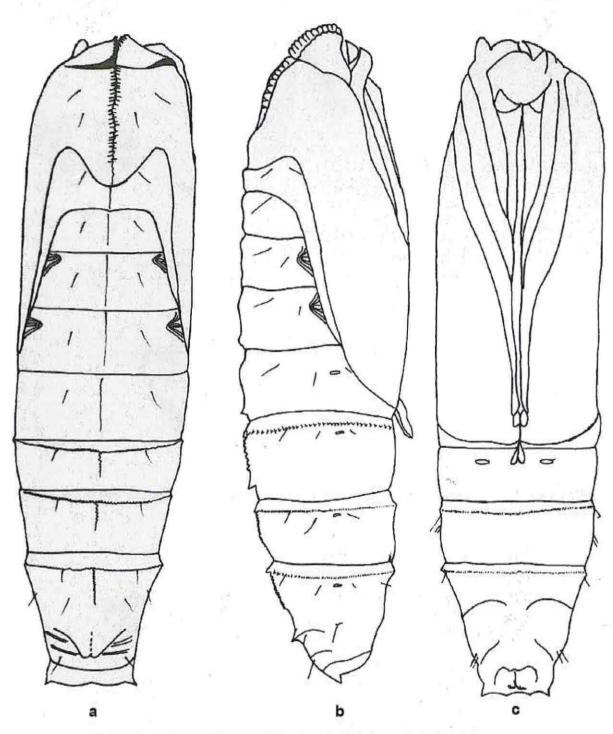


Figure 3 a-c. Dorsal (a), lateral (b) and ventral (c) views of a lepidopteran pupa

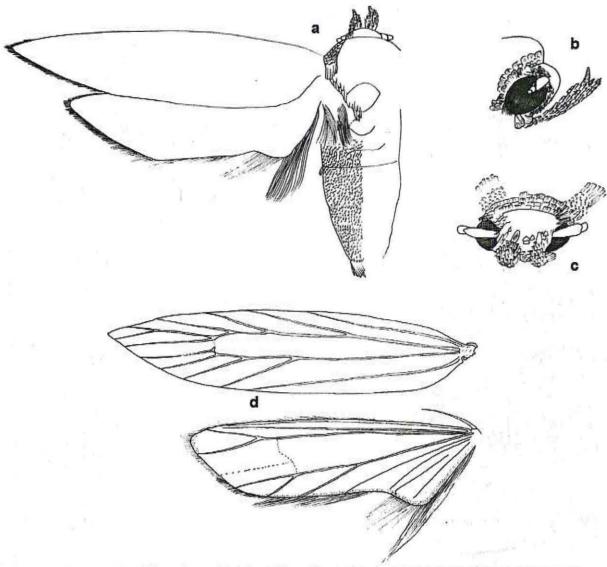


Figure 4 a-d. Adult of Stegasta sp. (a), lateral (b) and frontal (c) views of the head and wing venation (d)

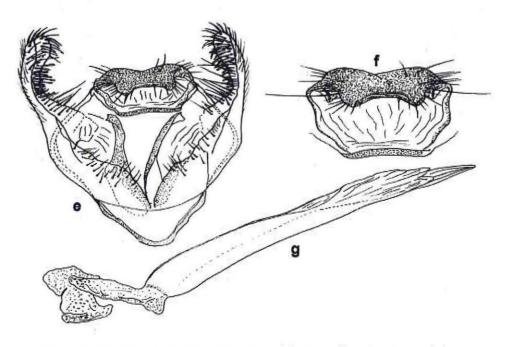


Figure 4 e-g. Male genitalia of Stegasta sp. (e), uncus (f) and aedeagus (g)

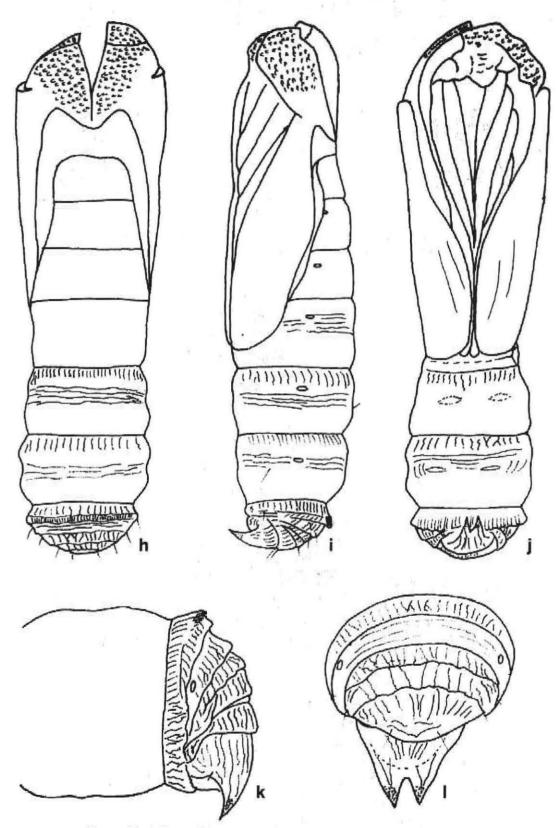


Figure 4 h–I. Pupa of Stegasta sp. in dorsal (h), lateral (i) and ventral (j) views and lateral (k) and frontal (l) views of cremaster

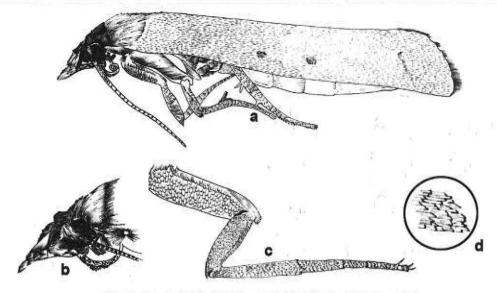


Figure 5 a-d. Adult Eldana saccharina Walker (a), head (b) and leg 1 (c) in lateral views, and closeup of hair scales (d)

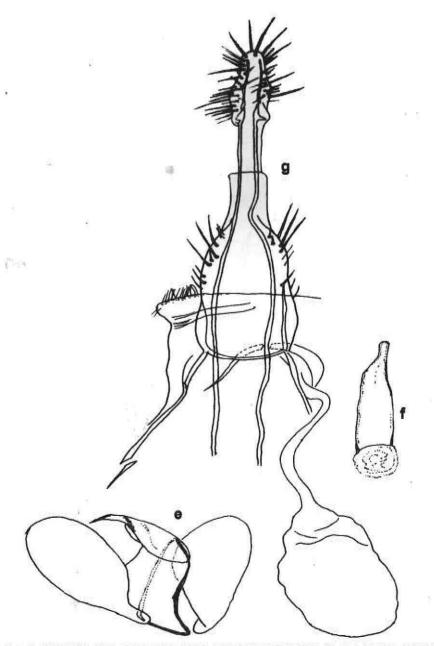


Figure 5 e-g. Genitalia of E. saccharina male (e) and its aedeagus (f) and female genitalia (g)

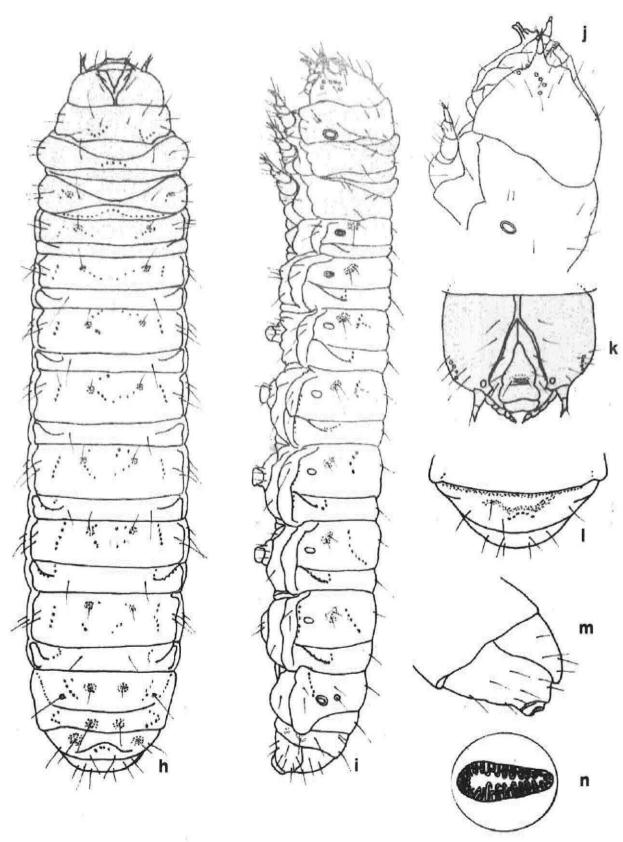


Figure 5 h-n. Dorsal (h) and lateral (i) views of *E. saccharina* larva, lateral (j) and frontal (k) views of head, dorsal (l) and lateral (m) views of anal plate and crochets (n)

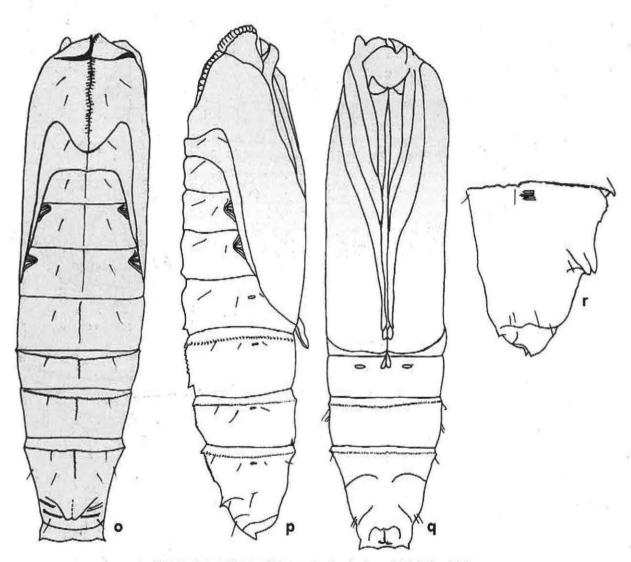


Figure 5 o-r. Pupa of *E. saccharina* in dorsal (o), lateral (p) and ventral (q) views and lateral view of cremaster and segments VIII–X (r)

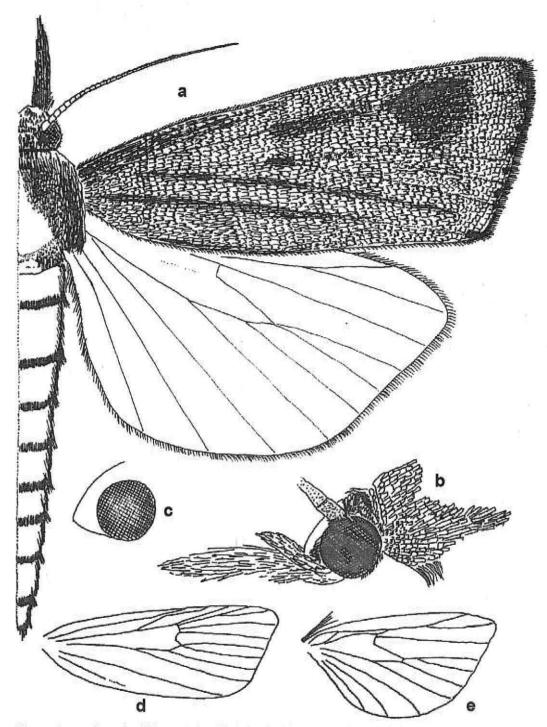


Figure 6 a-e. Female Chilo partellus (Swinhoe) with expanded wings (a), lateral views of head showing labial palp (b) and corneous point (c) and venation in the forewing (d) and hindwing (e)

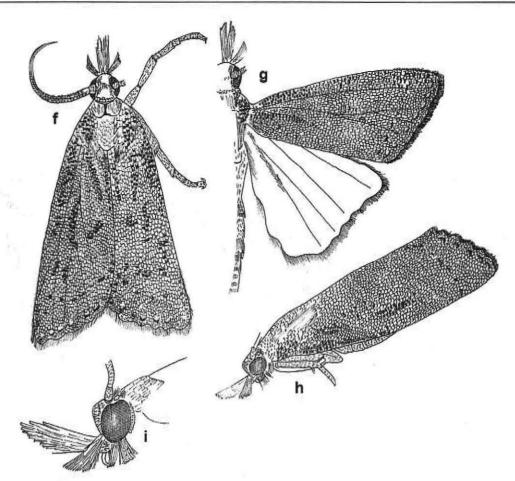


Figure 6 f-i. Male of C. partellus at rest (f), with expanded right forewing (g) lateral view (h), and head showing the ocelli, chaetosemata and labial palp (i)

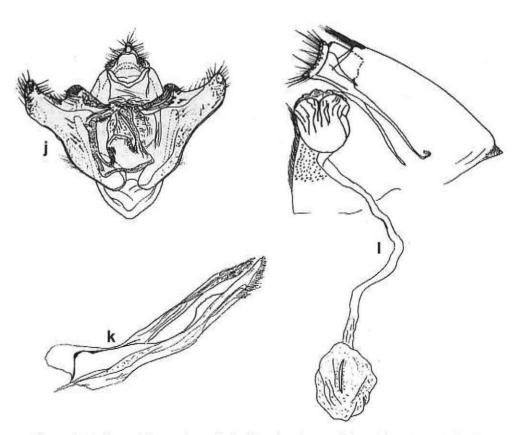
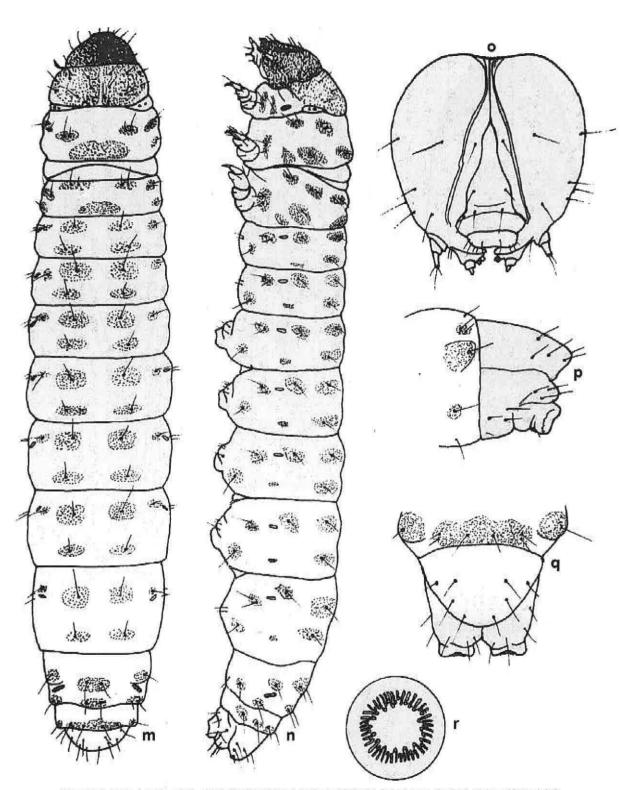


Figure 6 j-l. C. partellus male genitalia (j) and aedeagus (k), and female genitalia (l)



 $0.047164 p_{1}^{2}(4.46\pm5) = 0.04 p_{1}^{2}(5)$

Figure 6 m-r. Larva of *C. partellus* in dorsal (m) and lateral (n) views, frontal view of head (o), lateral (p) and dorsal (q) views of anal plate, and crochets (r)

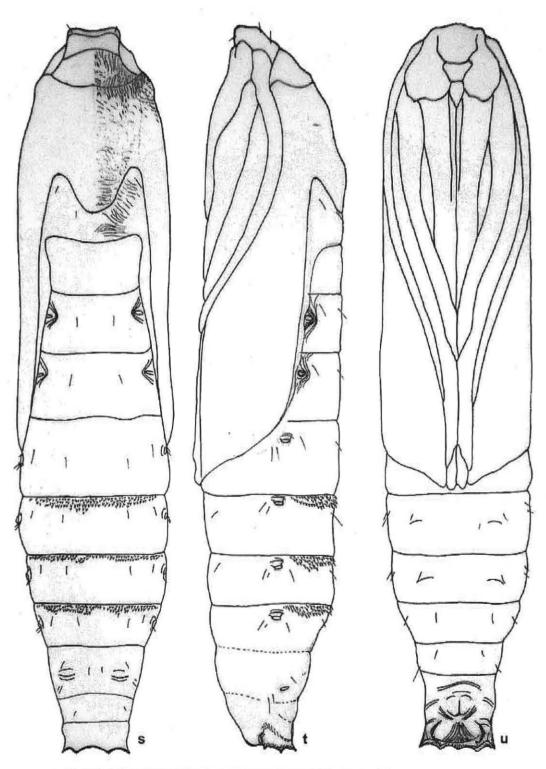


Figure 6 s-u. Dorsal (s), lateral (t) and ventral (u) views of C. partellus pupa

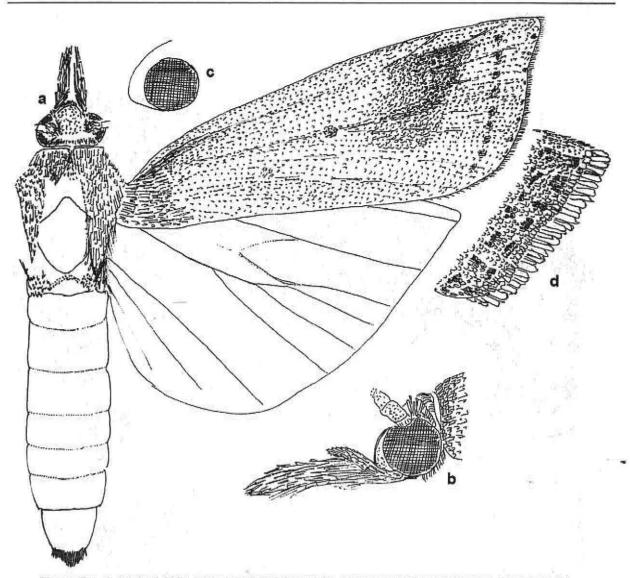


Figure 7 a-d. Adult of *Chilo orichalcociliellus* Strand (a), lateral view of head showing labial palp (b), head with rounded frons (c) and marking on the terminal margins of forewing (d)

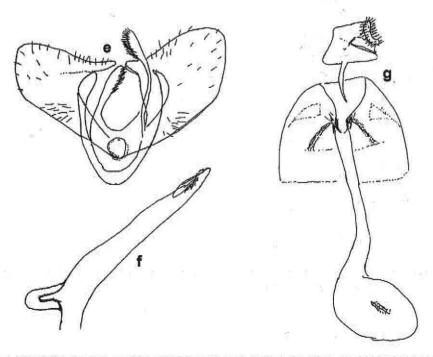


Figure 7 e-g. Genitalia of male (e) and aedeagus (f) and female genitalia (g) of Chilo orichalcociliellus

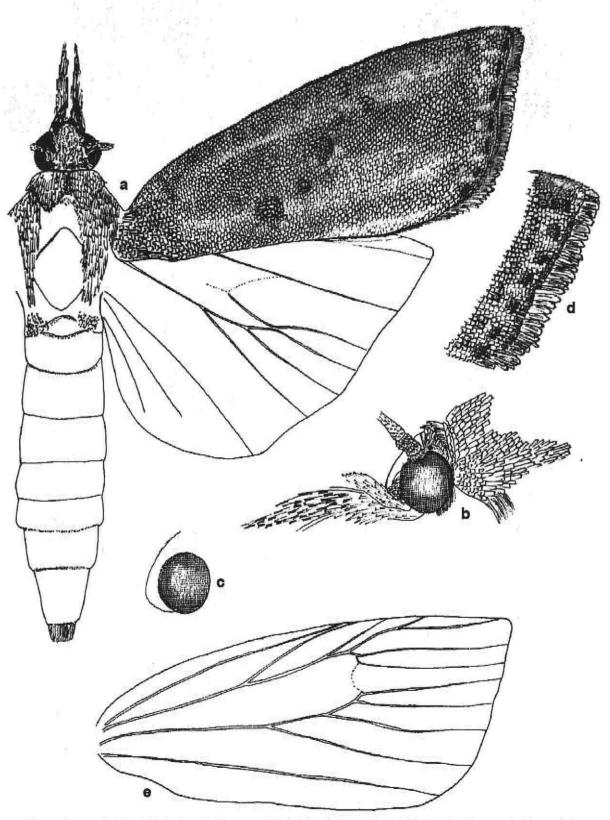


Figure 8 a-e. Adult of *Chilo thyrsis* Bleszynski (a), lateral view of head (b), head with rounded frons (c), markings on terminal margins of forewing (d) and forewing venation (e)

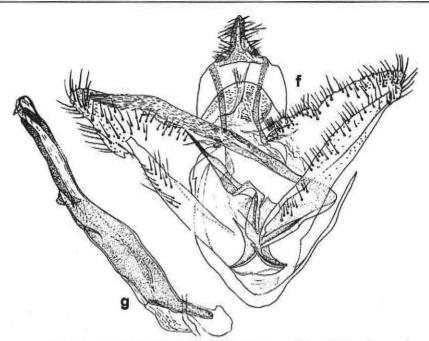


Figure 8 f-g. Male genitalia (f) and aedeagus (g) of C. thrysis

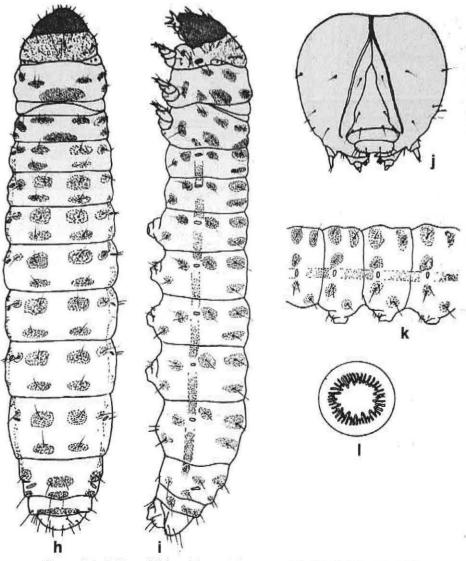


Figure 8 h–I. Dorsal (h) and lateral (i) views of *C. thrysis* larva, frontal view of head (j), abdominal prolegs (k) and crochets (l)

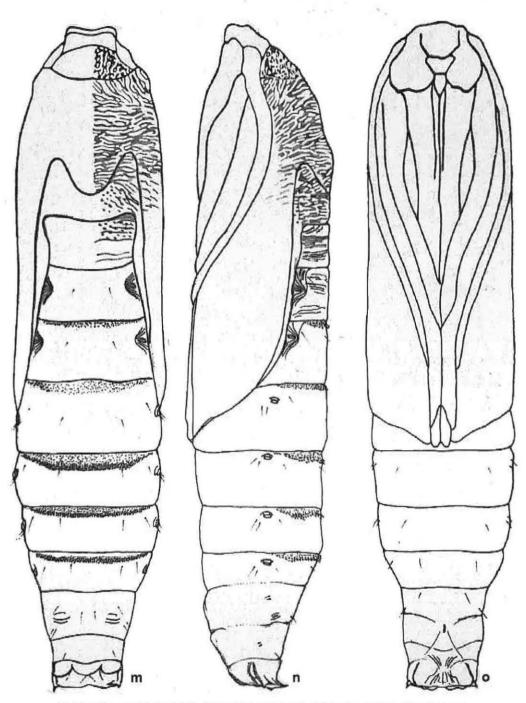


Figure 8 m-o. Dorsal (m), lateral (n) and ventral (o) views of C. thrysis pupa

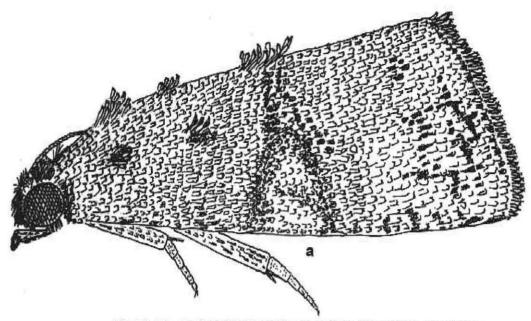


Figure 9 a. Adult of Thaumatotibia leucotreta (Meyrick) in lateral view

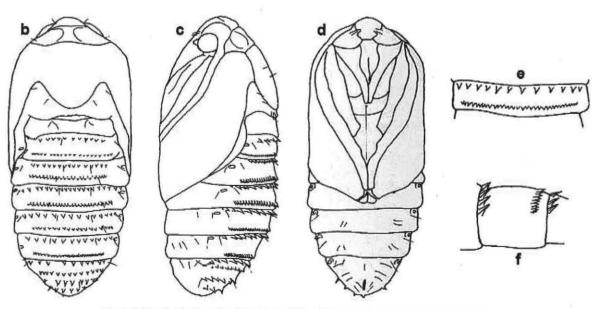


Figure 9 b–f. Pupa of *T. leucotreta* (Meyrick) in dorsal (b), lateral (c) and ventral (d) views and the abdominal spines in dorsal (e) and lateral (f) views

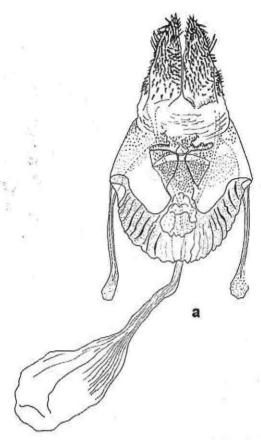


Figure 10 a. Female genitalia of Sesamia nonagrioides botanephaga

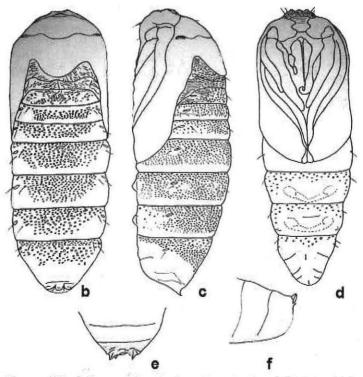


Figure 10 b-f. Pupa of S. n. botanephaga in dorsal (b), lateral (c) and ventral (d) views and cremaster in dorsal (e) and lateral (f) views

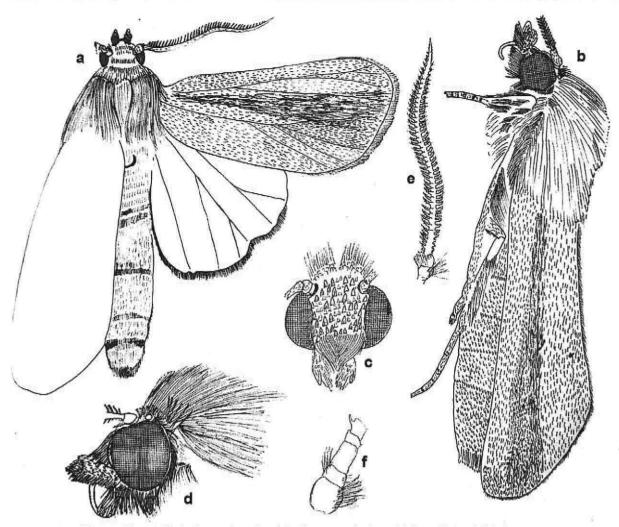


Figure 11 a-f. Male Sesamia calamistis Hampson in dorsal (a) and lateral (b) views, frontal (c) and lateral (d) views of head, antenna (e), and scape + pedicel + three funicles (f)

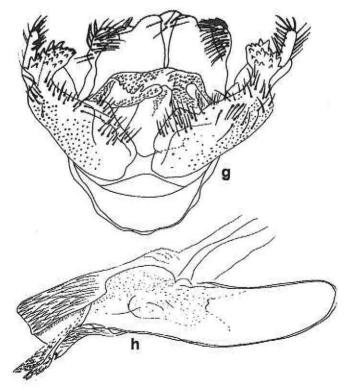


Figure 11 g-h. Male genitalia (g) and aedeagus (h) of S. calamistis Hampson

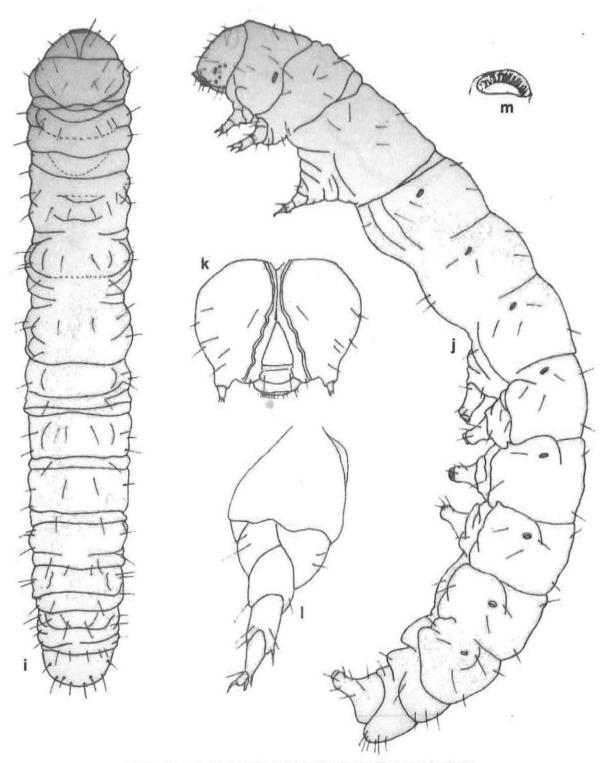


Figure 11 i-m. Larva of S. calamistis in dorsal (i), lateral (j) views, frontal view of head (k), leg I (l), and linear pattern of crochets (m)

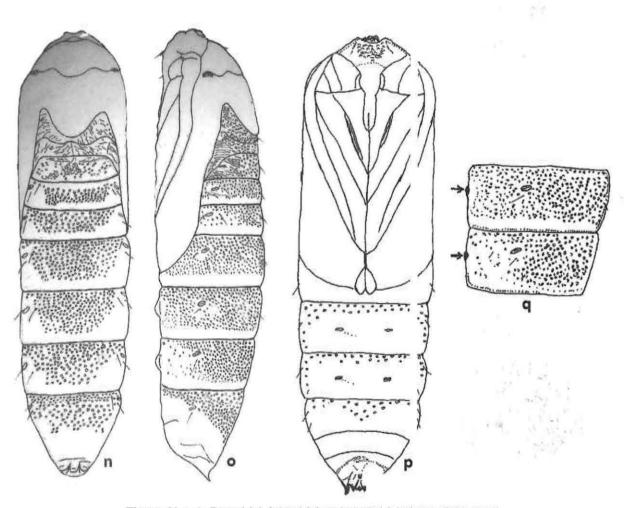


Figure 11 n-q. Dorsal (n), lateral (o) and ventral (p) views of the pupa of S. calamistis, lateral view of abdominal segments IV-V (q)

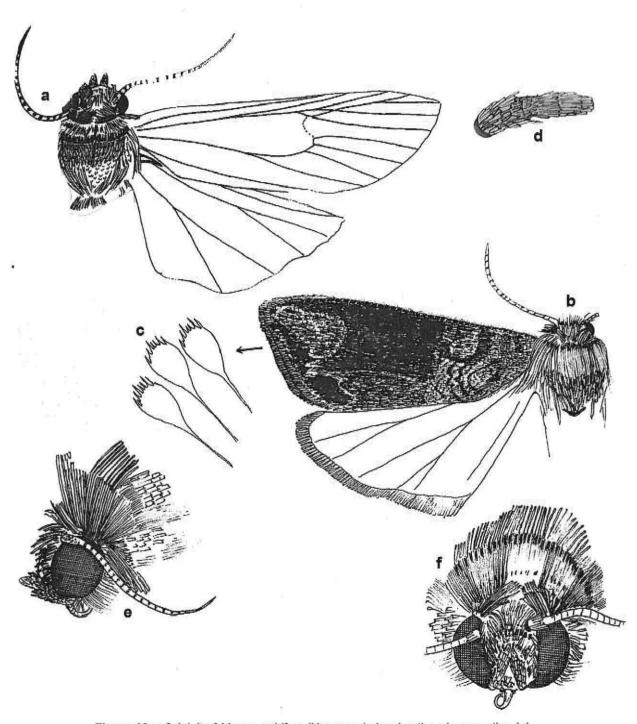


Figure 12 a–f. Adult of *Manga nubifera* (Hampson) showing the wing venation (a), wing pattern (b), hair scales on terminal fringes of wing (c), hairs on scape and pedicel of antenna (d), and lateral (e) and frontal (f) views of head

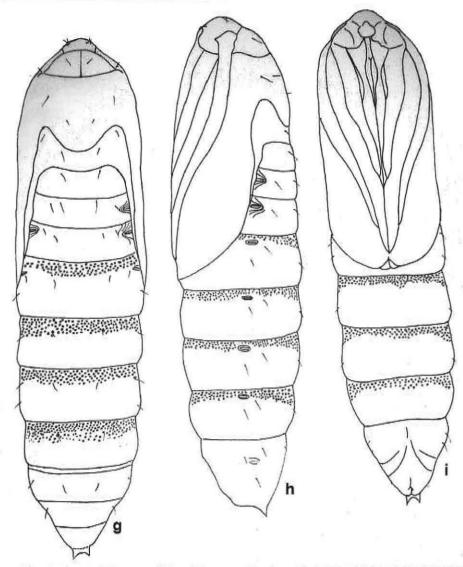


Figure 12 g-i. Pupa of Manga nubifera (Hampson) in dorsal (g), lateral (h) and ventral (i) views

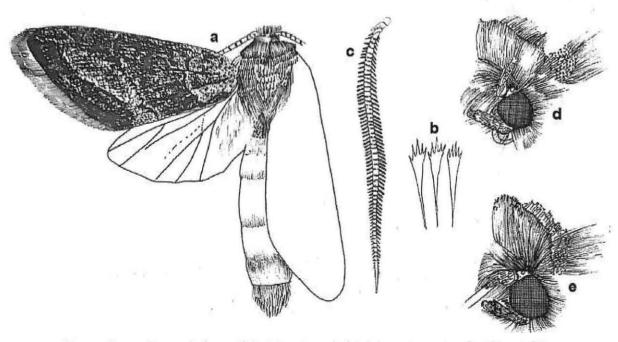


Figure 13 a-e. Busseola fusca (Fuller) female moth (a), hair scales on terminal fringes (b), male antenna (c), and lateral view of male (d) and female (e) heads

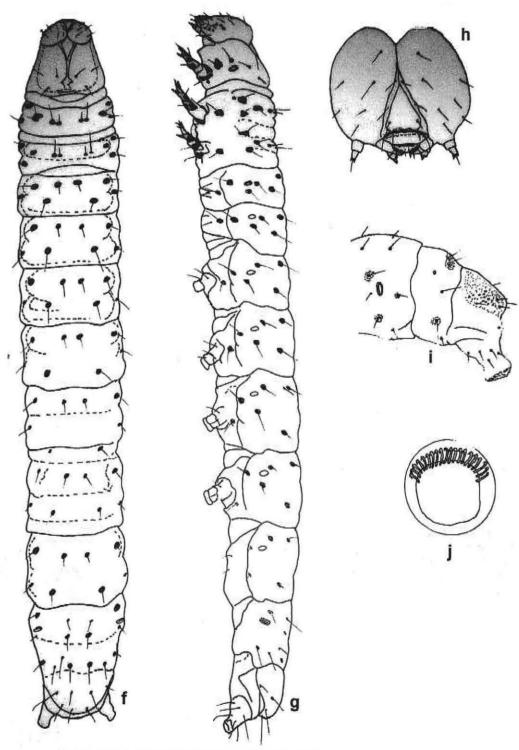


Figure 13 f-j. Dorsal (f) and lateral (g) views of B. fusca larva, frontal view of head (h), lateral view of anal plate and proleg (i) and the linear crochets (j)

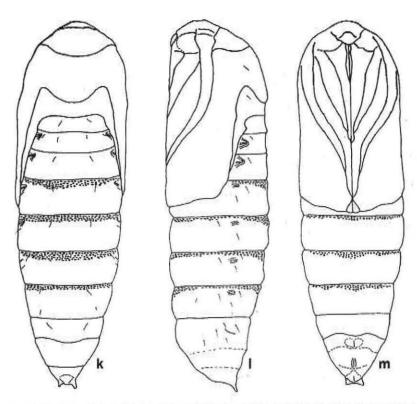


Figure 13 k-m. Dorsal (k), lateral (l) and ventral (m) views of B. fusca pupa

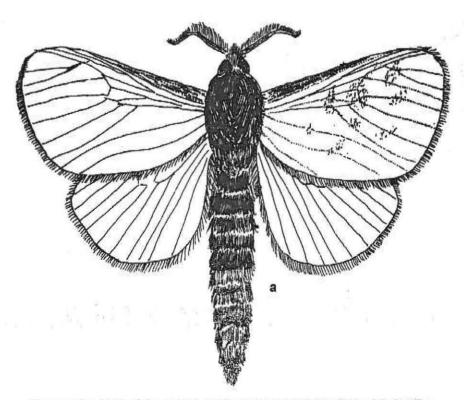


Figure 14 a. Moth of the cossid, Phragmataecia boisduvalii Herrich-Shäffer

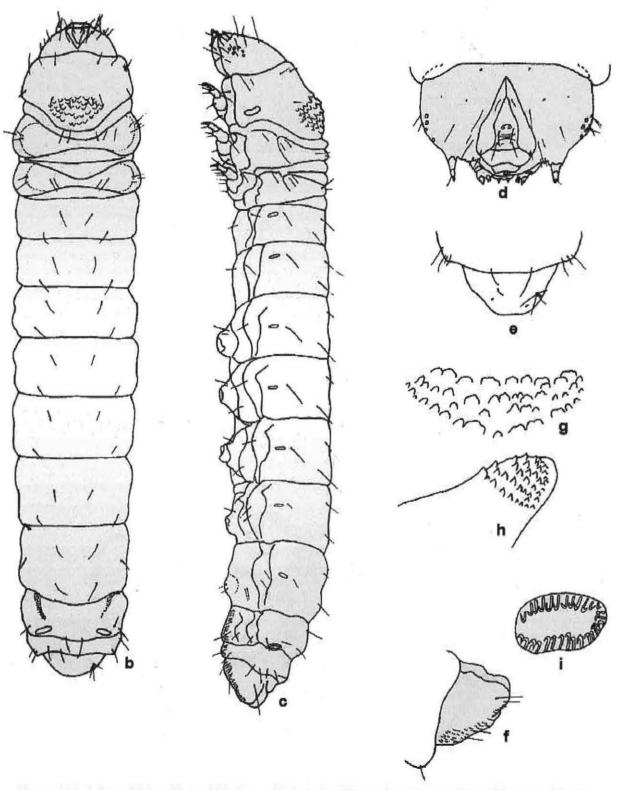


Figure 14 b-i. Dorsal (b) and lateral (c) views of *P. boisduvalii* larva, frontal view of head (d), dorsal (e) and lateral (f) views of anal plate, blunt spikes on the pronotum in dorsal (g) and lateral (h) views and crochets (i)

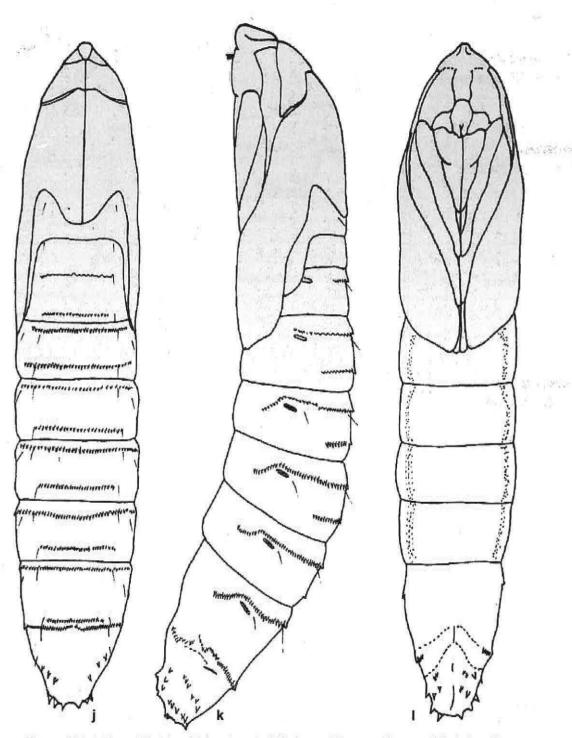


Figure 14 j-I. Dorsal (j), lateral (k) and ventral (l) views of the cossid pupa, P. boisduvalli

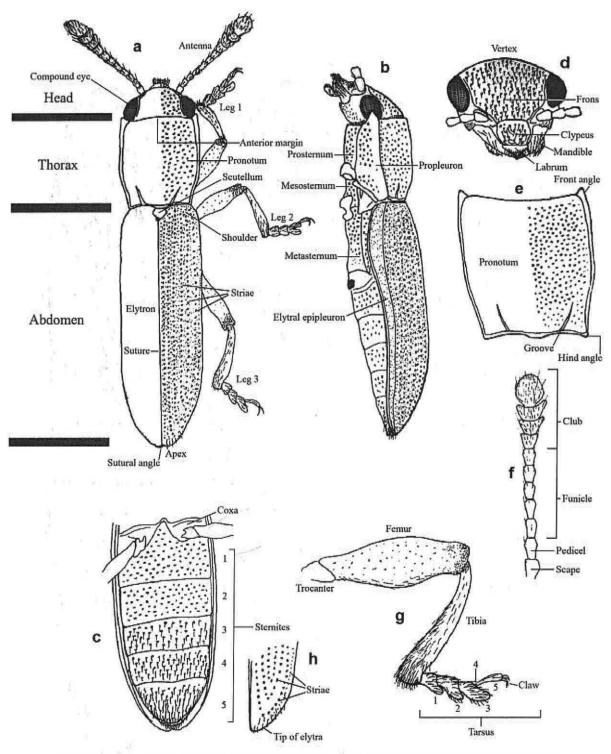


Figure 15 a-h. Morphology of a beetle showing the dorsal (a) and lateral (b) views, abdominal venter (c), frontal view of head (d), pronotum (e), antenna (f), leg III (g) and tip of elytra (h)

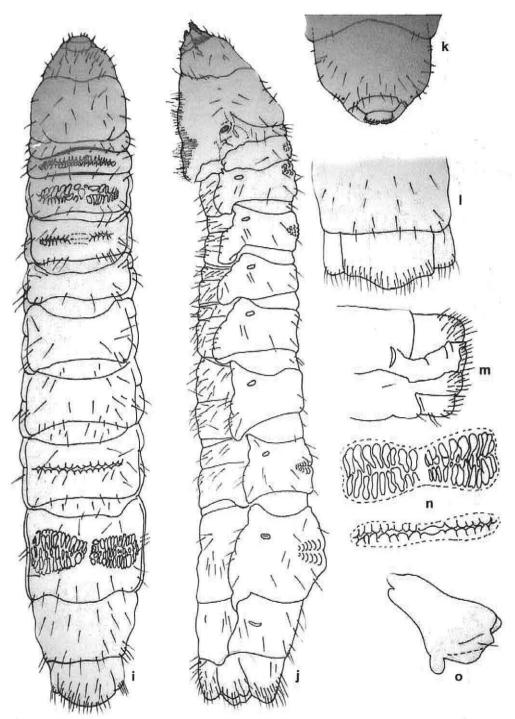


Figure 15 i–o. Dorsal (i) and lateral (j) views of coleopteran larva, head (k), dorsal (l) and lateral (m) views of anal plate, two forms of ampullae (n) and mandible (o)

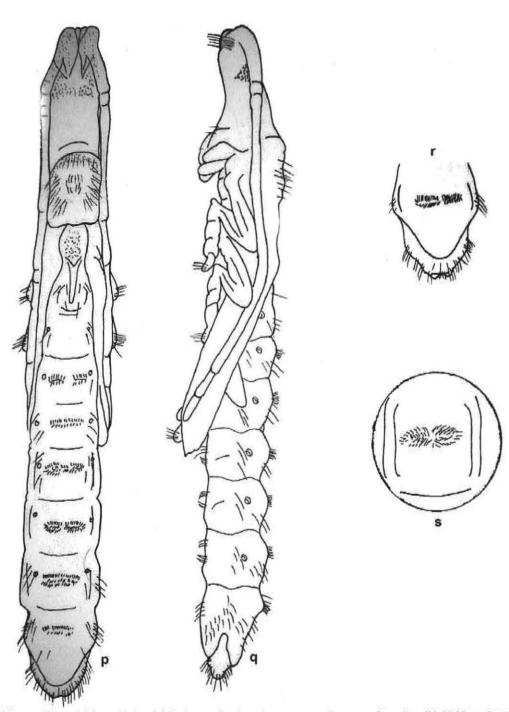


Figure 15 p-s. Dorsal (p) and lateral (q) views of coleopteran pupa, dorsum of tergites IX-X (r) and ampulla (s)

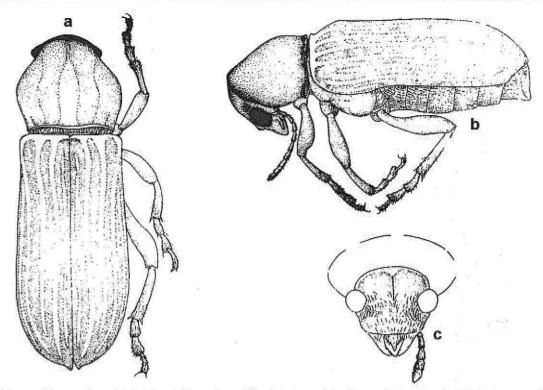


Figure 16 a-c. Dorsal (a), lateral (b) and head frontal views (c) of the anthribid beetle, Phloebius sp. A

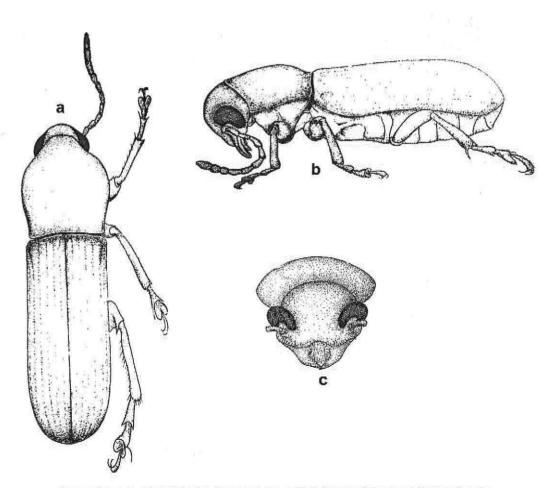


Figure 17 a-c. Phloebius sp. B dorsal (a), lateral (b) and frontal (c) views of head

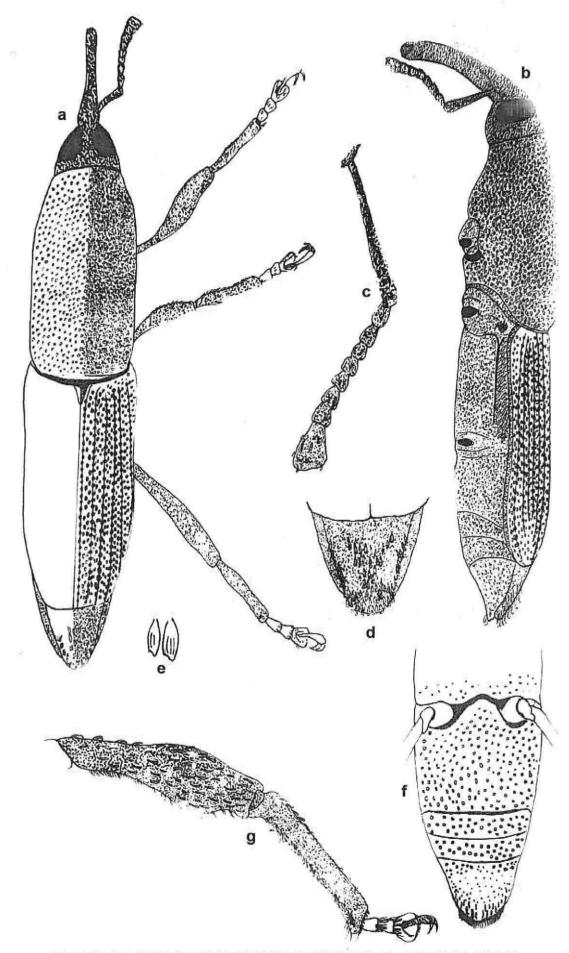


Figure 18 a-g. Dorsal (a) and lateral (b) views of ?Odioporus sp., antenna (c), hairs on the exposed abdominal tergite (d), closeup of hairs (e), abdominal venter (f) and leg I (g)

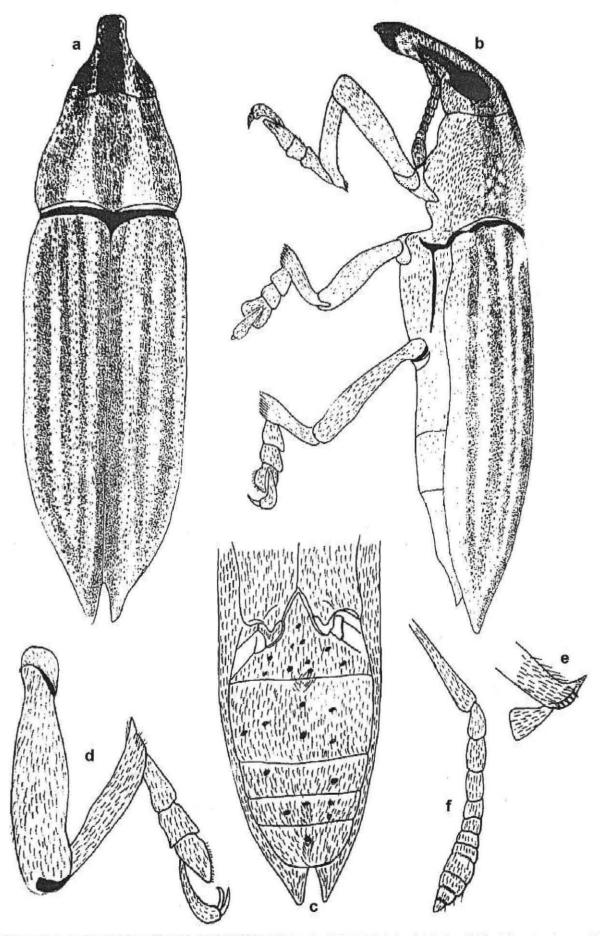


Figure 19 a-f. Dorsal (a) and lateral (b) views of Lixus sp. A, sternites (c), leg I (d), tip of tibia (e) and antenna (f)

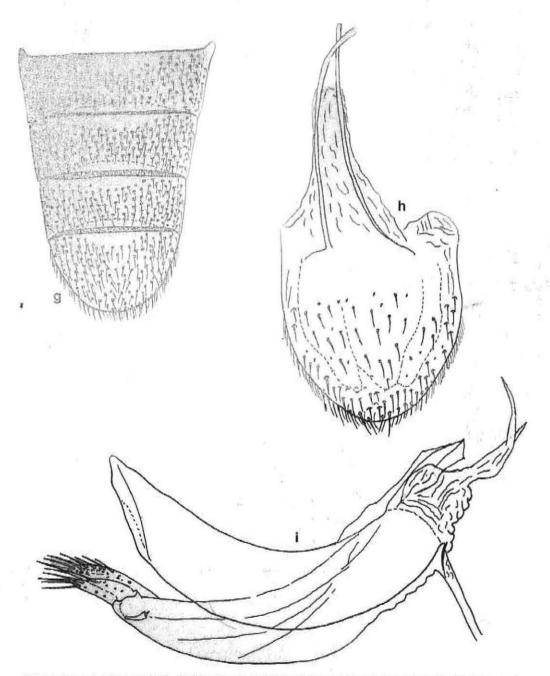


Figure 19 g-i. Sternites (g), abdominal sternite IX of male (h) and aedeagus (i) of Lixus sp. A

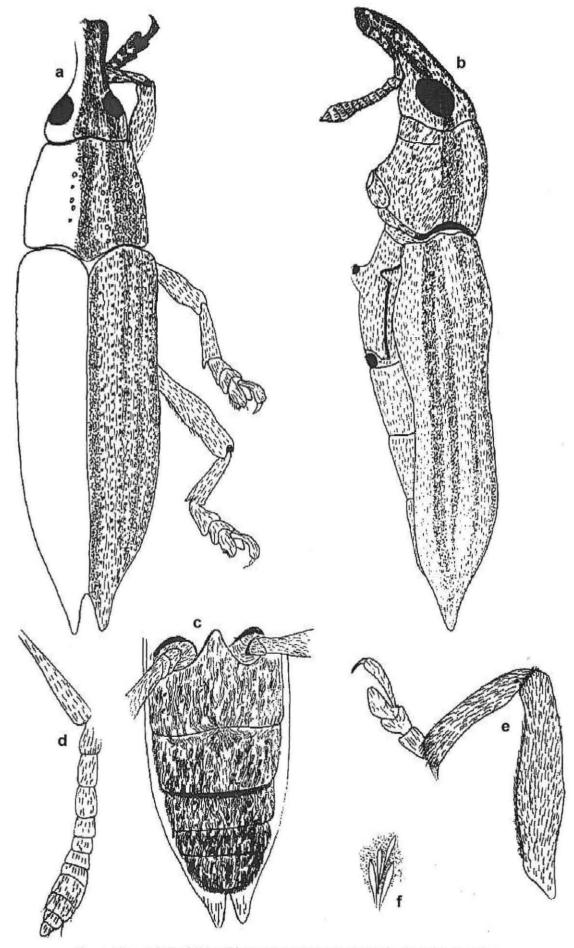


Figure 20 a–f. Dorsal (a) and lateral (b) views of *Lixus* near *germaini* Hustache, abdominal sternites (c), antenna (d) leg I (e) and hairs on abdominal venter (f)

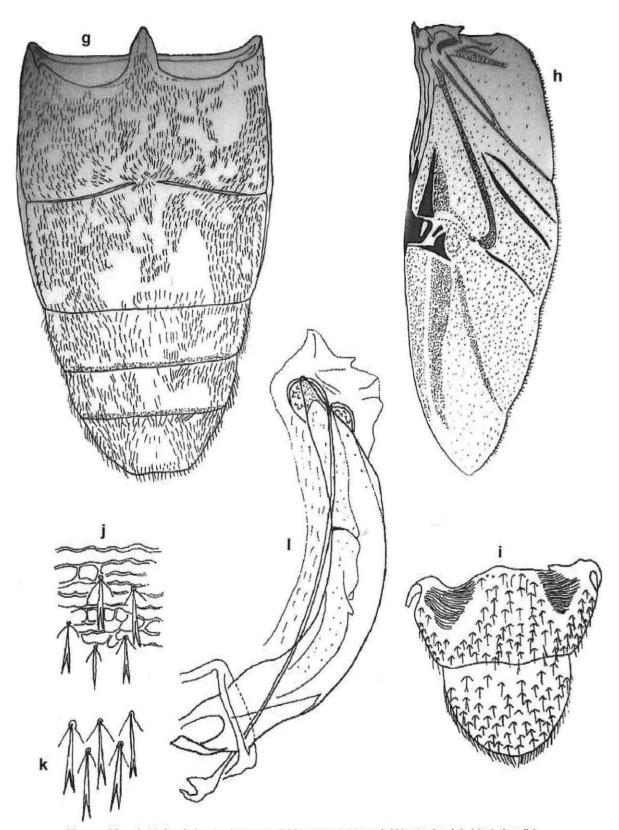


Figure 20 g–I. Male abdominal venter of *Lixus* near *germaini* Hustache (g), hindwing (h), sternites VIII–IX (i), sculpture (j) and hairs (k) in terminal sternite, and aedeagus (l)

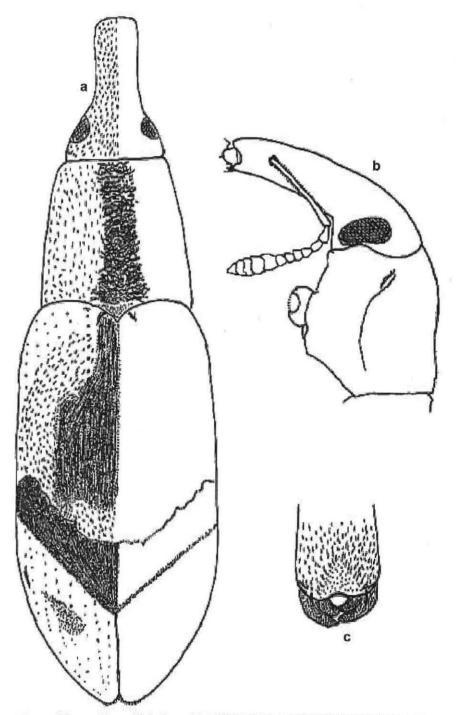


Figure 21 a-c. Dorsal (a) view of adult *Tanymecus* near *dilaticollis* Gyllenhal, lateral view of head (b) and mandible (c)

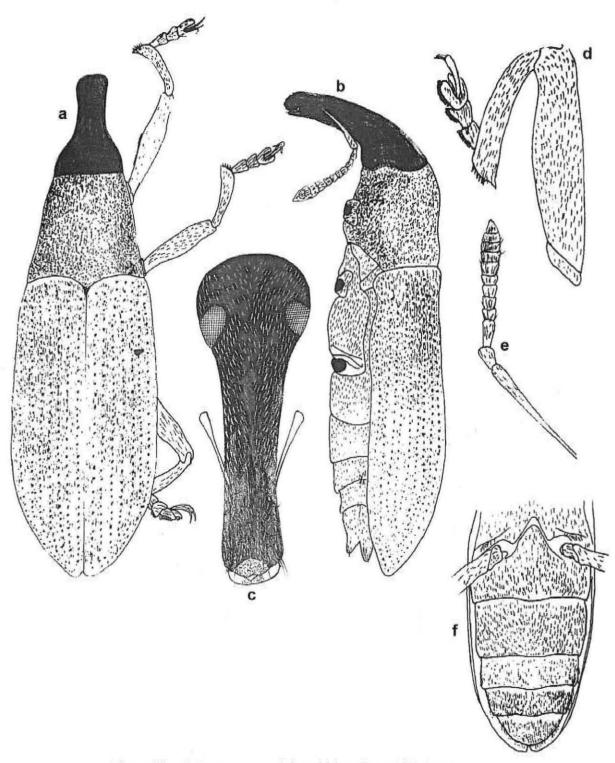


Figure 22 a-f. Tanymecus sp. A dorsal (a) and lateral (b) views, rostrum and head (c), leg I (d), antenna (e) and abdominal sternite (f)

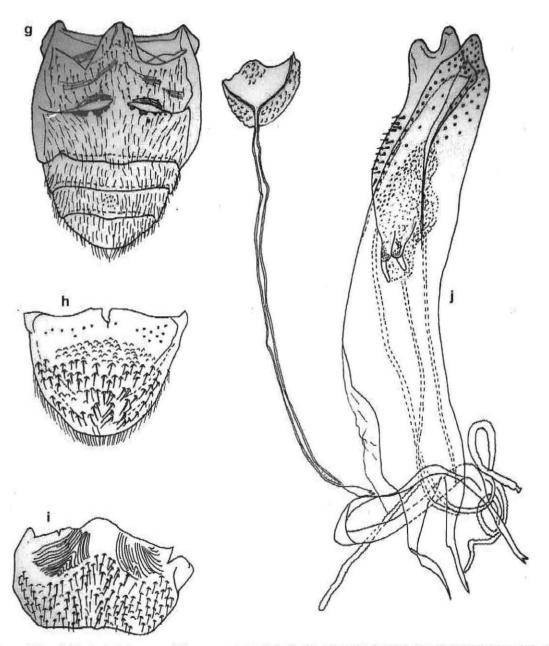


Figure 22 g-j. Ventral abdomen of Tanymecus sp. A male (g), sternites VIII (h) and IX (i) and aedeagus (j)

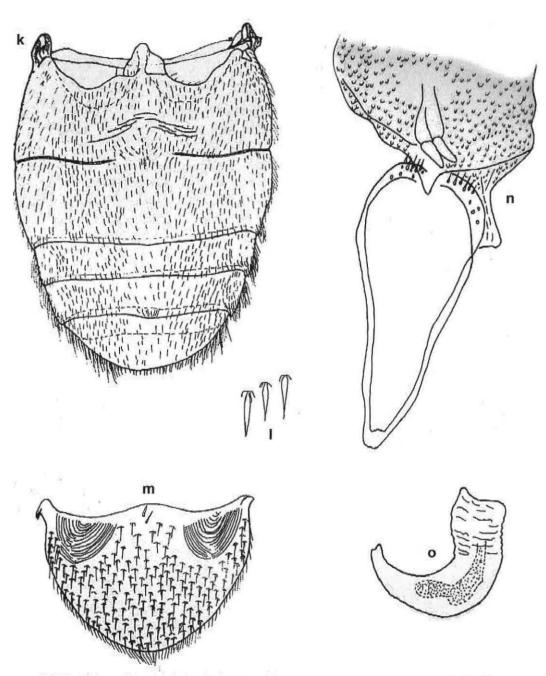


Figure 22 k-o. Female abdominal venter (k) of *Tanymecus* sp. A, abdominal hairs (l), sternite VIII (m), part of female genitalia (n) and spermatheca (o)

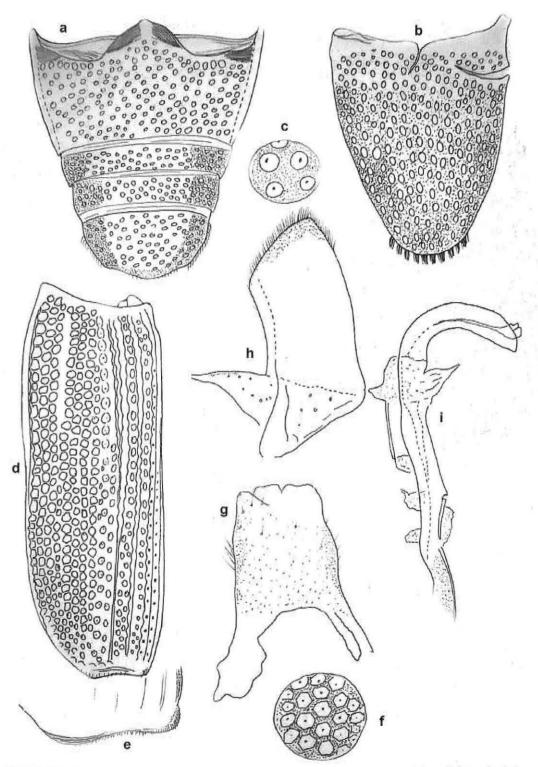


Figure 23 a-i. Abdominal venter (a) and dorsum (b) of ? Tanymecus sp., sculpture in sternite (c), left elytron (d) and its tip (e), sculpture of elytron (f), portions of male genitalia (g, h) and aedeagus (i)

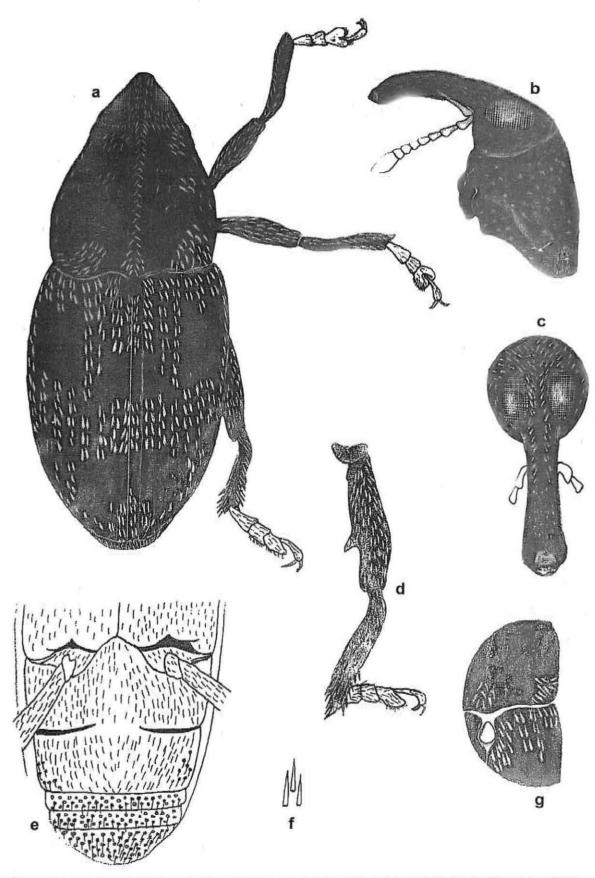


Figure 24 a-g. Dorsal (a) view of unknown genus sp. A, lateral (b) and frontal (c) views of head, leg III (d), abdominal venter (e) and its hairs (f) and the junction of posterolateral pronotum and shoulder (g)

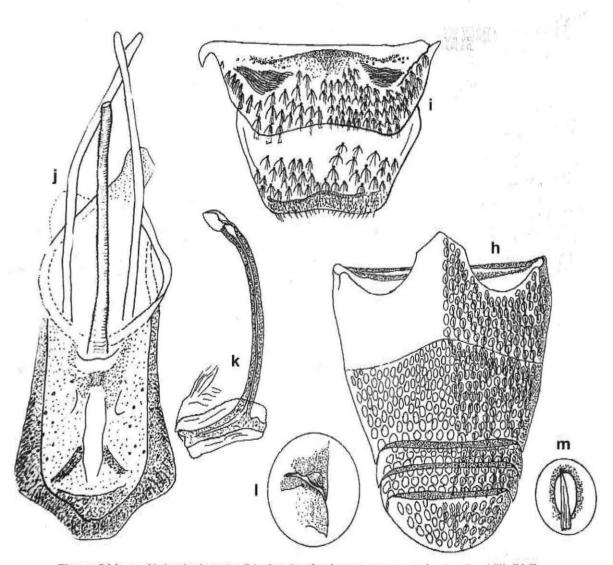


Figure 24 h-m. Abdominal venter (h) of male of unknown genus sp. A, sternites VIII-IX (i), male genitalia (j), siphon (k), lateral junction of two sternites (I) and hair in each puncture (m)

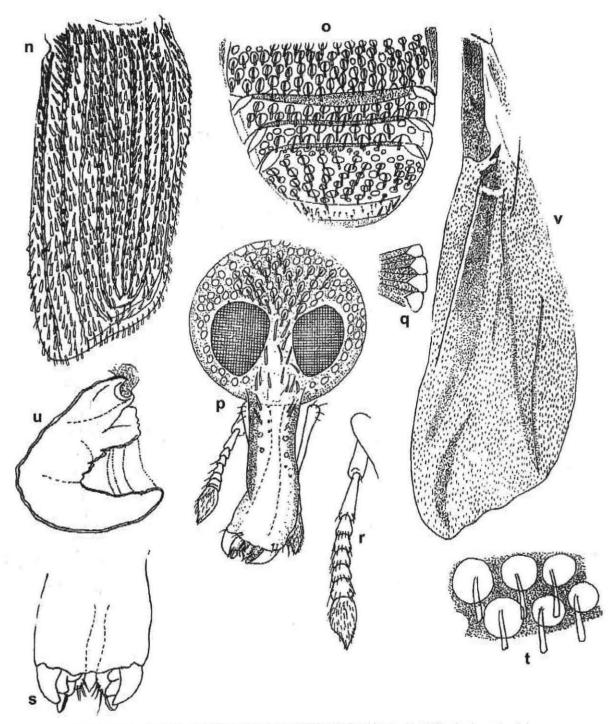


Figure 24 n-v. Female right elytron (n) of unknown genus sp. A, abdominal venter (o), frontal view of head (p), ommatidia (q), antenna (r), tip of rostrum showing the mandible (s), hairs in abdominal punctures (t), spermatheca (u) and hindwing (v)

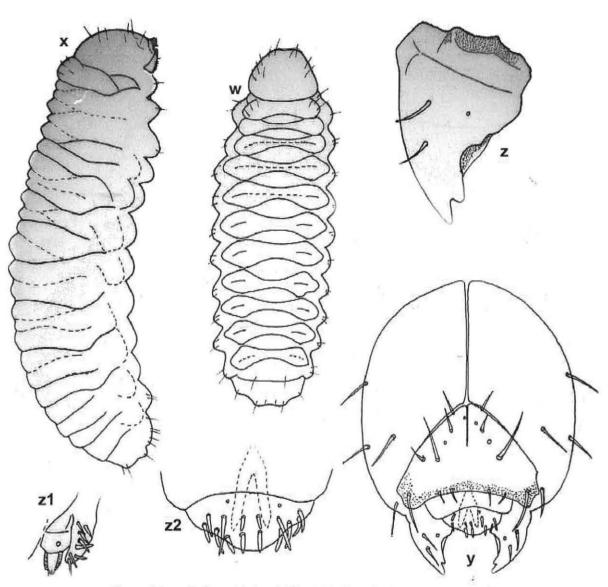


Figure 24 w-z2. Dorsal (w) and lateral (x) view of unknown genus sp. A larva, head (y), mandible (z), maxilla (z1) and setae on epipharynx (z2)

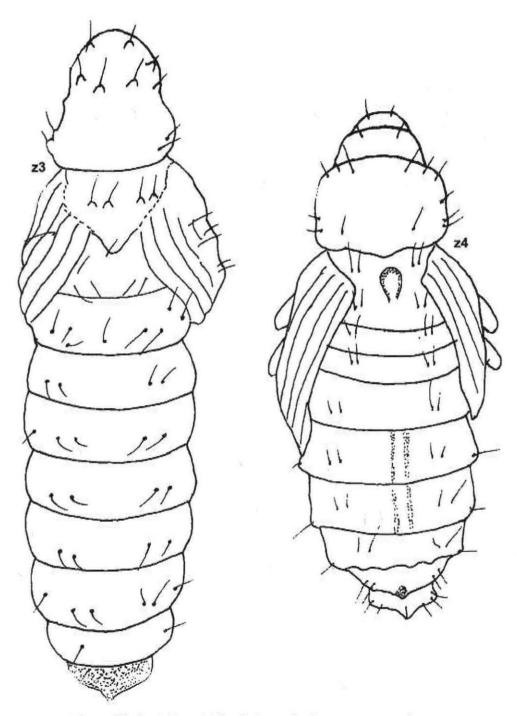


Figure 24 z3-z4. Dorsal (z3, z4) views of unknown genus sp. A pupa

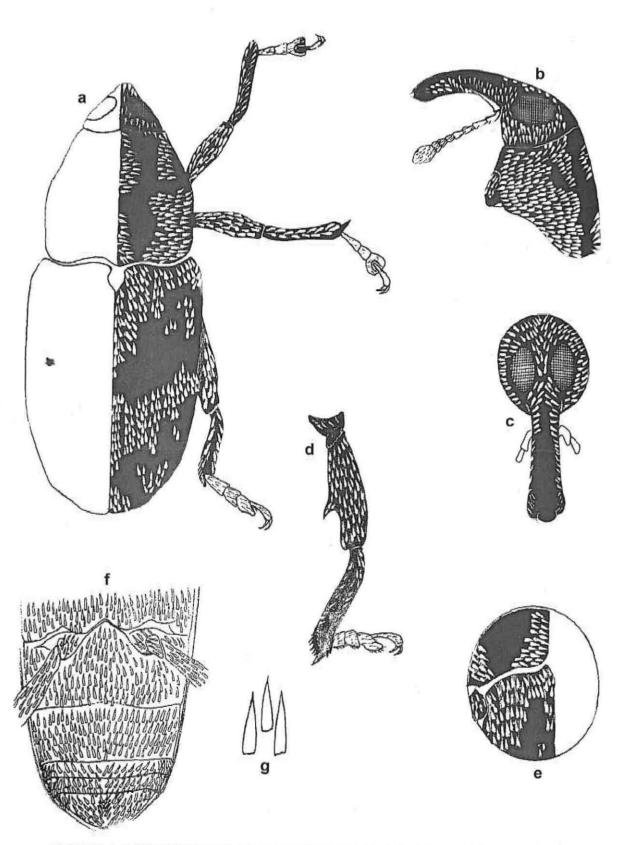


Figure 25 a-g. Dorsal (a) view of unknown genus sp. B, lateral (b) and frontal (c) views of head, leg III (d), pronotal and humeral hairs (e), abdominal venter (f) and abdominal hairs (g)

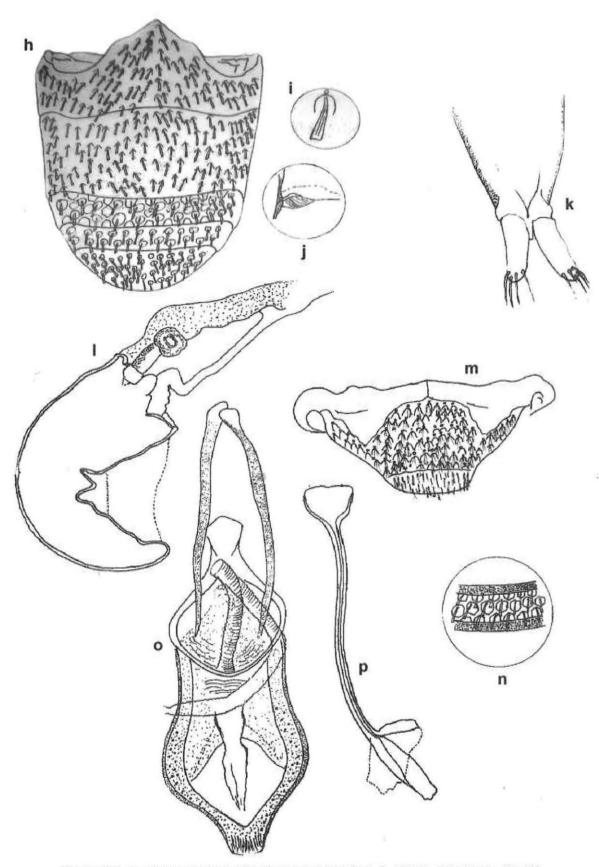


Figure 25 h–p. Abdominal venter (h) of unknown genus sp. B, hair on abdominal venter (i), separation of two sternites (j), female genitalia (k), spermatheca (I), male sternite VIII (m), sculpture and hairs on sternite V in female (n), male genitalia (o) and siphon (p)

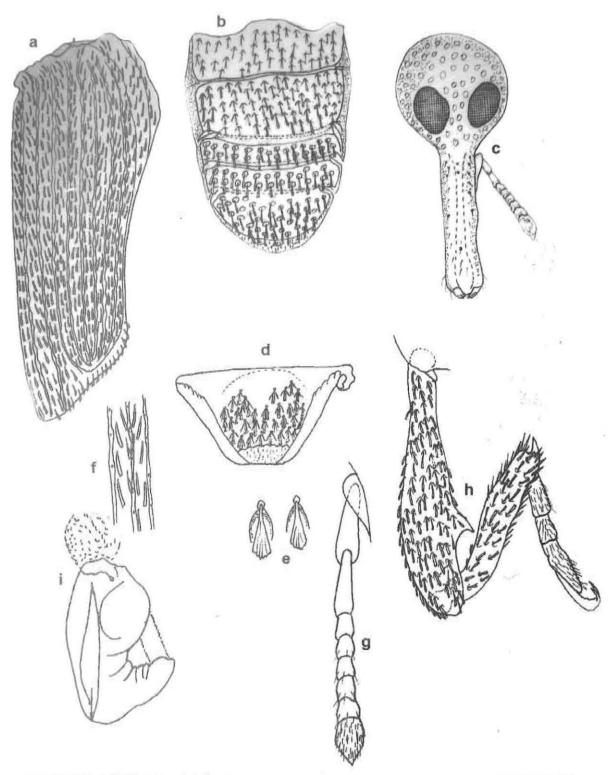


Figure 26 a-I. Right elytron (a) of unknown genus sp. C, abdominal venter (b), frontal view of head (c), last sternite (d), hairs on sternites (e), elytral hairs (f), antenna (g), leg III (h) and spermatheca (i)

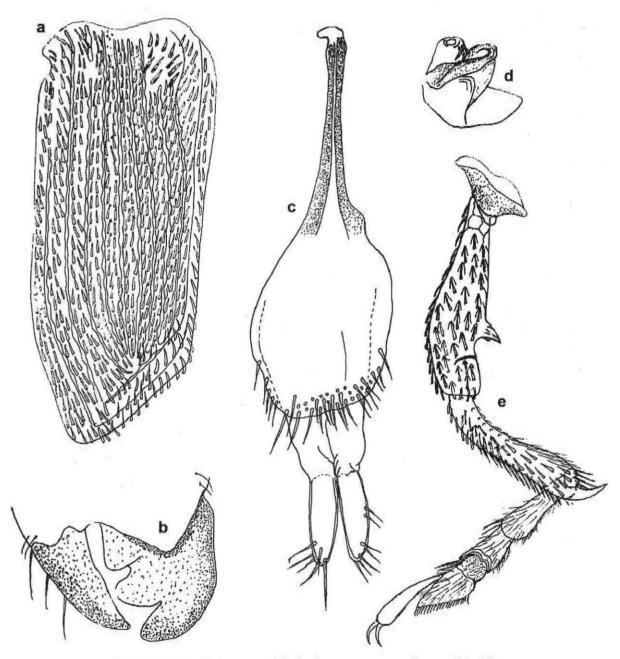


Figure 27 a-e. Right elytron (a) of unknown genus sp. D, mandible (b), portion of female genitalia (c), spermatheca (d) and leg III (e)

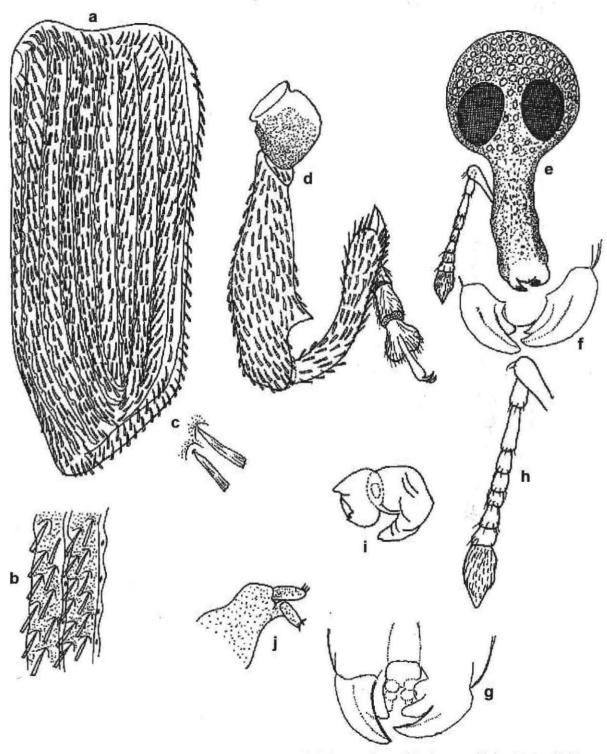


Figure 28 a-j. Right elytron (a) of unknown genus sp. E, hairs on elytron (b), closeup of hairs (c), leg III (d), frontal view of head (e), two types of mandibles (f, g), antenna (h), spermatheca (i) and part of female genitalia (j)

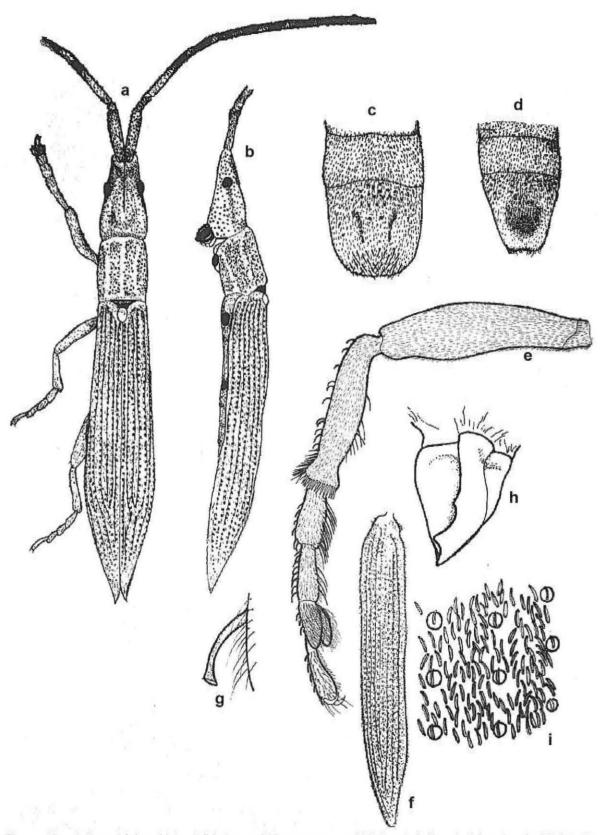


Figure 29 a-i. Dorsal (a) and lateral (b) views of *Hypamazso pauli* (Fairmaire), female (c) and male (d) tips of abdomen, leg III (e), left elytron (f), sensory hair on the leg (g), mandible (h) and pattern of elytral hairs (i)

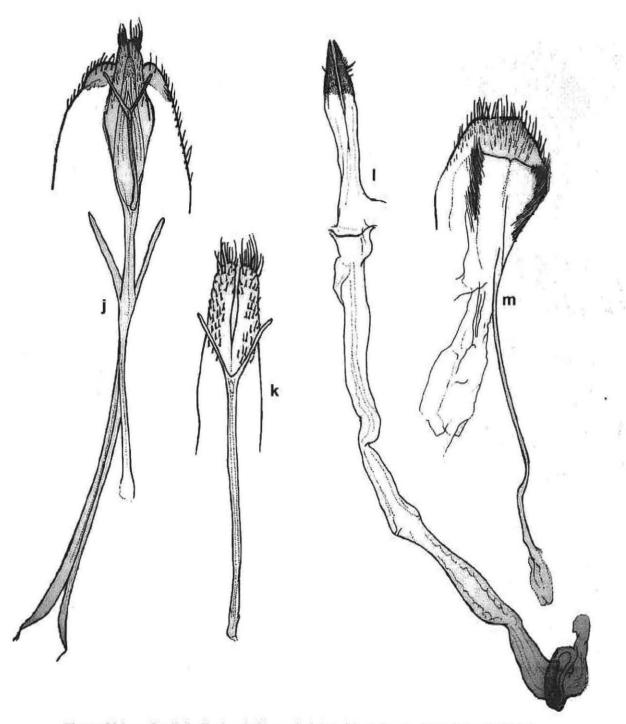


Figure 29 j-m. Genitalia (j) of male H. pauli, siphon (k), aedeagus (l) and sternite IX (m)

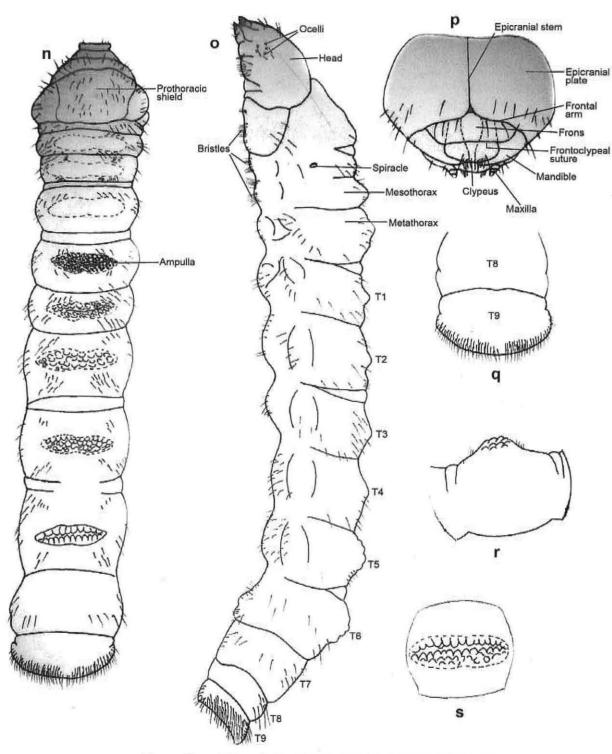


Figure 29 n-s. Dorsal (n) and lateral (o) views of *H. pauli* larva, frontal view of head (p), anal segment (q) lateral (r) and dorsal (s) views of ampulla

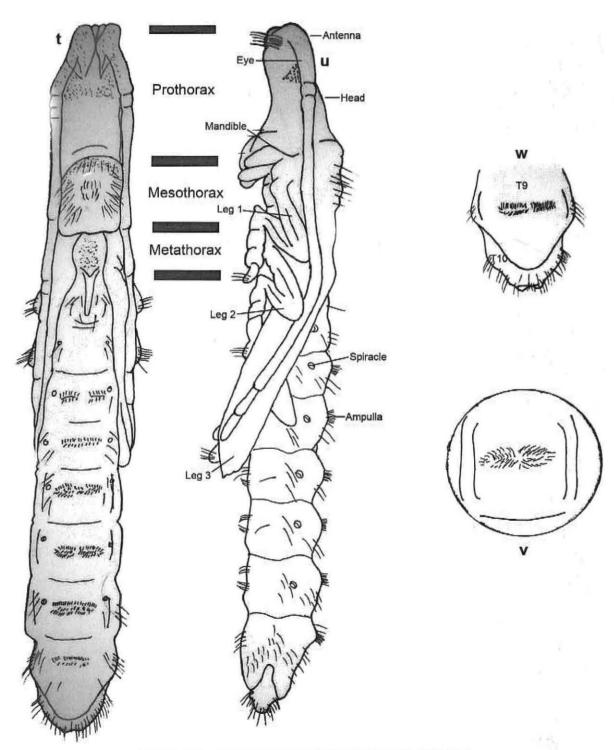


Figure 29 t-w. Dorsal (t) and lateral (u) views of *H. pauli* pupa, ampulla on anal plate (v) and on abdominal dorsum (w)

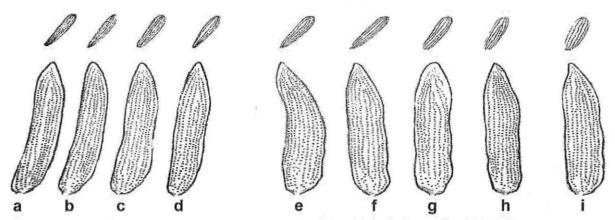


Figure 30 a-i. Wing patterns and form of elytral hairs in the adults of H. pauli

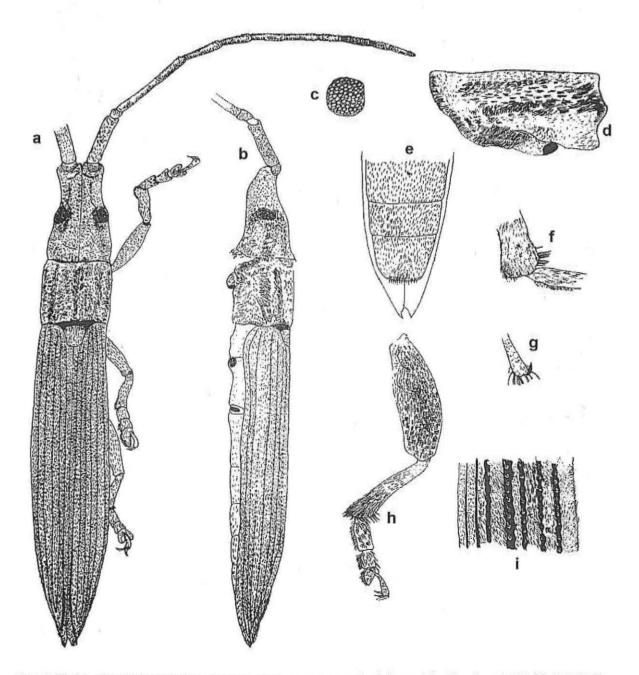


Figure 31 a-i. Dorsal (a) and lateral (b) views of *Hypamazso* sp. B adult, eye (c), side of pronotum (d), last three sternites of female (e), apex of tibia III (f), tip of claw (g), leg III (h) and sculpture on the dorso-lateral side of elytra (i)

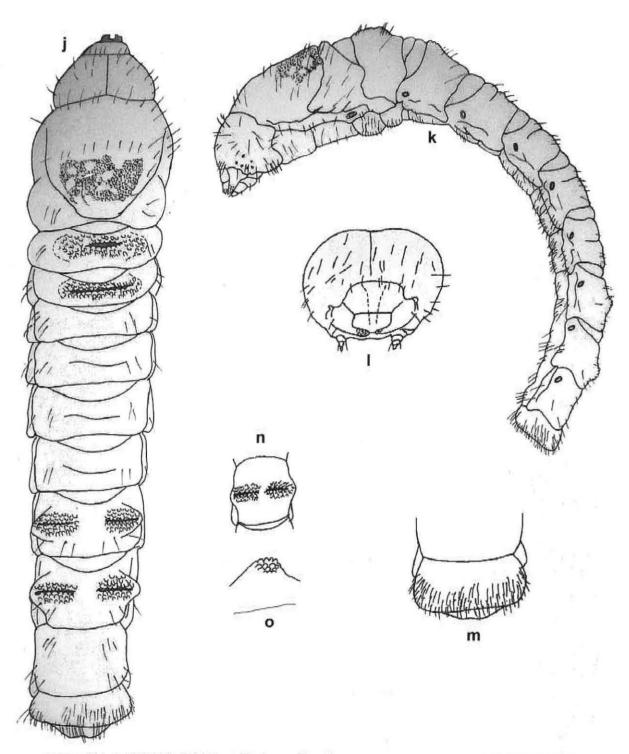


Figure 31 j-o. Dorsal (j) and lateral (k) views of the *Hypamazso* sp. B larva, frontal view of head (l), anal plate (m), paired ampulla on abdominal dorsum (n) and lateral (o) view of ampulla

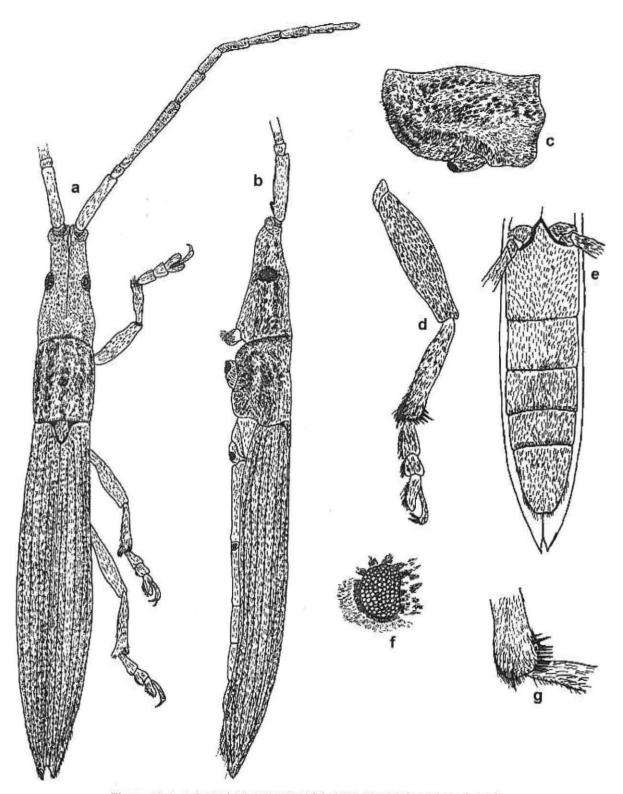


Figure 32 a-g. Dorsal (a) and lateral (b) views of *Hypamazso* sp. C adult, propleuron (c), leg III (d), abdominal venter (e), eye (f) and tip of tibia III (g)

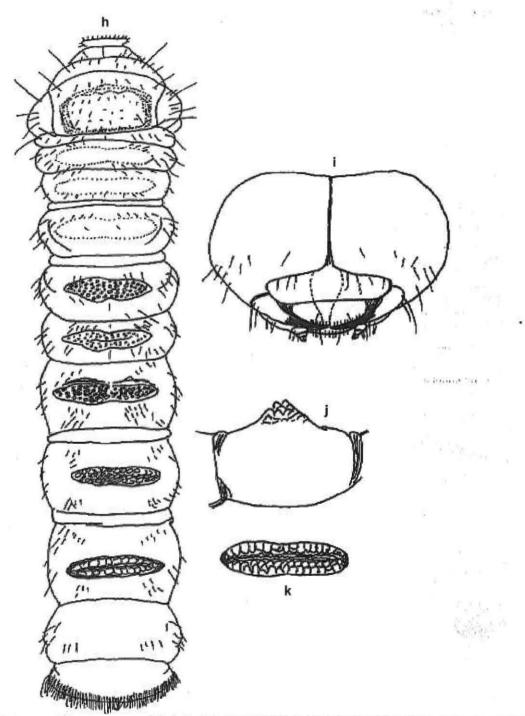


Figure 32 h-k. Larva of Hypamazso sp. C (h), frontal view of head (i), lateral (j) and dorsal (k) views of ampulla

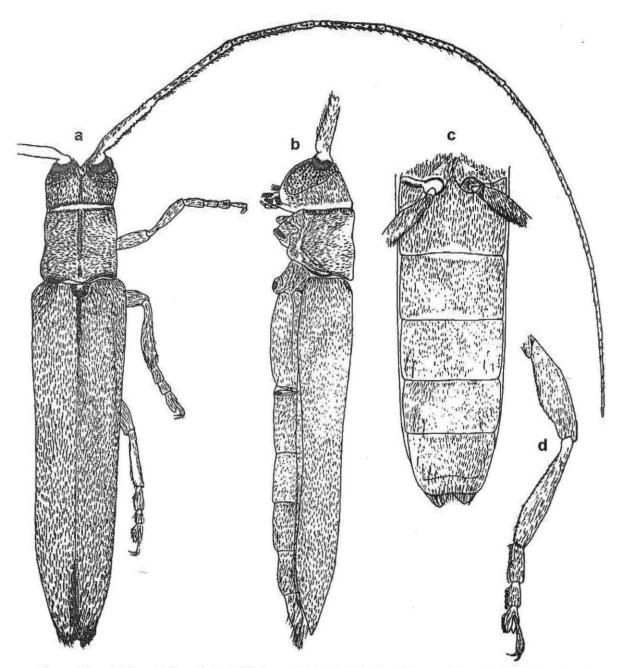


Figure 33 a-d. Dorsal (a) and lateral (b) views of Obeneopsis sp. adult, sternites I-V (c) and leg III (d)

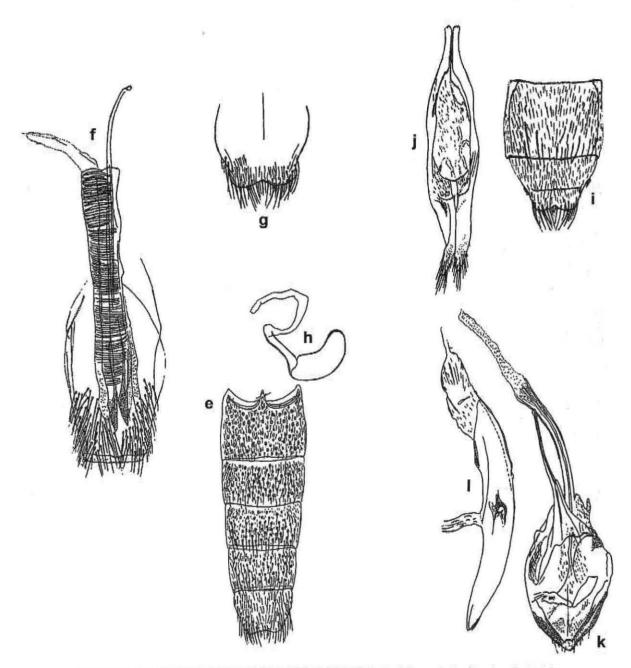


Figure 33 e-I. Female abdominal venter of *Obeneopsis* sp. (e), genitalia (f), sternite V (g), spermatheca (h) and male sternites IV-V (i), part of male genitalia (j), aedeagus (k) and tegmen (l)

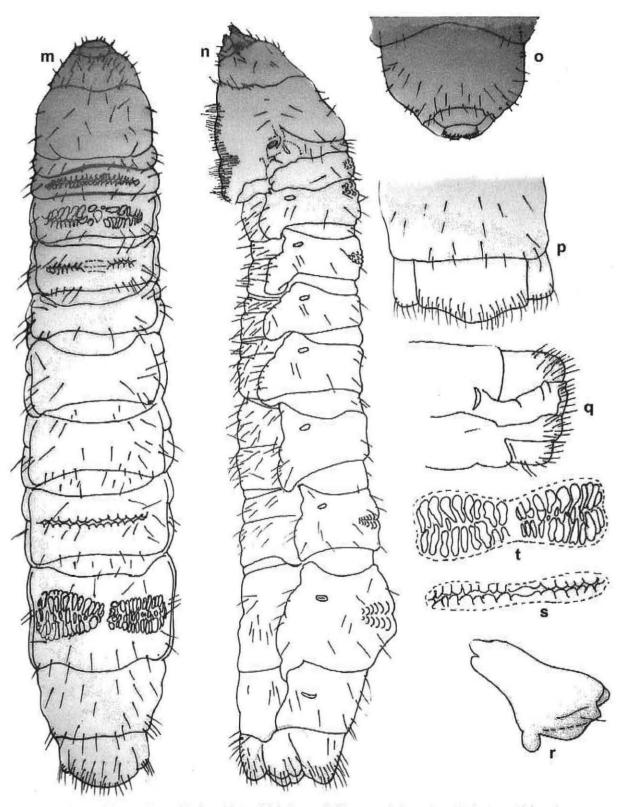


Figure 33 m-t. Dorsal (m) and lateral (n) views of *Obeneopsis* larva, head (o), dorsal (p) and lateral (q) views of segment IX, mandible (r) and ampulla on segment VII (s) and segment VI (t)

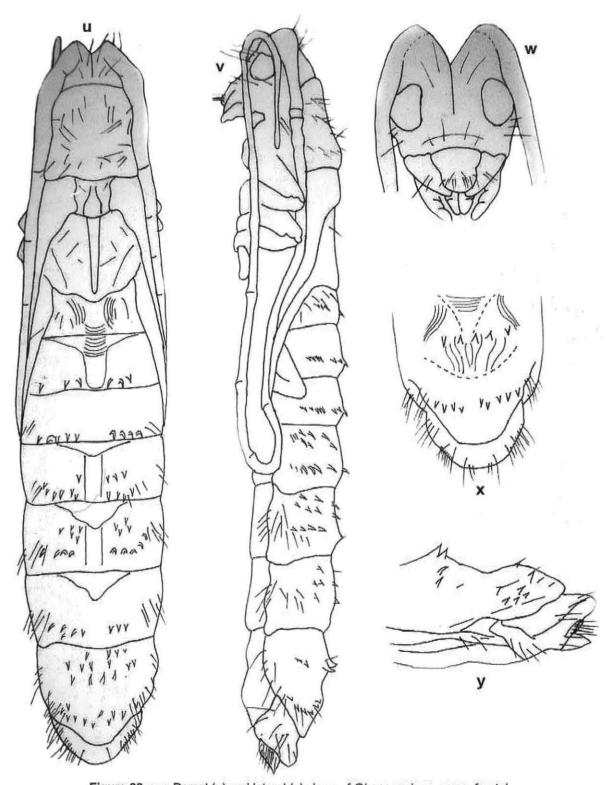


Figure 33 u-y. Dorsal (u) and lateral (v) views of Obeneopsis sp. pupa, frontal view of head (w), spines on the dorsal (x) and lateral (y) portions of abdominal tip

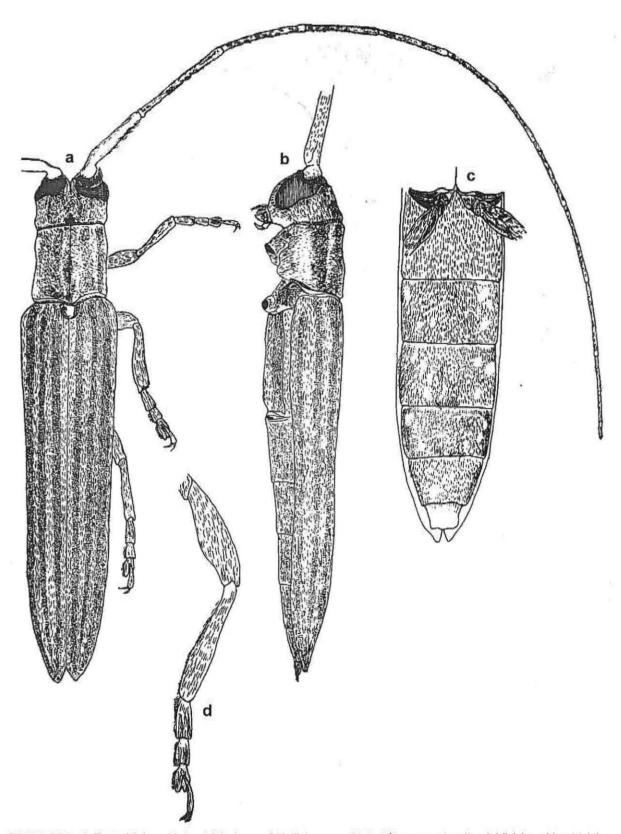


Figure 34 a-d. Dorsal (a) and lateral (b) views of Hyllisia near vittata Fåhraeus, sternites I-VI (c) and leg III (d)

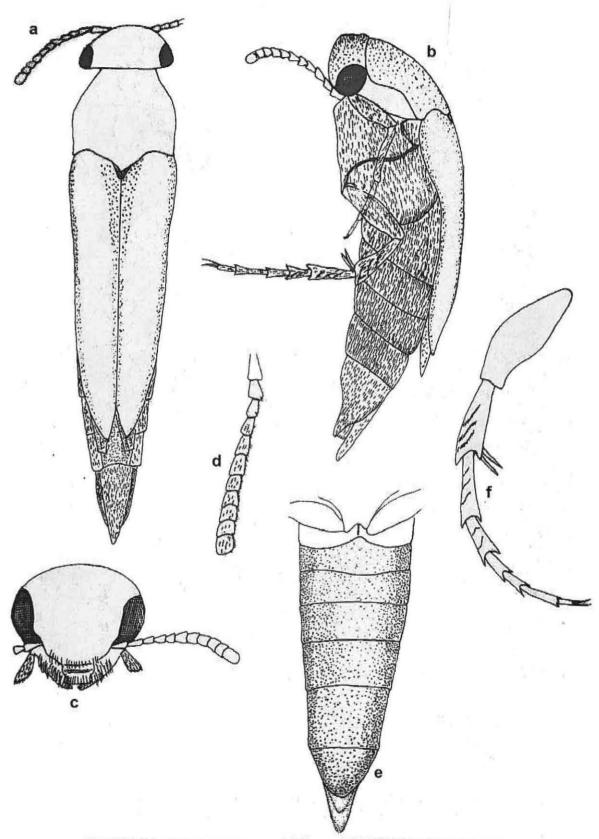


Figure 35 a–f. Dorsal (a) and lateral (b) views of Stenalia near occidentalis Pic, frontal view of head (c), antenna (d), abdominal venter (e) and leg III (f)

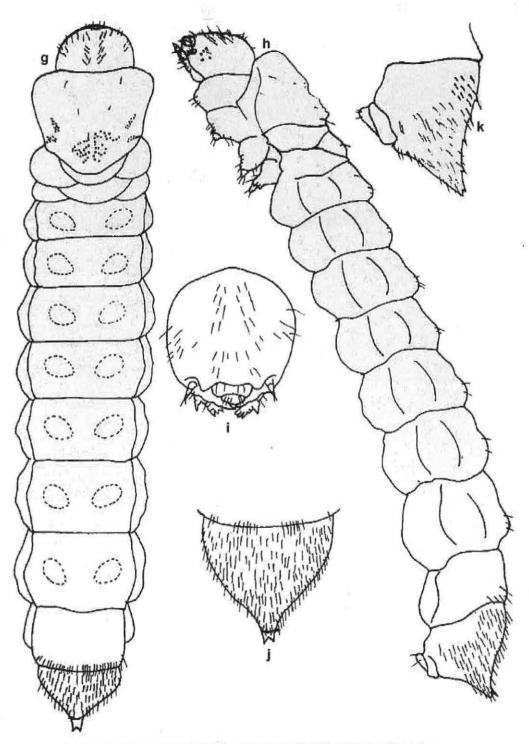


Figure 35 g-k. Dorsal (g) and lateral (h) views of S. near occidentalis larva, frontal view of head (i), dorsal (j) and lateral (k) views of urogomphi

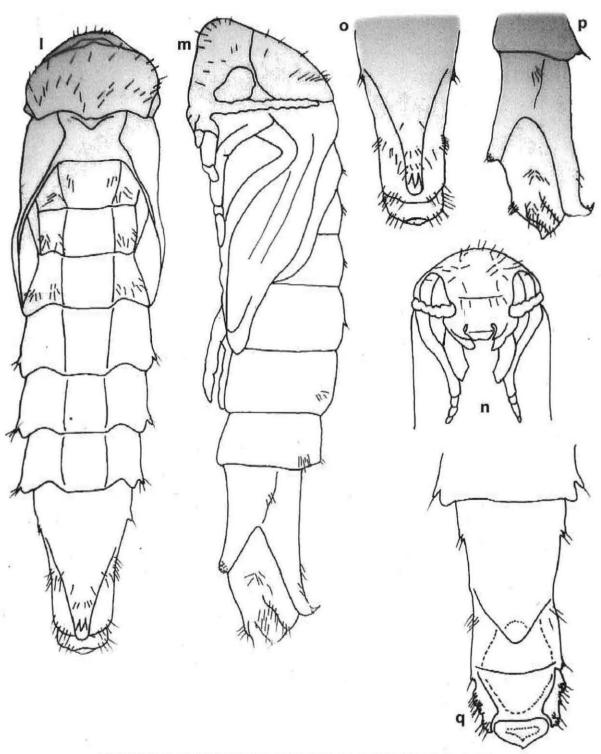


Figure 35 I–q. Dorsal (I) and lateral (m) views of S. near occidentalis pupa, frontal view of head (n), dorsal (o), lateral (p) and ventral (q) views of urogomphi

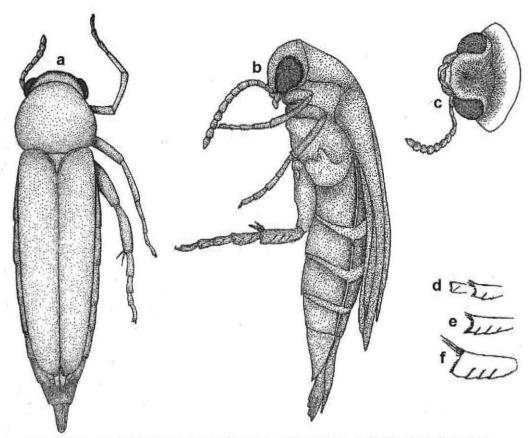


Figure 36 a-f. Dorsal (a) and lateral (b) views of Stenalia sp. B, frontal view of head (c), oblique serrated spines on leg III in tarsal segment II and III (d), tarsus I (e) and tibia III (f)

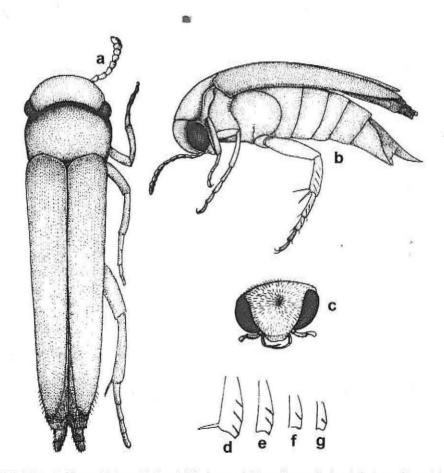


Figure 37 a-g. Dorsal (a) and lateral (b) views of Stenalia sp. C, frontal view of head (c), oblique serrated spines in tibia III (d), basitarsus (e), tarsal segment II (f), and III (g) of leg III

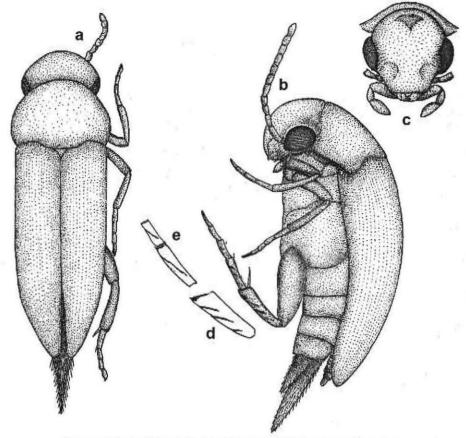


Figure 38 a-e. Dorsal (a) and lateral (b) views of Stenalia sp. D, frontal view of head (c), tibia III (d), and basal two segments of tarsus (e)

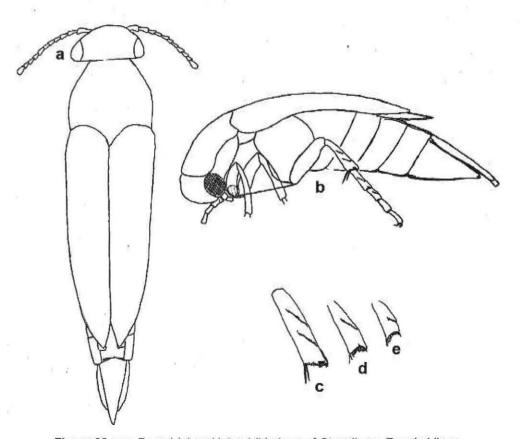


Figure 39 a-e. Dorsal (a) and lateral (b) views of Stenalia sp. E and oblique spines of tibia III (c), basitarsus (d), and second tarsal segment (e) of leg III

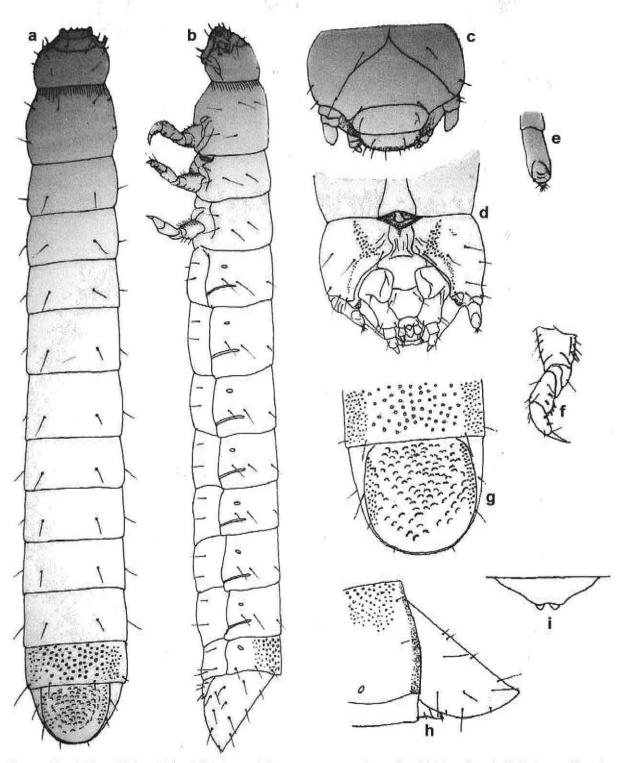


Figure 40 a–i. Dorsal (a) and lateral (b) views of *Paramarygmus* sp. larva, frontal (c) and ventral (d) views of head, antenna (e), proleg I (f), dorsal (g) and lateral (h) views of segments VIII–IX and ventral view of sternite X (i)

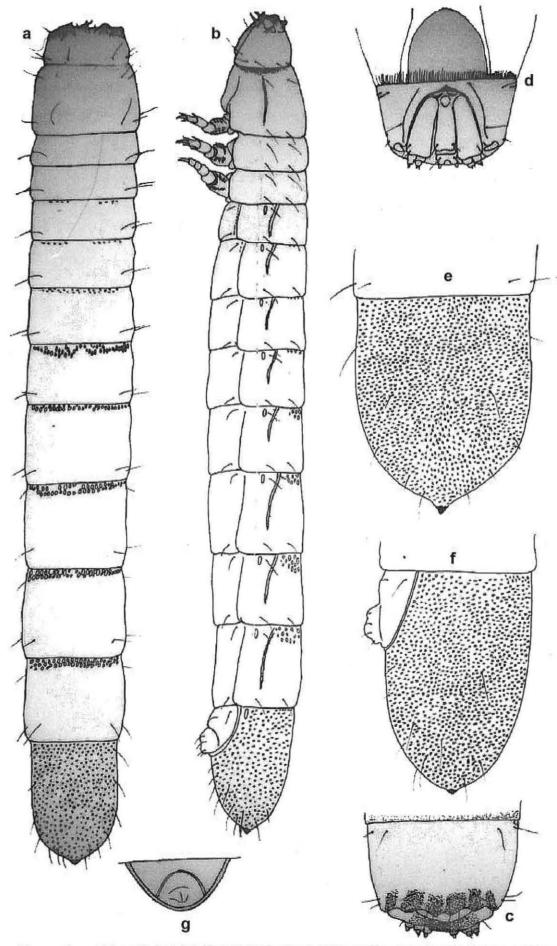


Figure 41 a-g. Dorsal (a) and lateral (b) views of Zophodes sp. larva, frontal (c) and ventral (d) views of head, dorsum of abdominal segment IX (e), lateral view of segment IX-X (f) and ventral view of segment X (g)

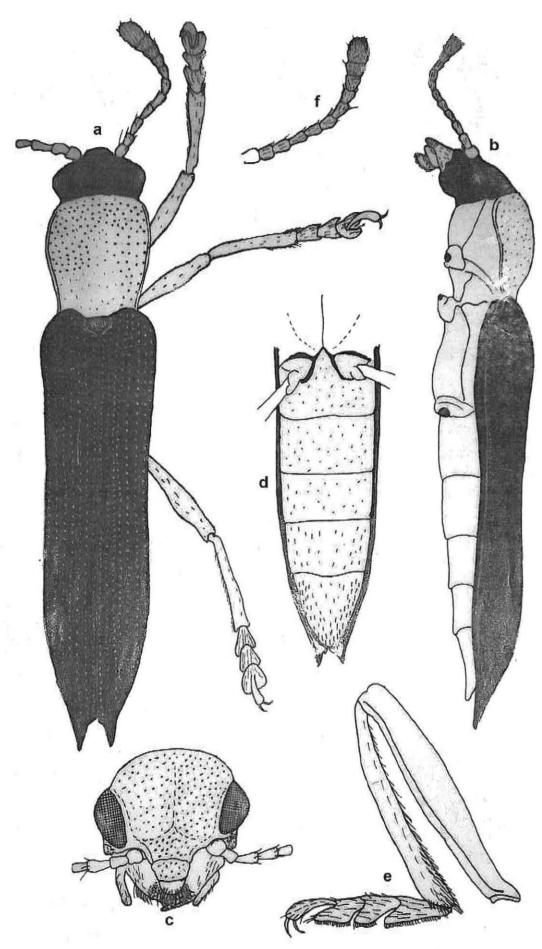


Figure 42 a-f. Dorsal (a) and lateral (b) views of Stenolanguria caudata, frontal view of head (c), abdominal venter (d), leg III (e) and antenna (f)

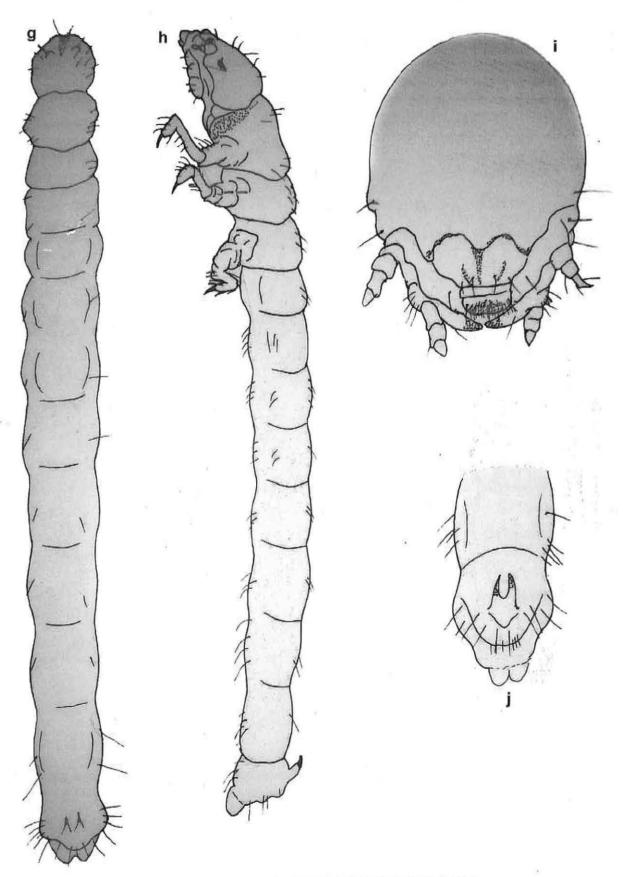


Figure 42 g-j. Dorsal (g) and lateral (h) views of S. caudata larva, frontal view of head (i), and dorsal view of urogomphi (j)

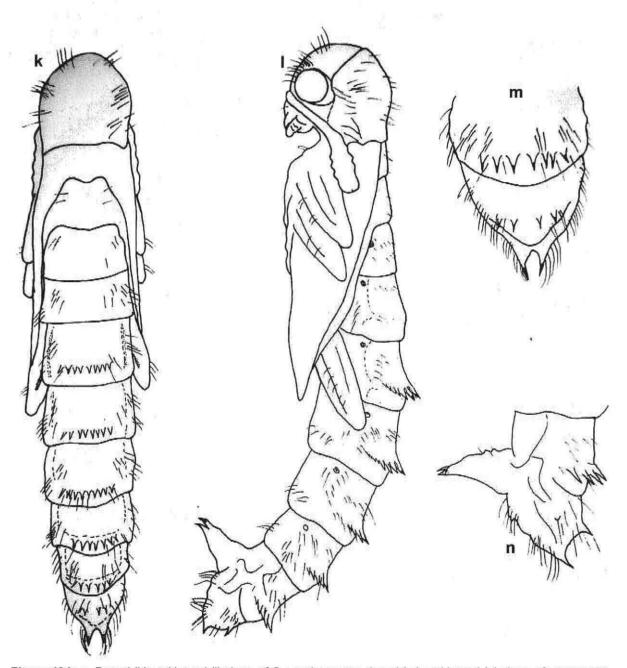


Figure 42 k-n. Dorsal (k) and lateral (l) views of S. caudata pupa, dorsal (m) and lateral (n) views of urogomphi

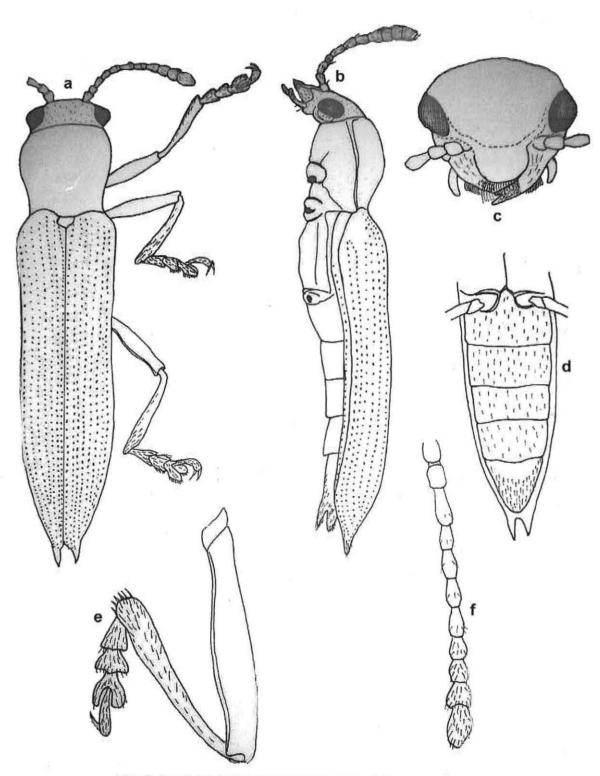


Figure 43 a-f. Dorsal (a) and lateral (b) views of Stenolanguria sp., frontal view of head (c), abdominal venter (d), leg III (e) and antenna (f)

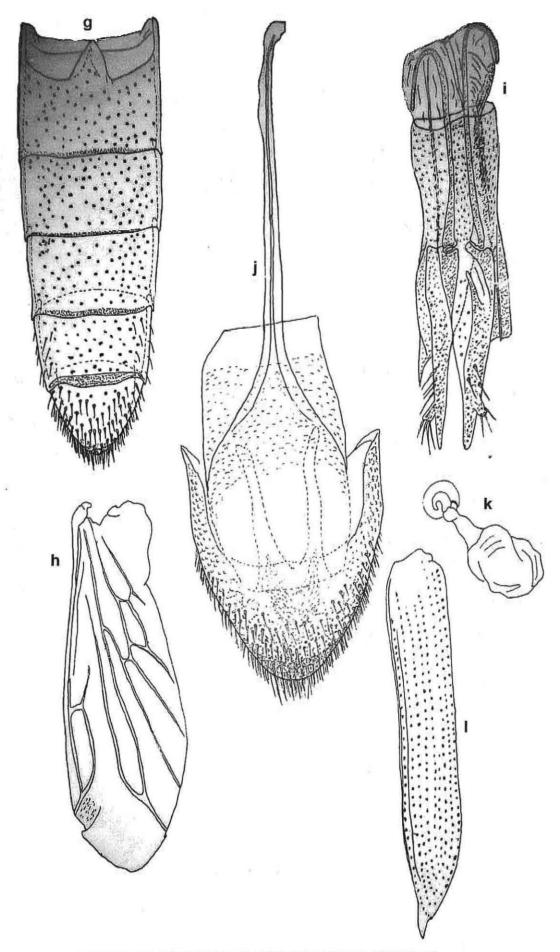


Figure 43 g-I. Stenolanguria sp. abdominal venter (g), hindwing (h), female genitalia (i, j), spermatheca (k) and right elytron (l)

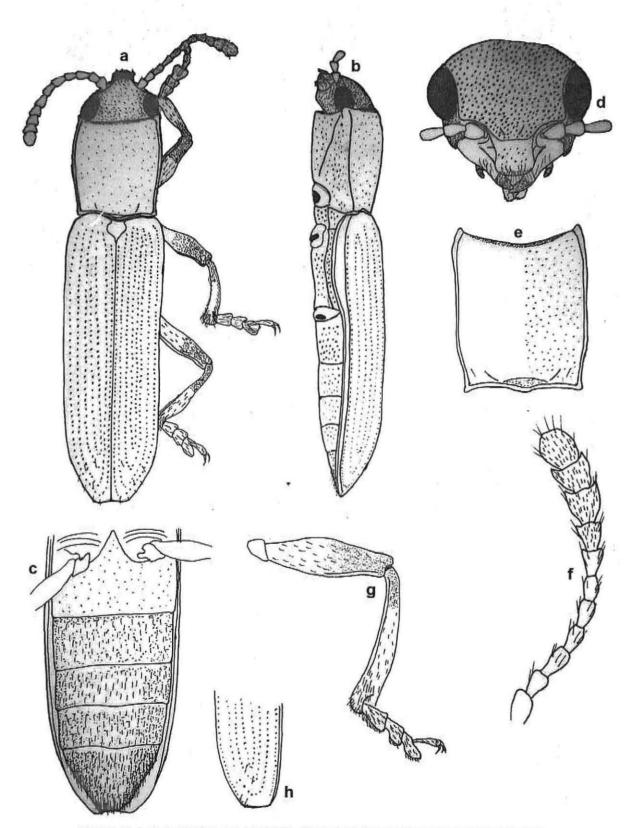


Figure 44 a-h. Dorsal (a) and lateral (b) views of Barbaropus near olseni Pic, venter of abdomen (c), frontal view of head (d), pronotum (e), antenna (f), leg I (g) and tip of elytron (h)

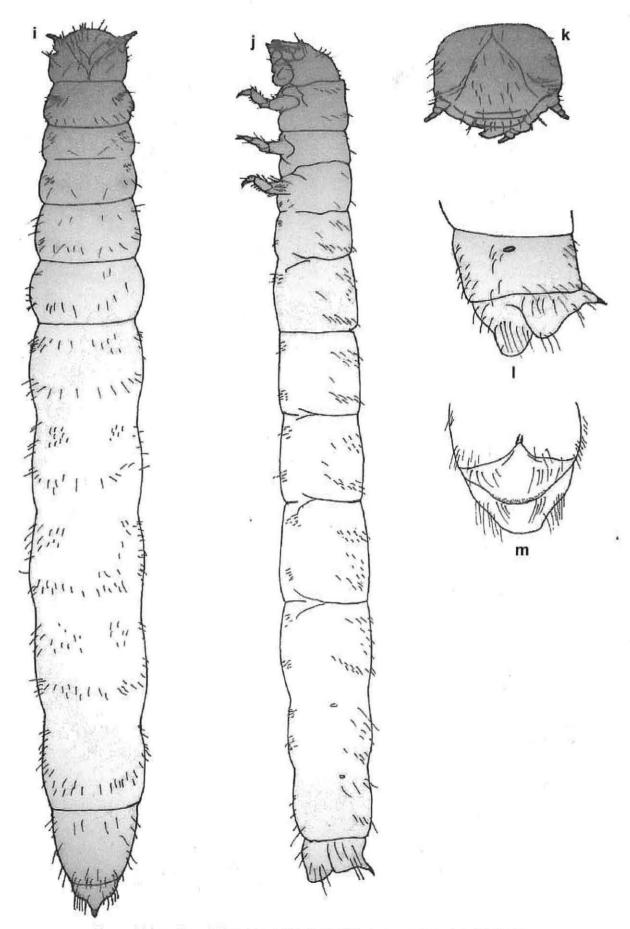


Figure 44 i-m. Dorsal (i) and lateral (j) views of Barbaropus near olseni Pic larva, frontal view of head (k), lateral (l) and dorsal (m) views of urogomphi

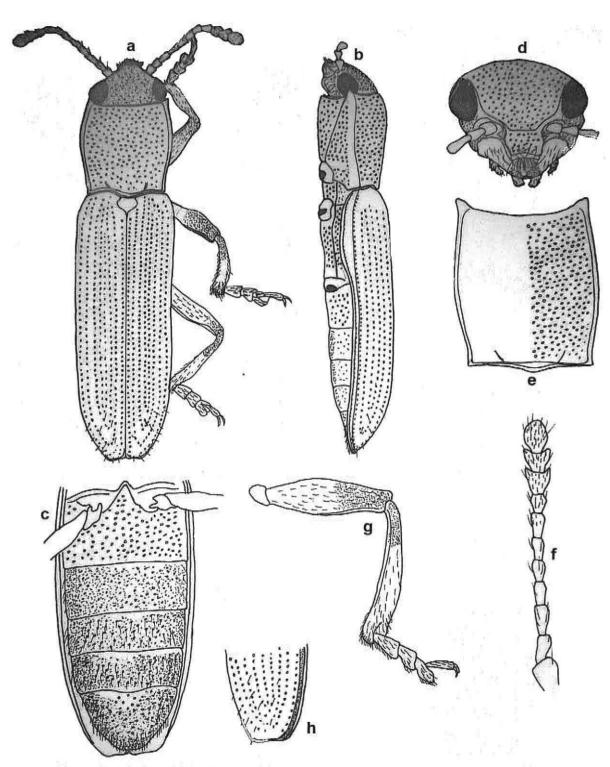


Figure 45 a-h. Dorsal (a) and lateral (b) views of *Barbaropus* sp. C, venter of abdomen (c), frontal view of head (d), pronotum (e), antenna (f), leg I (g) and tip of elytron (h)

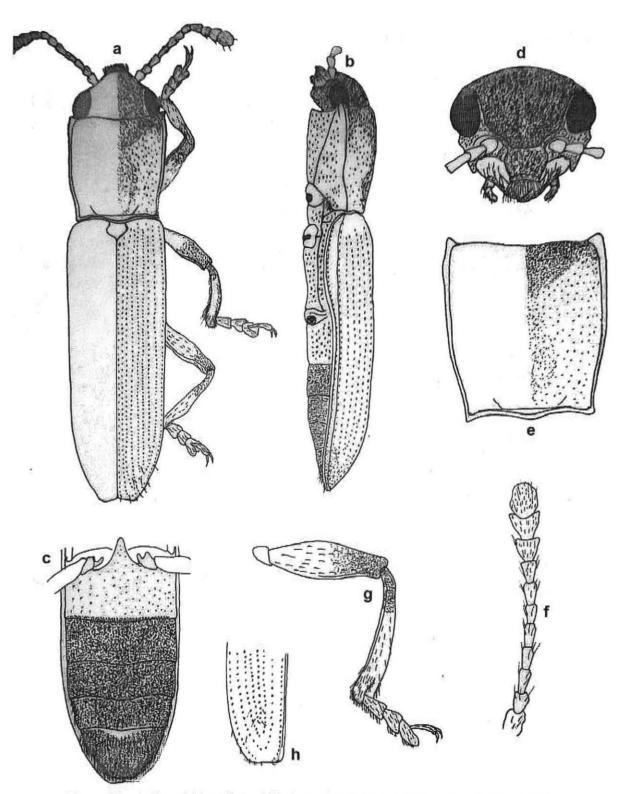


Figure 46 a-h. Dorsal (a) and lateral (b) views of *Barbaropus* sp. B, venter of abdomen (c), frontal view of head (d), pronotum (e), antenna (f), leg I (g) and tip of elytra (h)

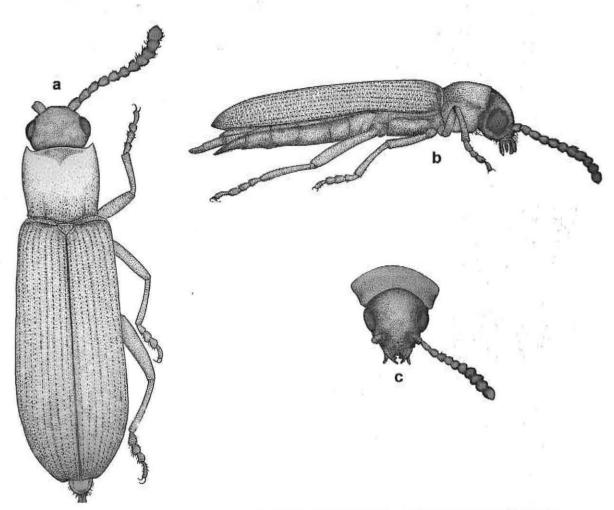


Figure 47 a-c. Dorsal (a) and lateral (b) views of Barbaropus sp. D and frontal view of head (c)

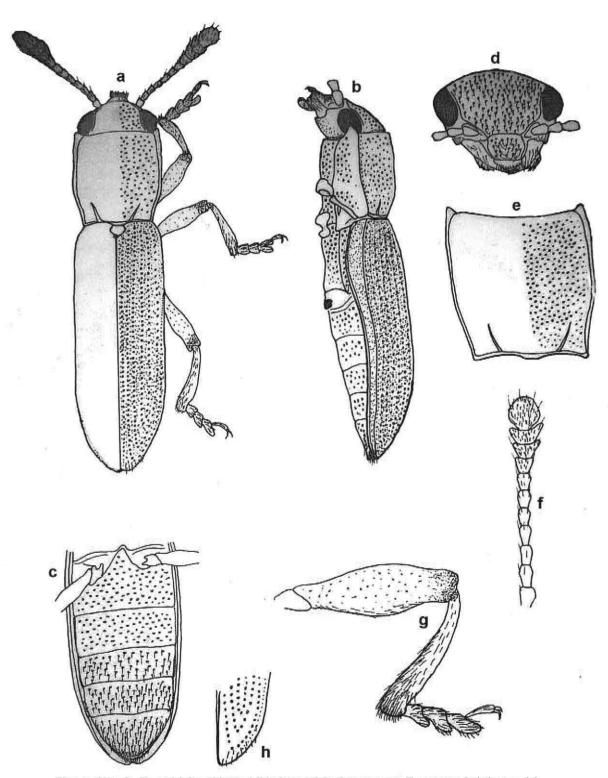


Figure 48 a-h. Dorsal (a) and lateral (b) views of *Barbaropus* sp. E, venter of abdomen (c), frontal view of head (d), pronotum (e), antenna (f), leg I (g) and tip of elytron (h)

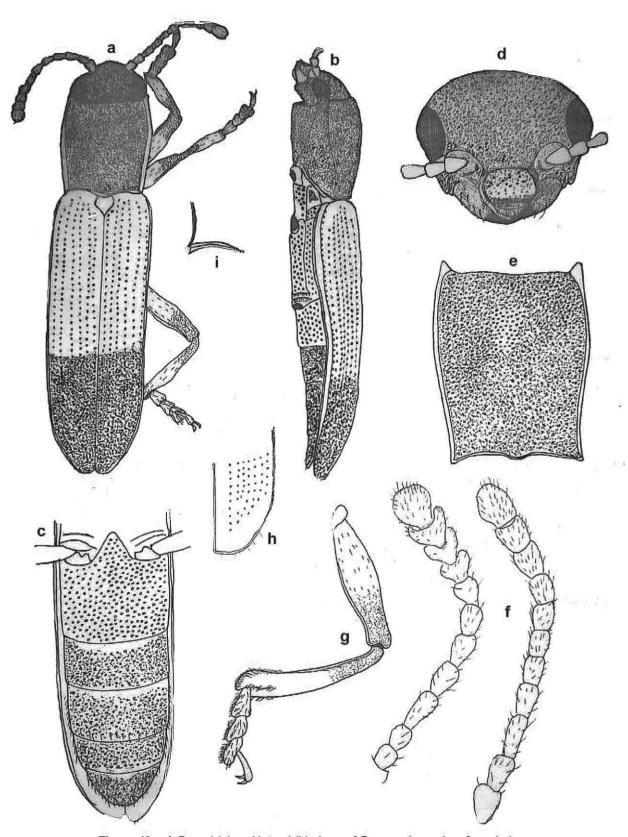


Figure 49 a-i. Dorsal (a) and lateral (b) views of *Promecolanguria rufocephala*, venter of abdomen (c), frontal view of head (d), pronotum (e), two forms of antenna (f), leg I (g), tip of elytron (h) and posterior lateral angle of pronotum (i)

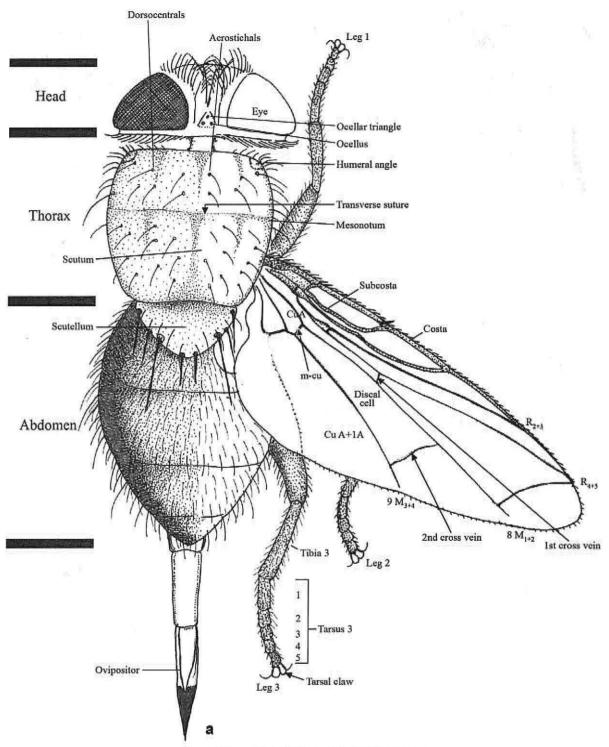


Figure 50 a. Dorsal view of a hypothetical fly

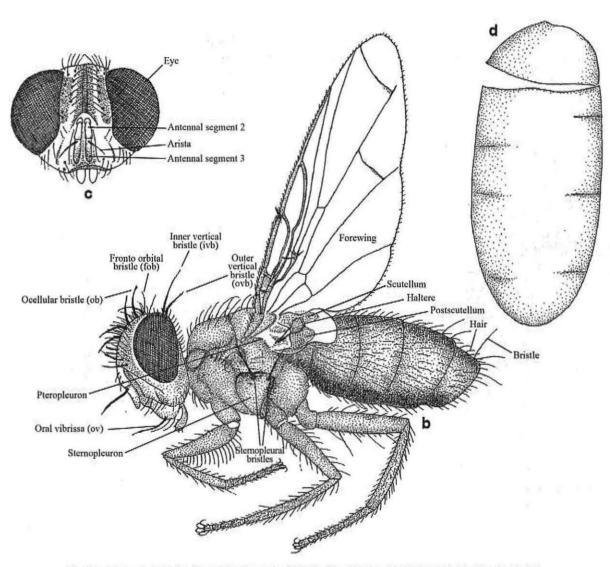


Figure 50 b-d. Lateral view of a hypothetical fly (b), frontal view of head (c) and pupa (d)

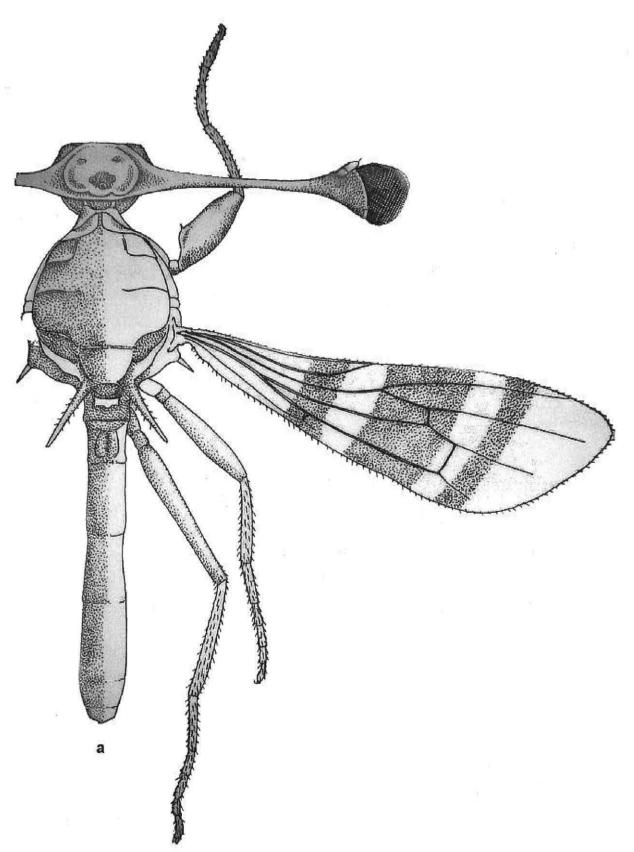


Figure 51 a. Dorsal view of Diopsina sp.

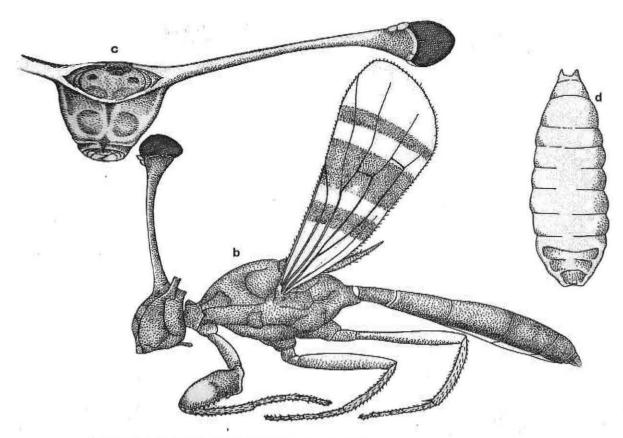


Figure 51 b-d. Lateral view of Diopsina sp. (b), frontal view of head (c) and pupa (d)

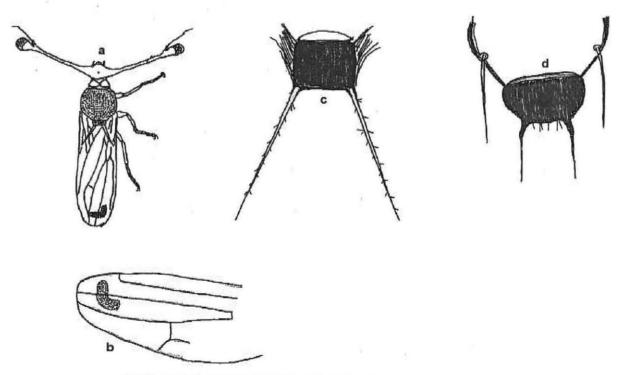


Figure 52 a-d. Dorsal view of *Diopsis* near *lindneri* Feijen (a), spot on forewing (b), scutellum (c) and scutellum of *Diopsis* sp. B (d)

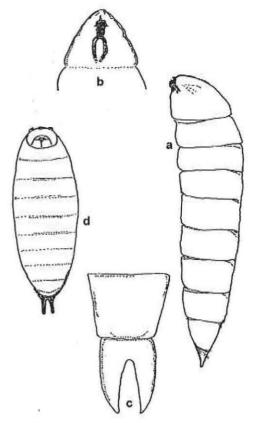


Figure 53 a-d. Lateral view of Mepachymerus sp. larva (a), dorsal view of head (b), posterior end (c) and dorsal view of pupa (d)

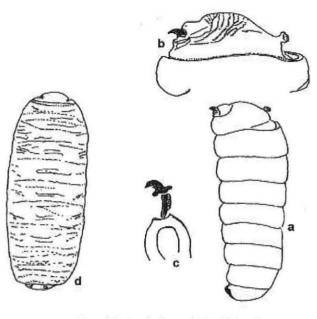


Figure 54 a-d. Lateral view of Elachiptereicus abessynicus Becker larva (a), head (b), cephalopharyngeal skeleton (c) and pupa (d)

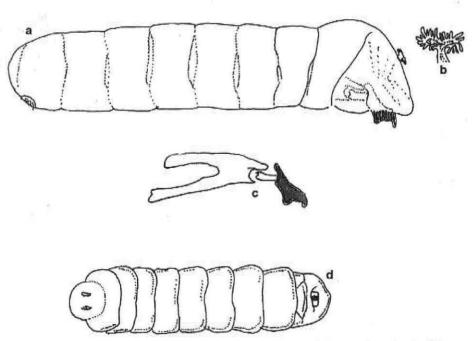


Figure 55 a–d. Lateral view of *Pachylophus* sp. larva (a), anterior spiracle (b), cephalopharyngeal skeleton (c) and pupa (d)

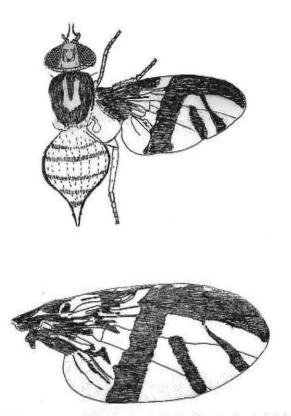


Figure 56 a-b. Dorsal view of Bistrispinaria fortis (Speiser) adult (a) and wing (b)

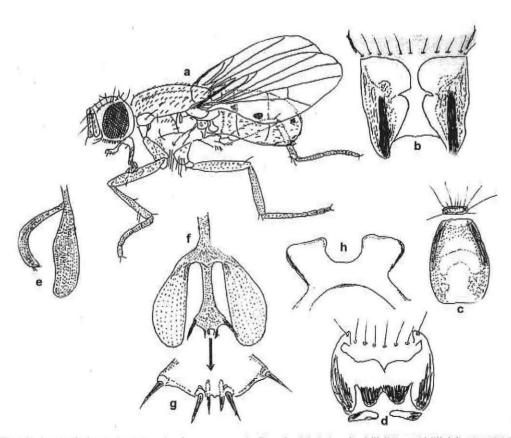


Figure 57 a-h. Lateral view of adult Atherigona soccata Rondani (a), tergite VII (b) and VIII (c), sternite VII (d), lateral (e) and dorsal (f) views of trifoliate process, tip of median process (g) and hypopygial prominence (h)

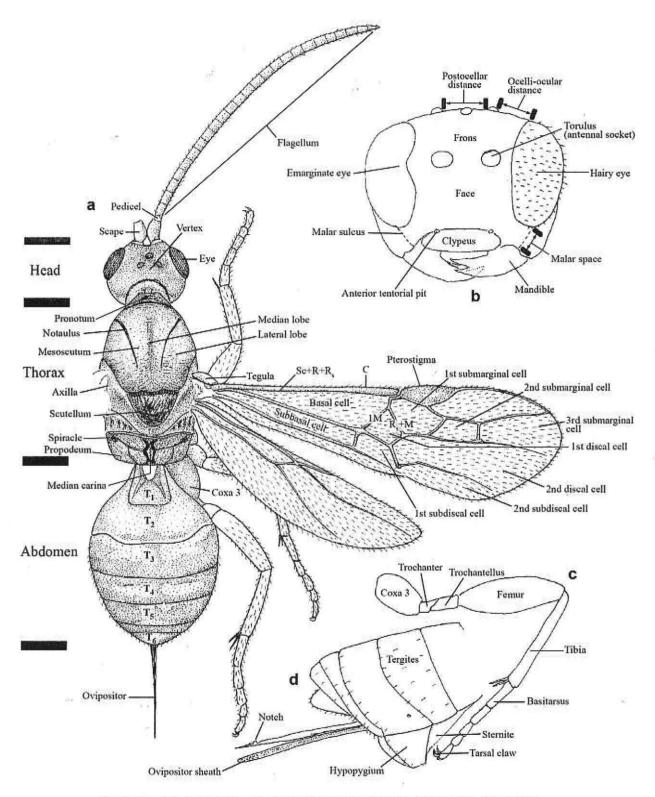


Figure 58 a–d. Dorsal view of a parasitic hymenoptera (a), frontal view of head (b), leg III (c) and posterior abdomen showing ovipositor (d)

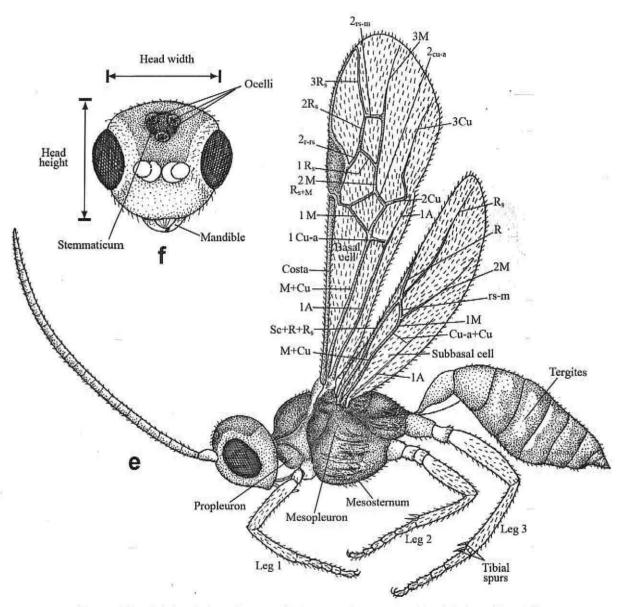


Figure 58 e-f. Lateral view of a parasitic hymenopteran (e) and frontal view of head (f)

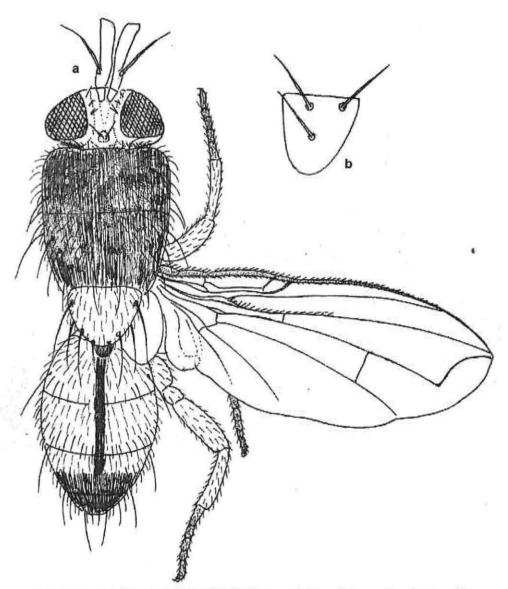


Figure 59 a-b. Dorsal view of adult Siphona sp. A (a) and sternopleural setae (b)

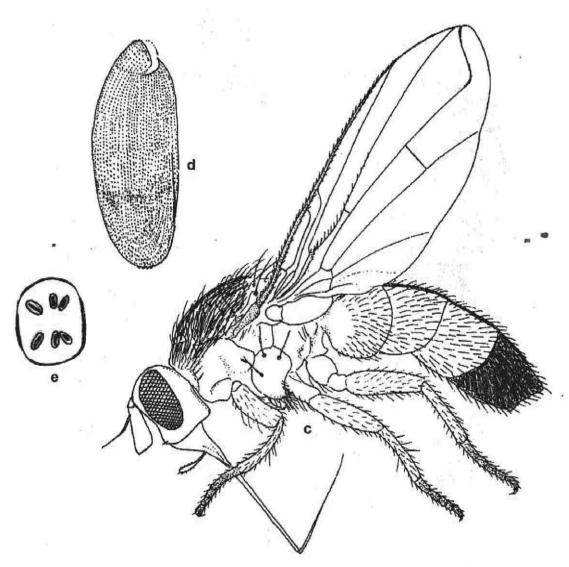


Figure 59 c-e. Lateral view of Siphona sp. A (c), pupa (d) and spiracle (e)

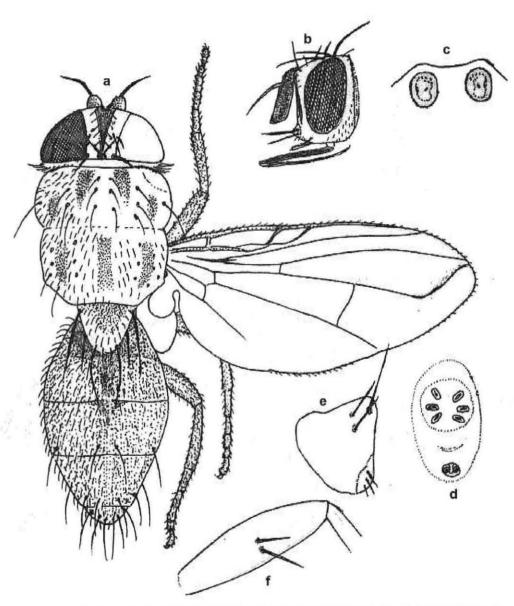


Figure 60 a–f. Dorsal view of Siphona sp. B (a), lateral view of head (b), anterior spiracle (c) and enlarged view (d), sternopleural setae (e) and spines on femur III (f)

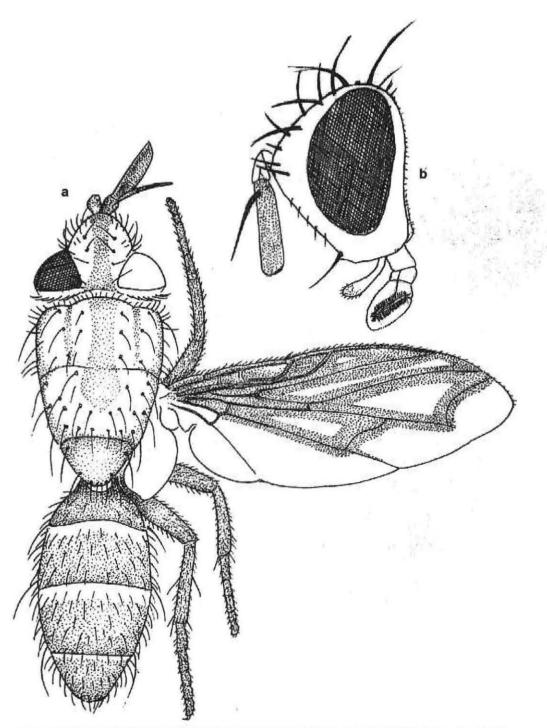


Figure 61 a-b. Dorsal view of Lydella near sesamiae Mesnil (a) and lateral view of head (b)

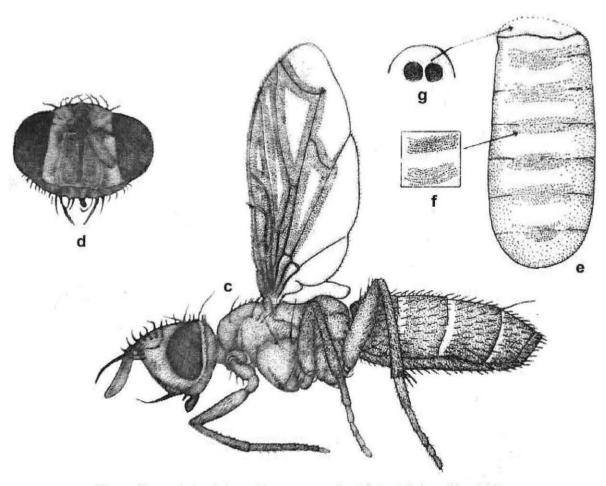


Figure 61 c-g. Lateral view of L. near sesamiae (c), frontal view of head (d), pupa (e), sculpture on pupal surface (f) and spiracle (g)

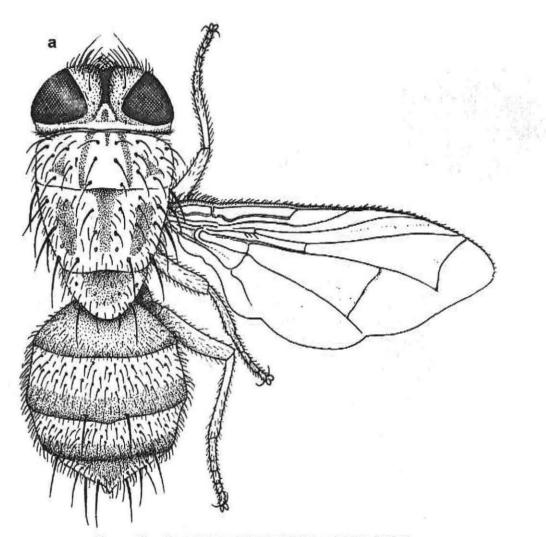


Figure 62 a. Dorsal view of Descampsina sesamiae Mesnil

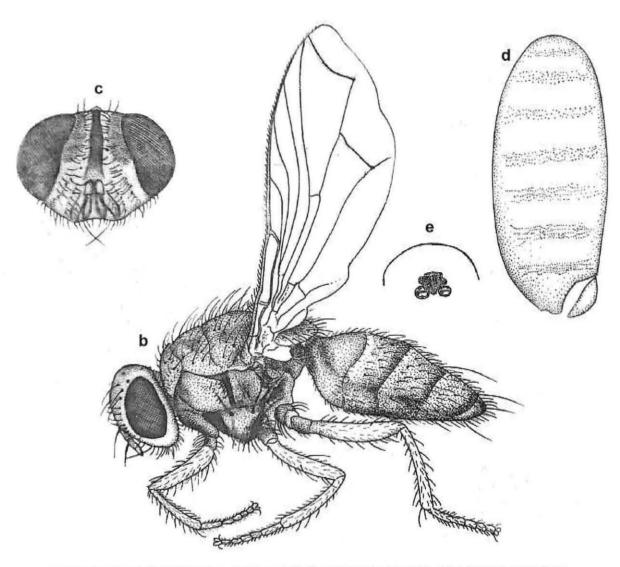


Figure 62 b-e. Lateral view of D. sesamiae (b), frontal view of head (c), pupa (d) and spiracle (e)

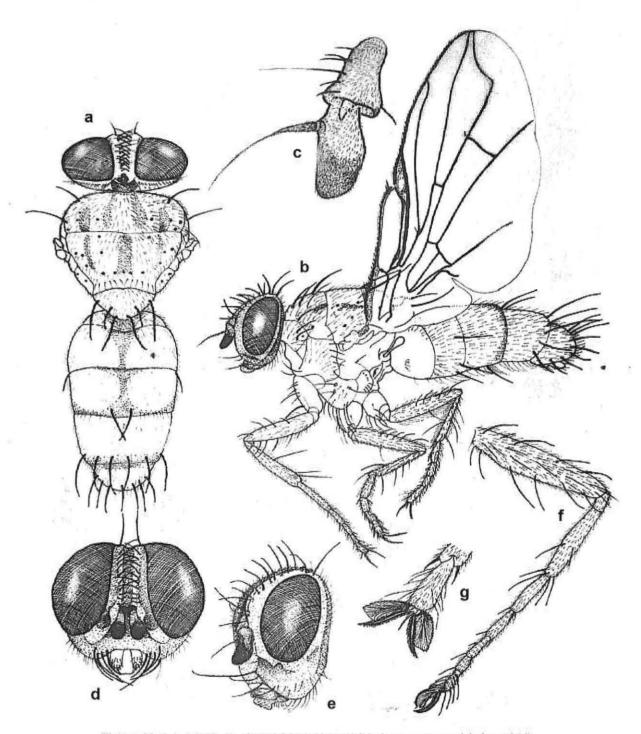


Figure 63 a-g. Leskia sp. dorsal (a) and lateral (b) views, antenna (c), frontal (d) and lateral (e) views of head, leg III (f) and tarsal claw (g)

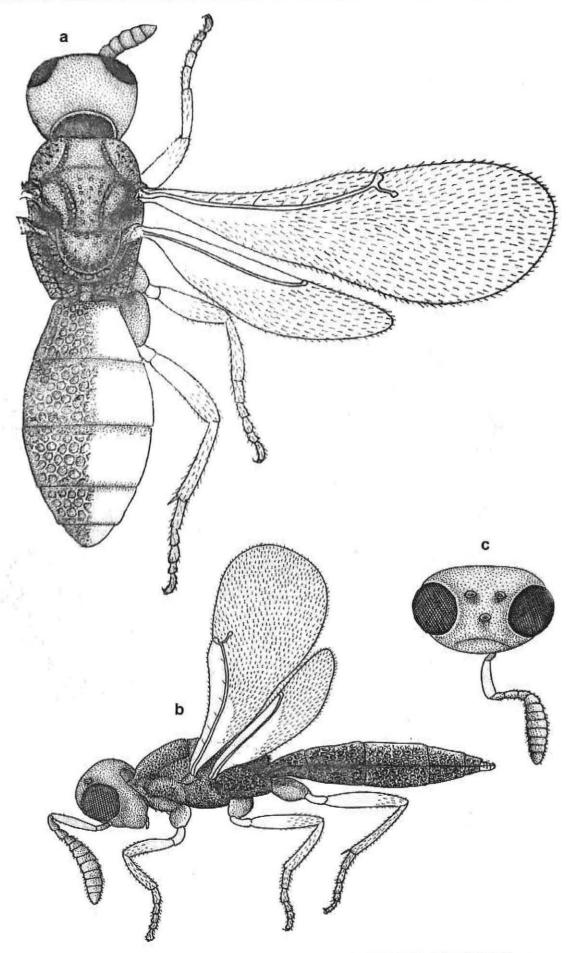


Figure 64 a-c. Dorsal (a) and lateral (b) views of Scelio sp. and frontal view of head (c)

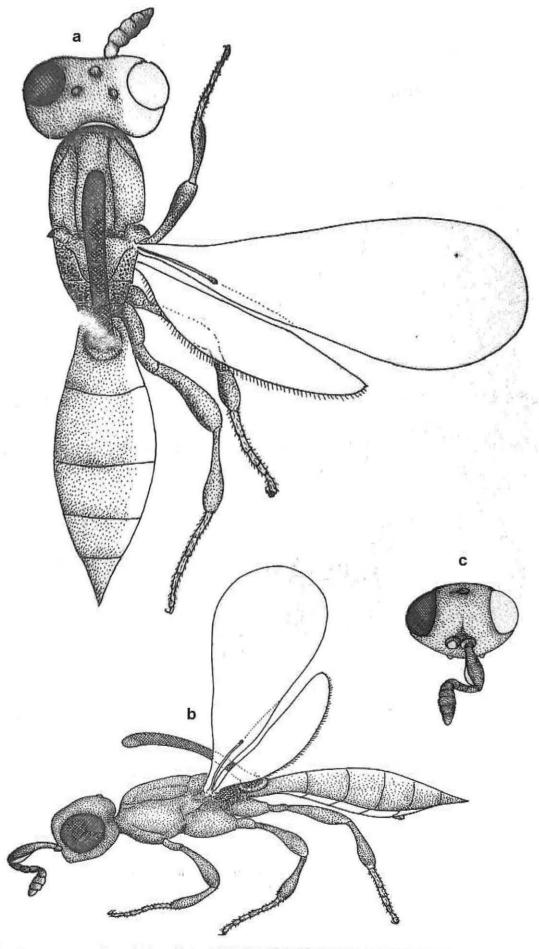


Figure 65 a-c. Dorsal (a) and lateral (b) views of Inostemma sp. and frontal view of head (c)

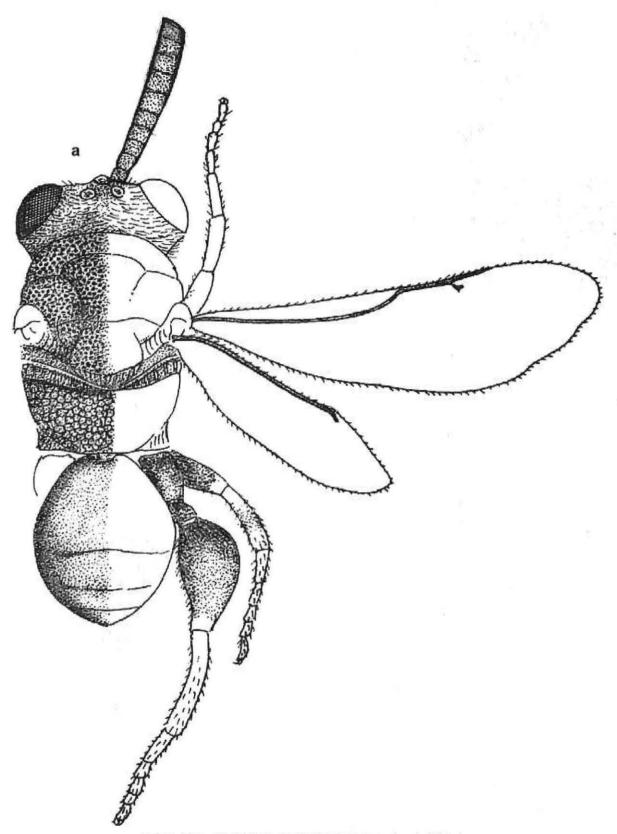


Figure 66 a. Dorsal view of Brachymeria kassalensis (Kirby)

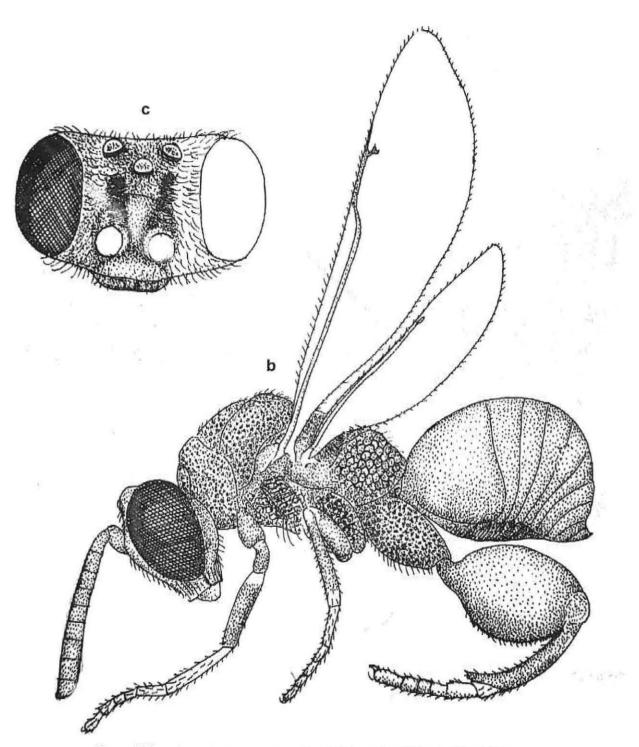


Figure 66 b-c. Lateral view of B. kassalensis (b) and frontal view of head (c)

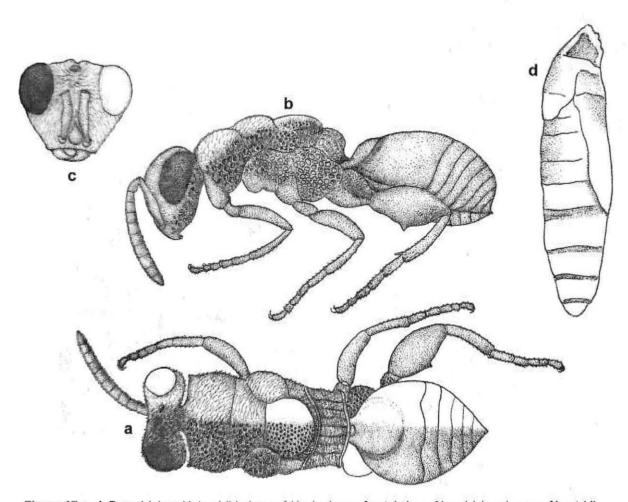


Figure 67 a-d. Dorsal (a) and lateral (b) views of Hockeria sp., frontal view of head (c) and pupa of host (d)

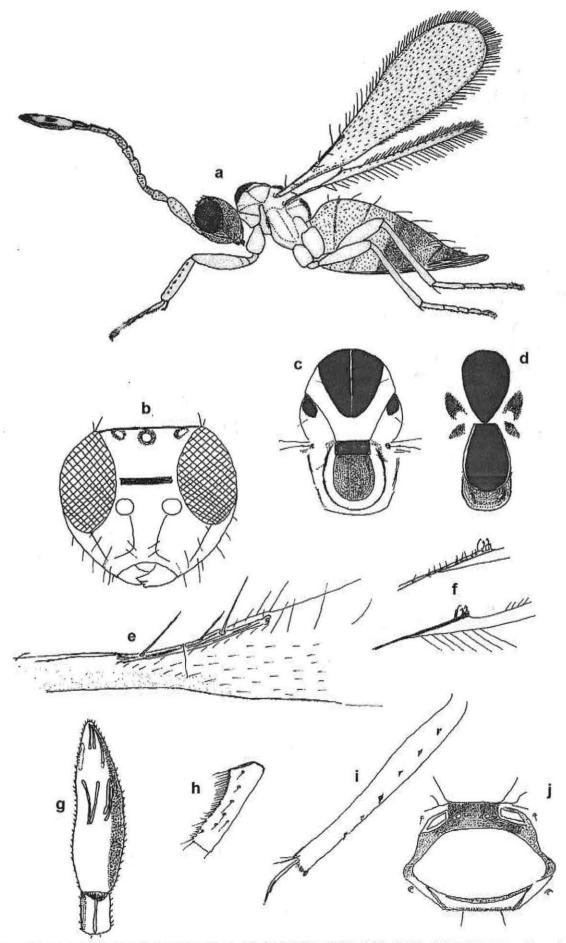


Figure 68 a-j. Lateral view of Gonatocerus sp. (a), frontal view of head (b), meso- and metanotum of female (c) and male (d), wing venation (e), hindwing (f), antennal club (g), tarsal bristles (h), tibia I (i) and propodeum (j)

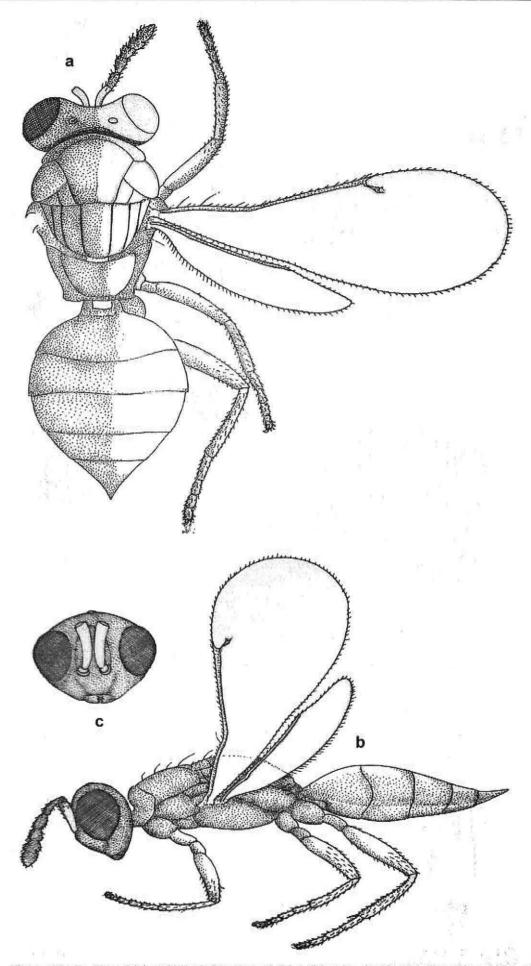


Figure 69 a-c. Dorsal (a) and lateral (b) views of Tetrastichus sp. and frontal view of head (c)

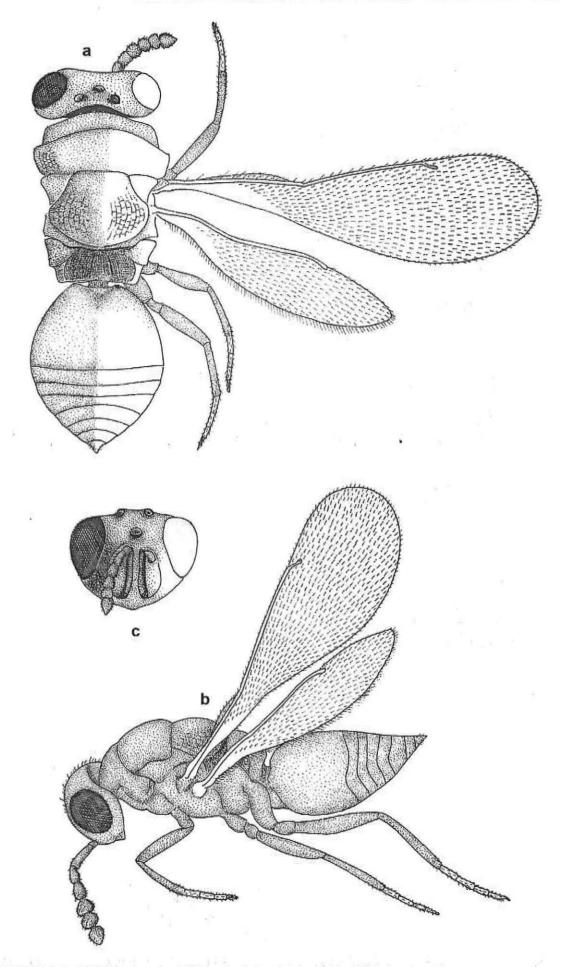


Figure 70 a-c. Pediobius homoeus (Waterston) dorsal (a) and lateral (b) views and frontal view of head (c)

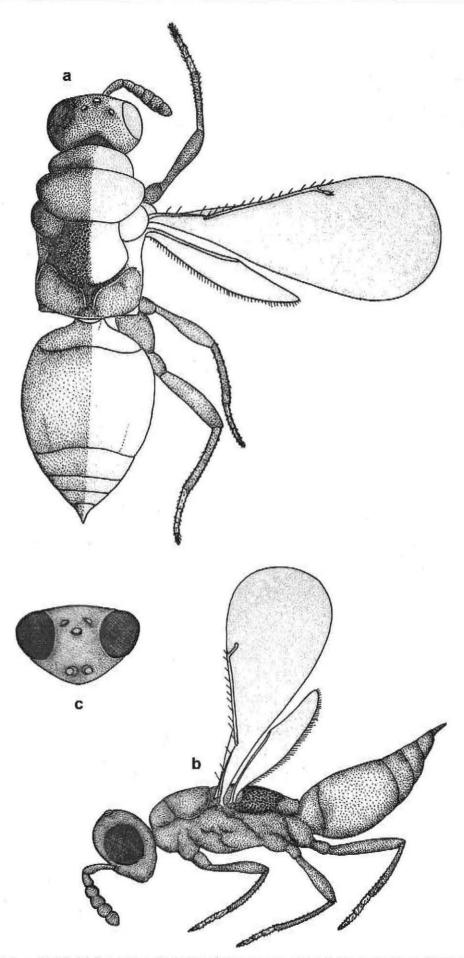


Figure 71 a-c. Dorsal (a) and lateral (b) views and frontal view of head (c) of Pediobius furvus (Gahan)

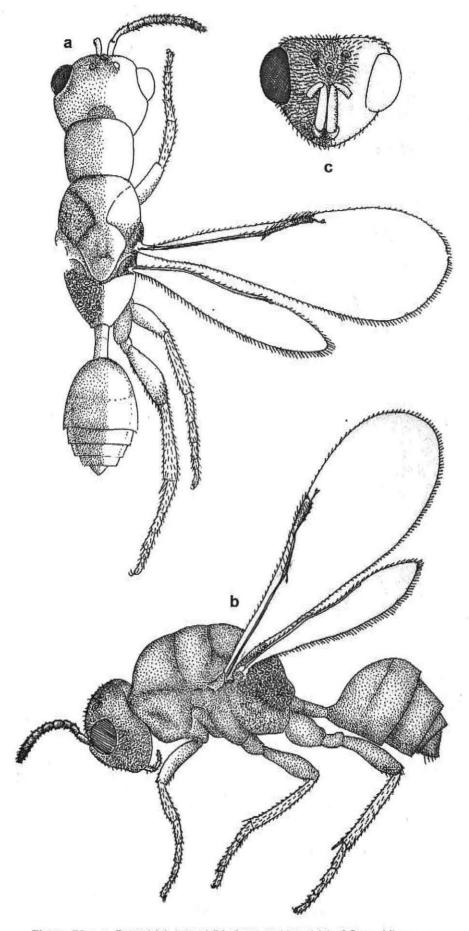


Figure 72 a-c. Dorsal (a), lateral (b) views and head (c) of Sycophila sp.

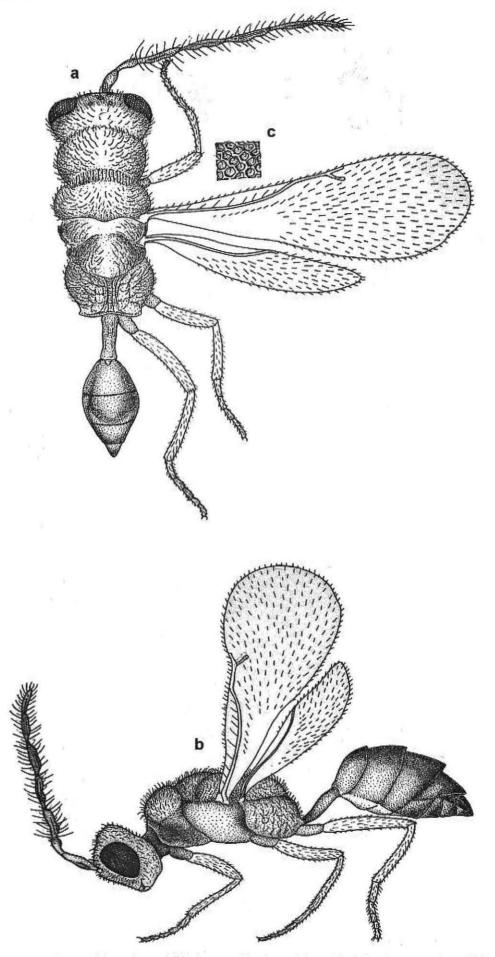


Figure 73 a-c. Dorsal (a) and lateral (b) views and body sculpture (c) of Eurytoma oryzivora Delvare

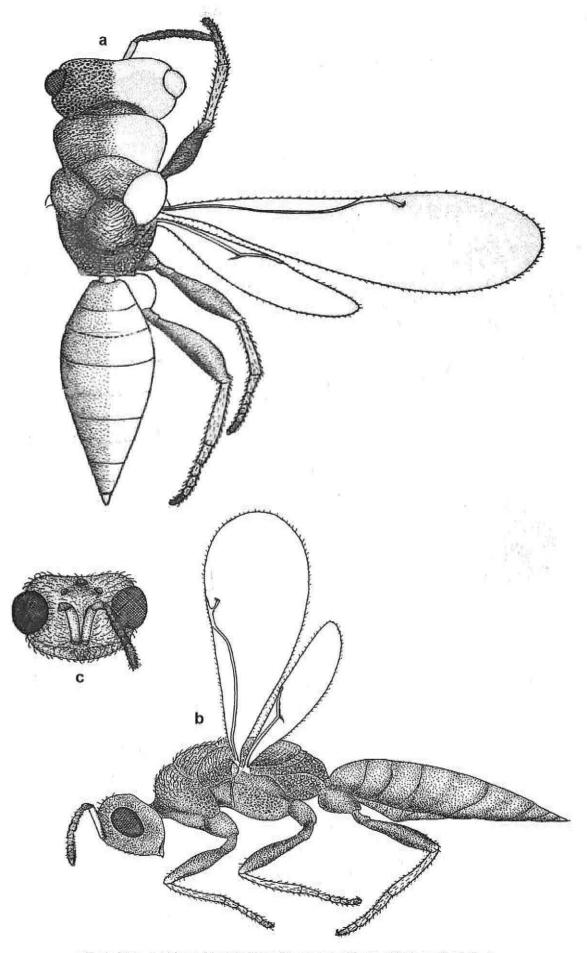


Figure 74 a-c. Dorsal (a) and lateral (b) views and head (c) of Eurytoma sp. A

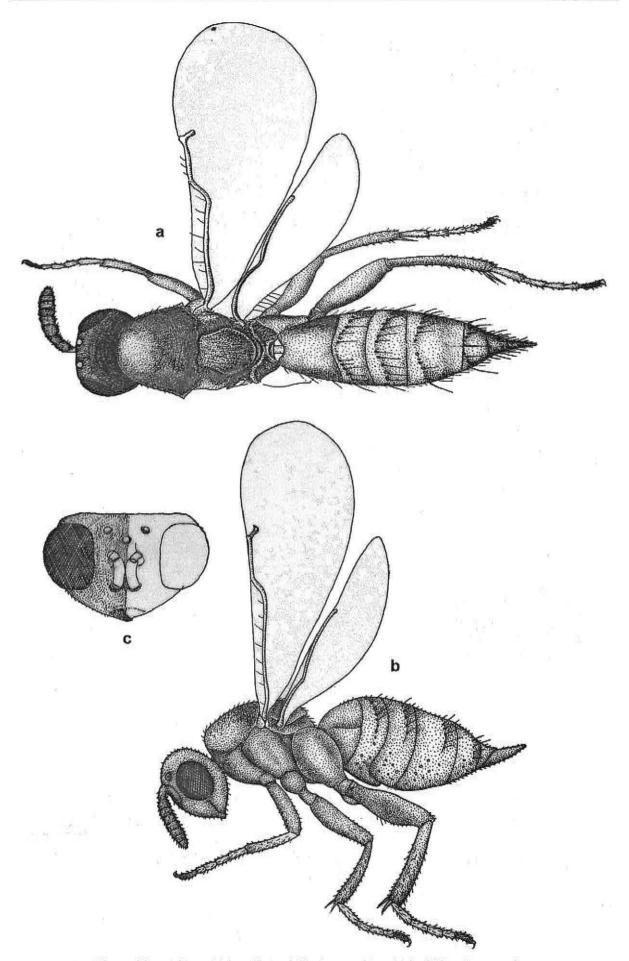


Figure 75 a-c. Dorsal (a) and lateral (b) views and head (c) of ?Eurytoma sp. C

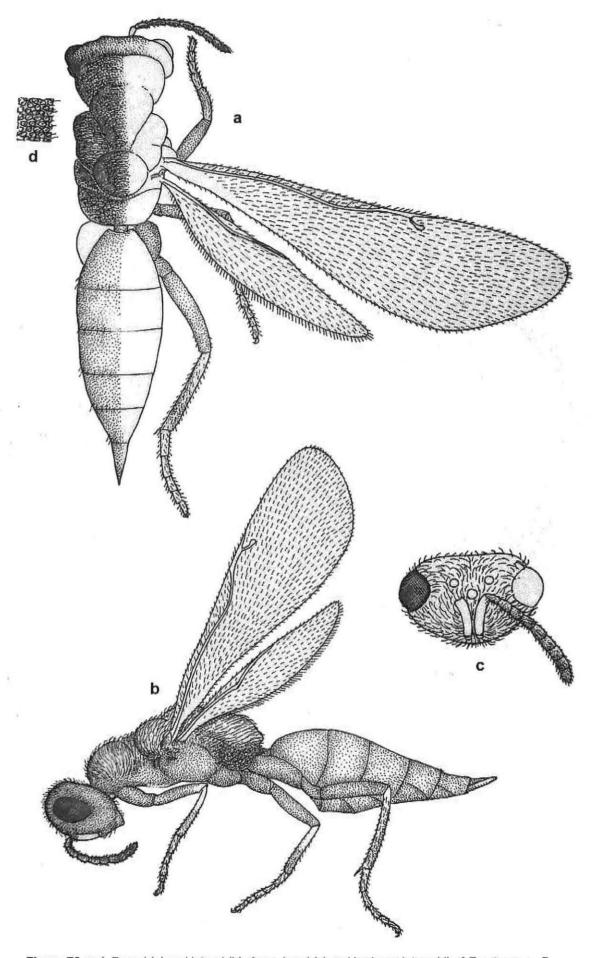


Figure 76 a-d. Dorsal (a) and lateral (b) views, head (c) and body sculpture (d) of Eurytoma sp. D

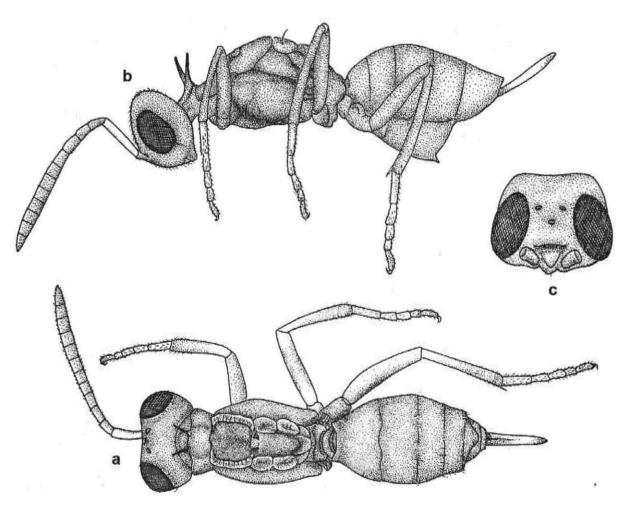


Figure 77 a-c. Dorsal (a) and lateral (b) views and head (c) of Macroneura sp.

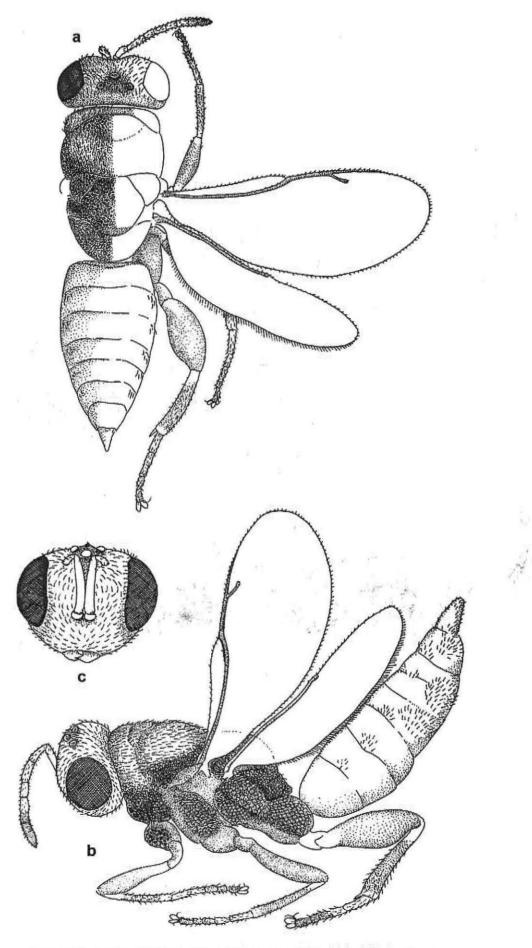


Figure 78 a-c. Dorsal (a) and lateral (b) views and head (c) of Norbanus sp.

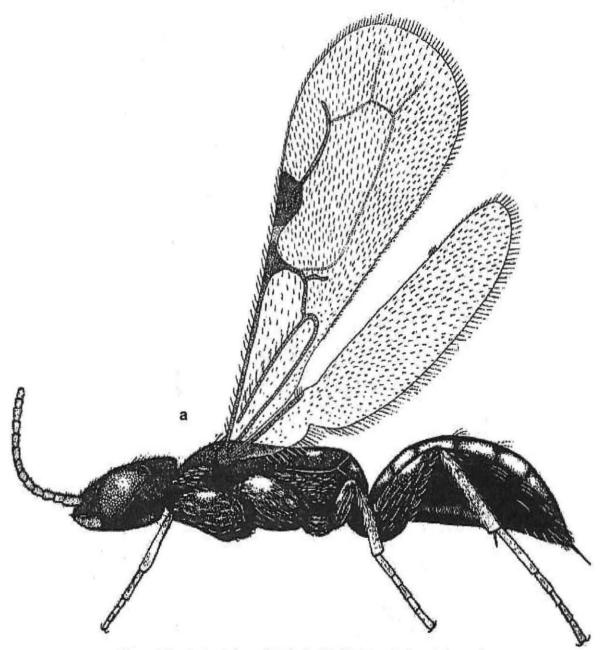


Figure 79 a. Lateral view of the bethylid, Goniozus indicus Ashmead

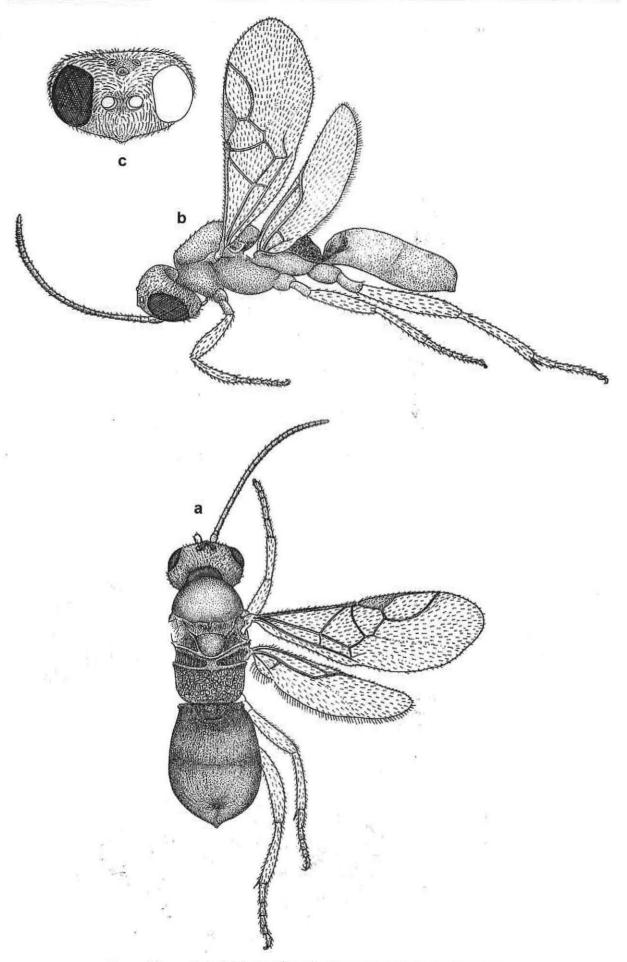


Figure 80 a-c. Dorsal (a) and lateral (b) views and head (c) of Triaspis sp.

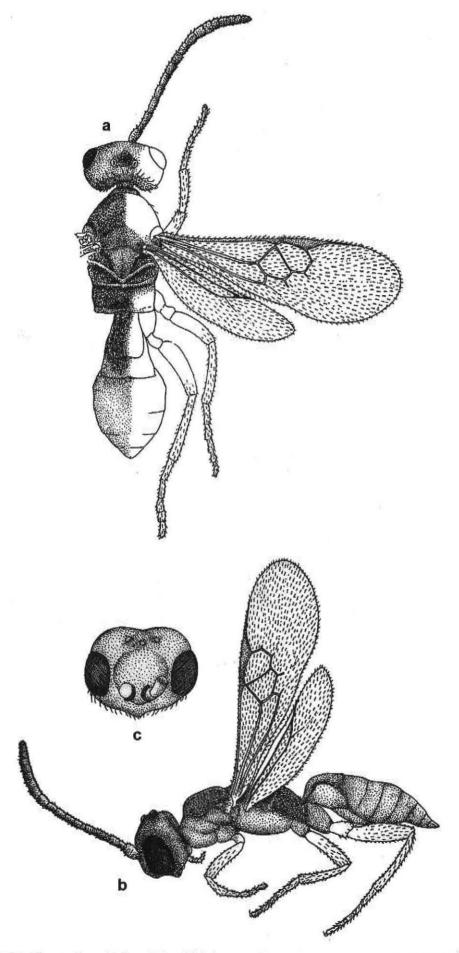


Figure 81 a-c. Dorsal (a) and lateral (b) views and head (c) of Cotesia flavipes Cameron

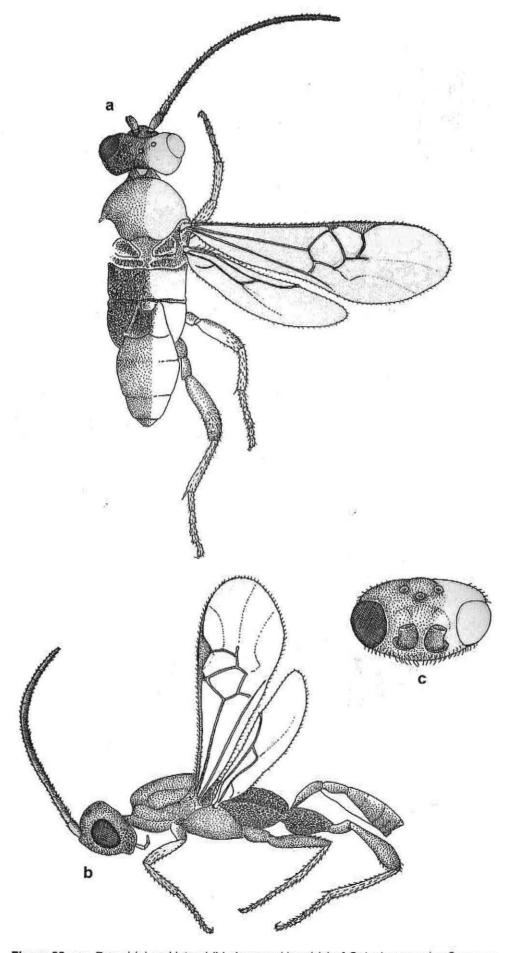


Figure 82 a-c. Dorsal (a) and lateral (b) views and head (c) of Cotesia sesamiae Cameron

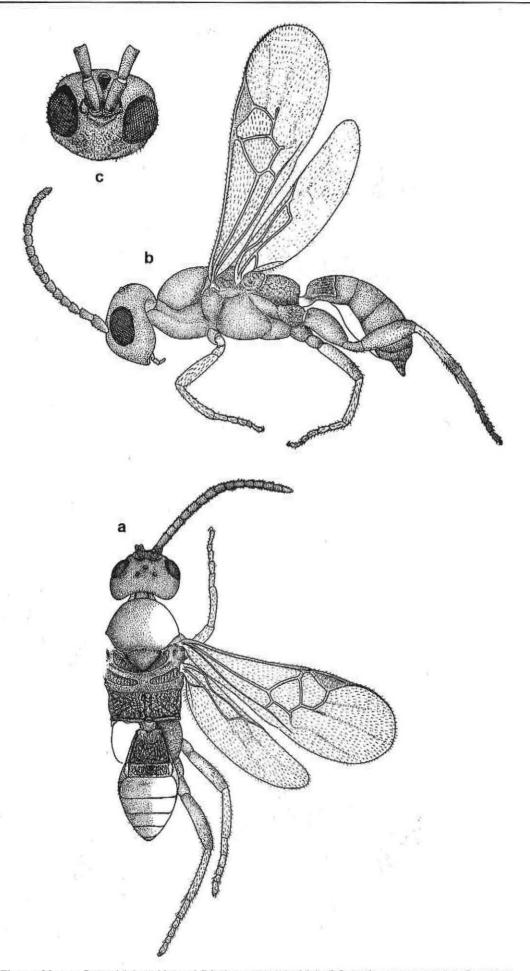


Figure 83 a-c. Dorsal (a) and lateral (b) views and head (c) of Cotesia near sesamiae Cameron

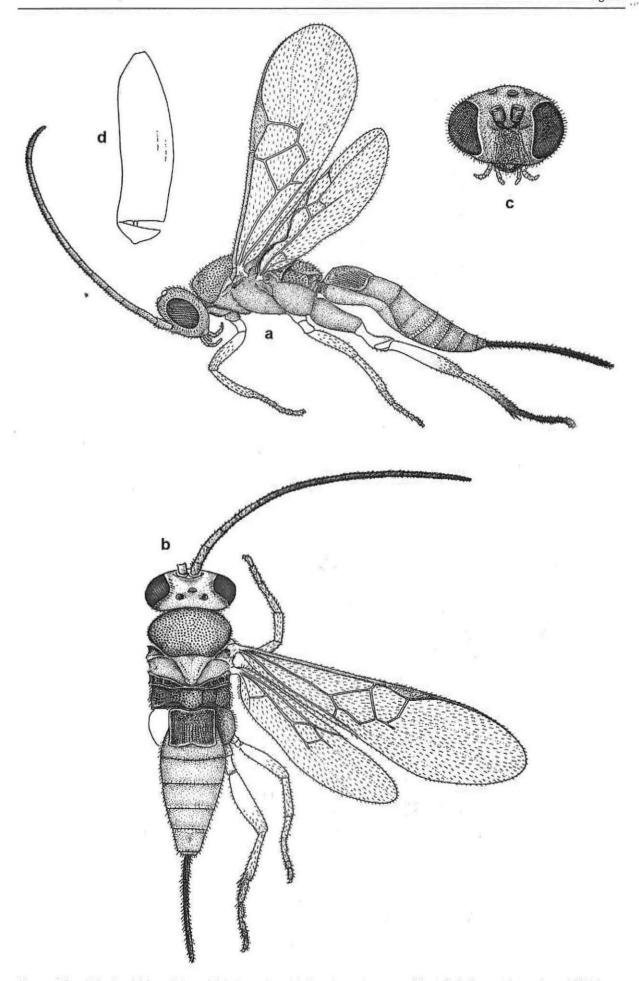


Figure 84 a-d. Lateral (a) and dorsal (b) views, head (c) and pupal cocoon (d) of Dolichogenidea polaszeki Walker

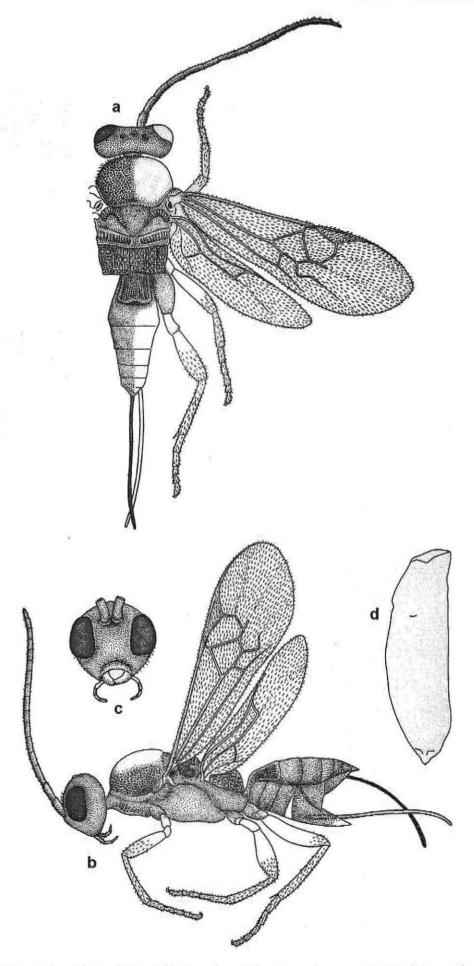


Figure 85 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Dolichogenidea sp. C

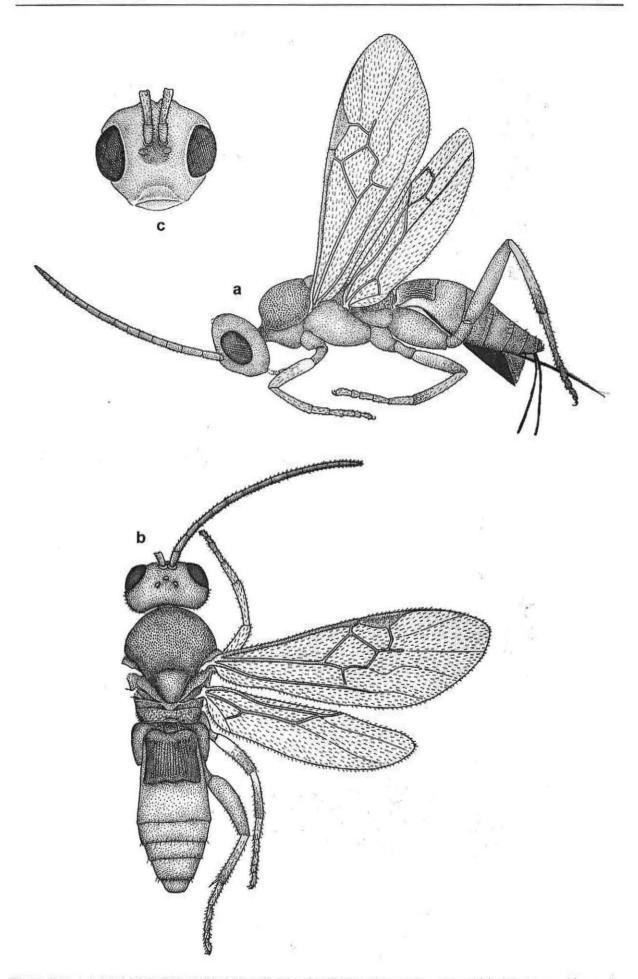


Figure 86 a-c. Lateral (a) and dorsal (b) views with ovipositor hidden underneath and head (c) of Dolichogenidea sp. A

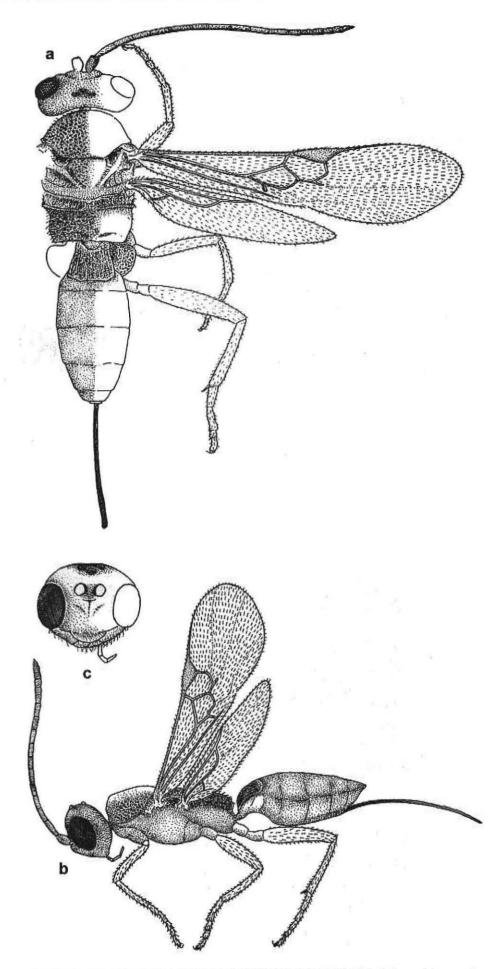


Figure 87 a-c. Dorsal (a) and lateral (b) views and head (c) of Dolichogenidea sp. B

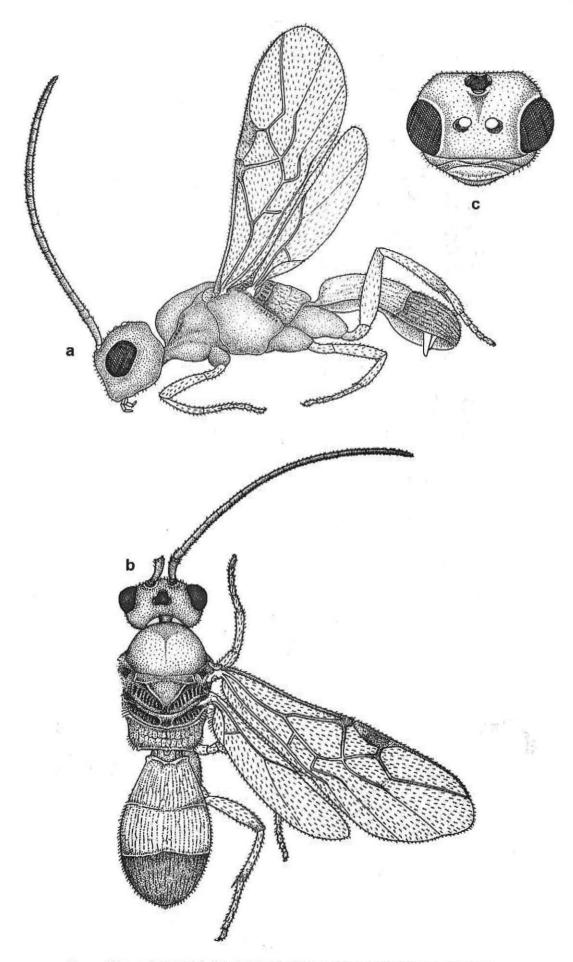


Figure 88 a-c. Lateral (a) and dorsal (b) views and head (c) of Phanerotoma sp.

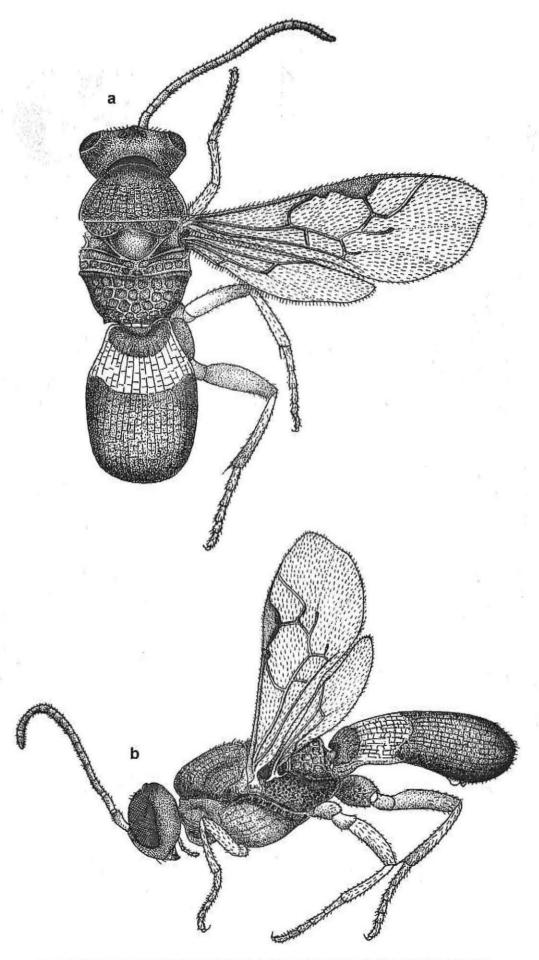


Figure 89 a-b. Dorsal (a) and lateral (b) views of Chelonus curvimaculatus Cameron

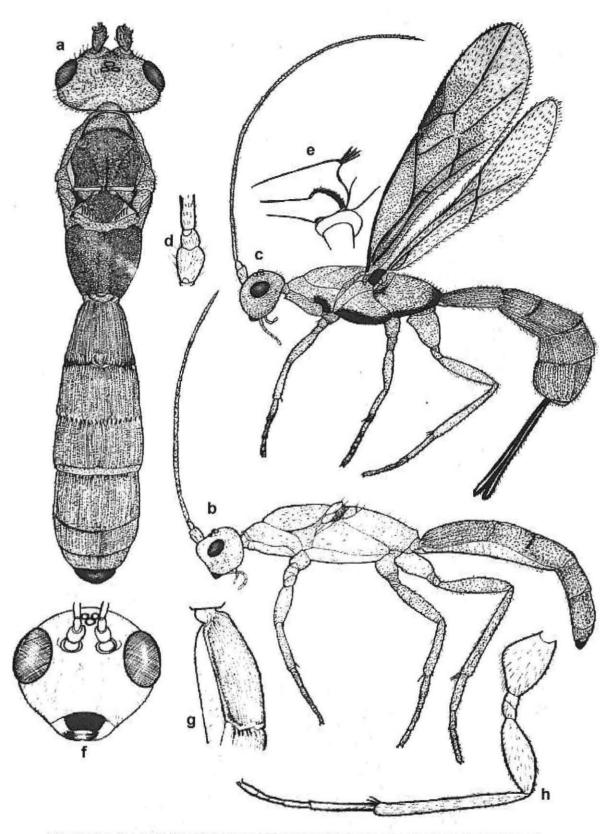


Figure 90 a-h. Dorsal (a) and lateral views of male (b) and female (c), scape and pedicel (d), coxa I and propleuron (e), head (f), side of T1 + 2 (g) and leg III (h) of *Rhaconotus* sp. A

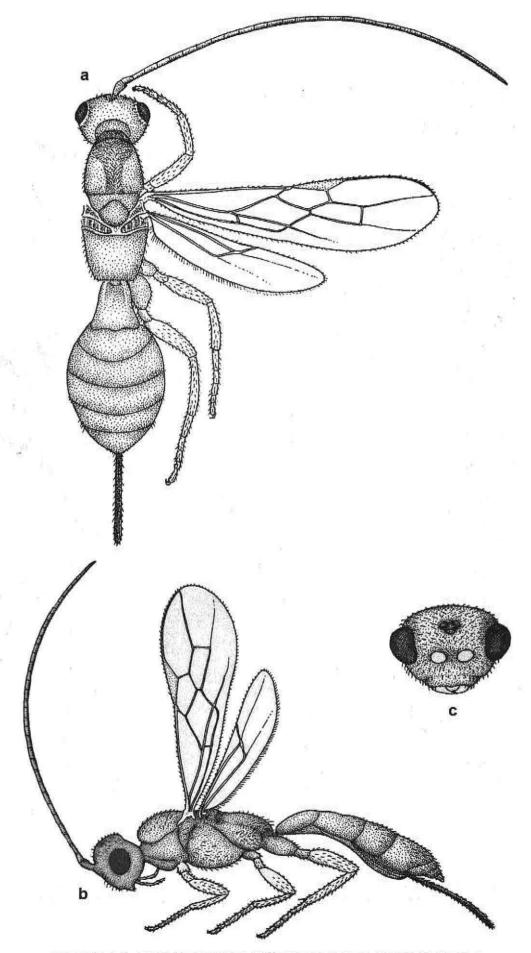


Figure 91 a-c. Dorsal (a) and lateral (b) views and head (c) of Rhaconotus sp. B

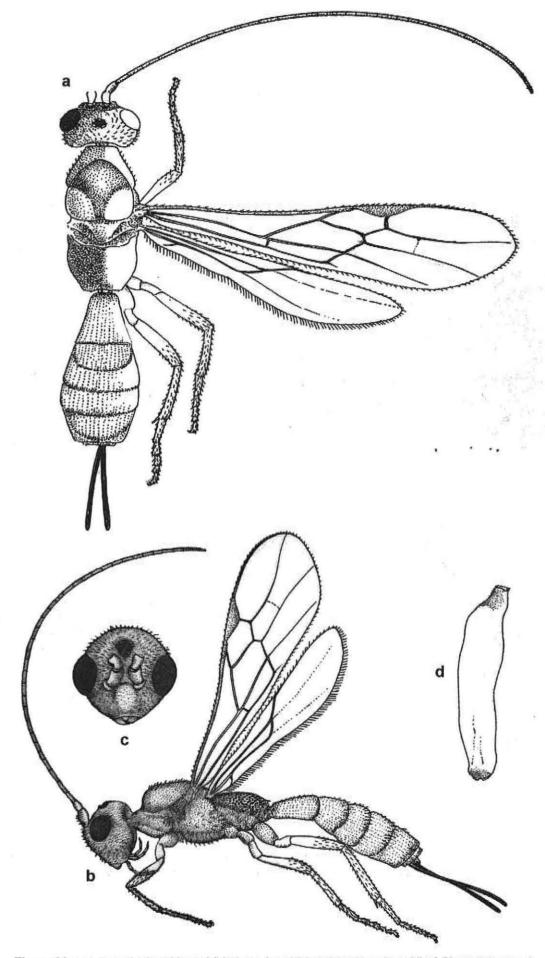


Figure 92 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Rhaconotus sp. C

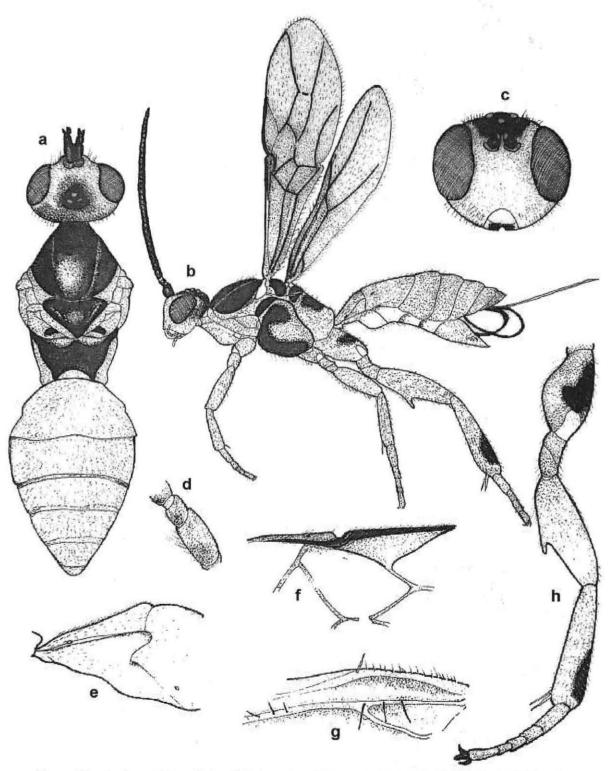


Figure 93 a-h. Dorsal (a) and lateral (b) views, head (c), scape and pedicel (d), side of tergite I (e), stigma and first submarginal cell (f), hindwing venation (g) and leg III (h) of *Habrobracon* sp. A

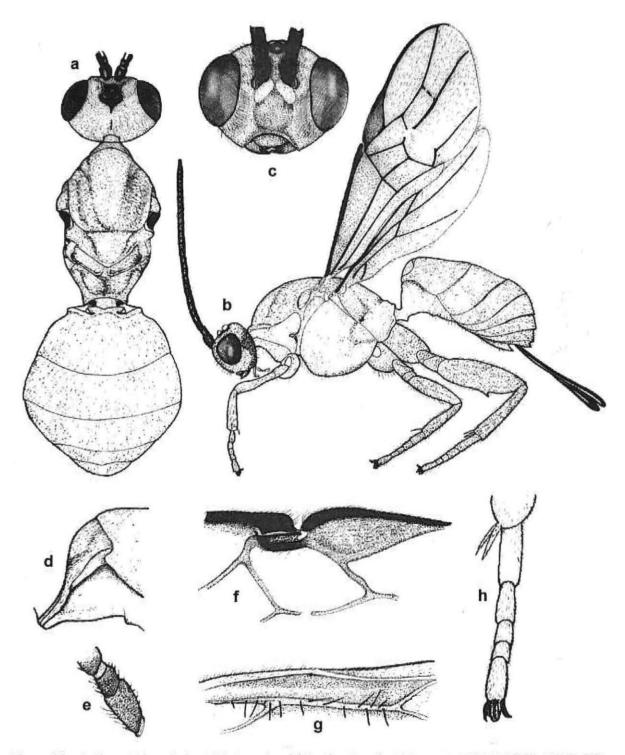


Figure 94 a-h. Dorsal (a) and lateral (b) views, head (c), side of tergite I (d), scape and pedicel (e), stigma and first submarginal cell (f), hindwing venation (g) and tip of tibia and tarsus of leg III (h) of Habrobracon sp. B

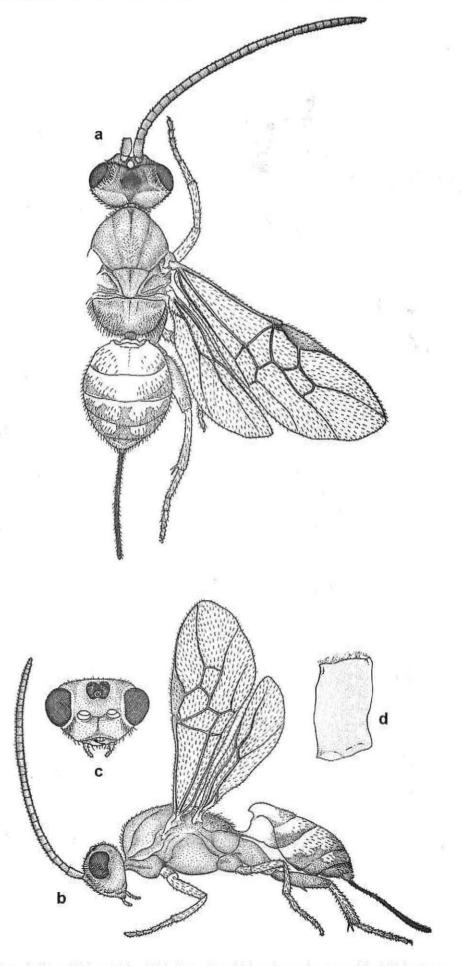


Figure 95 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Habrobracon sp. C

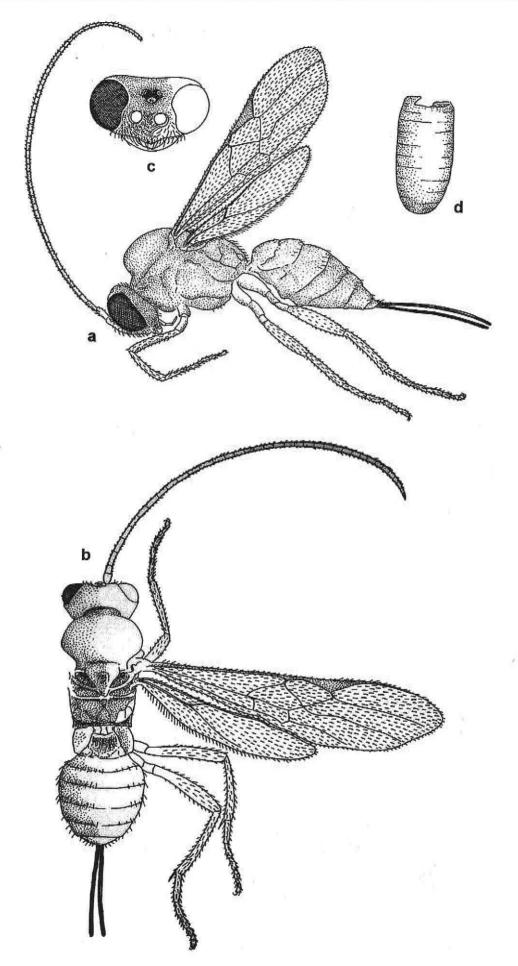


Figure 96 a-d. Lateral (a) and dorsal (b) views, head (c) and pupa (d) of host of Bracon sp.

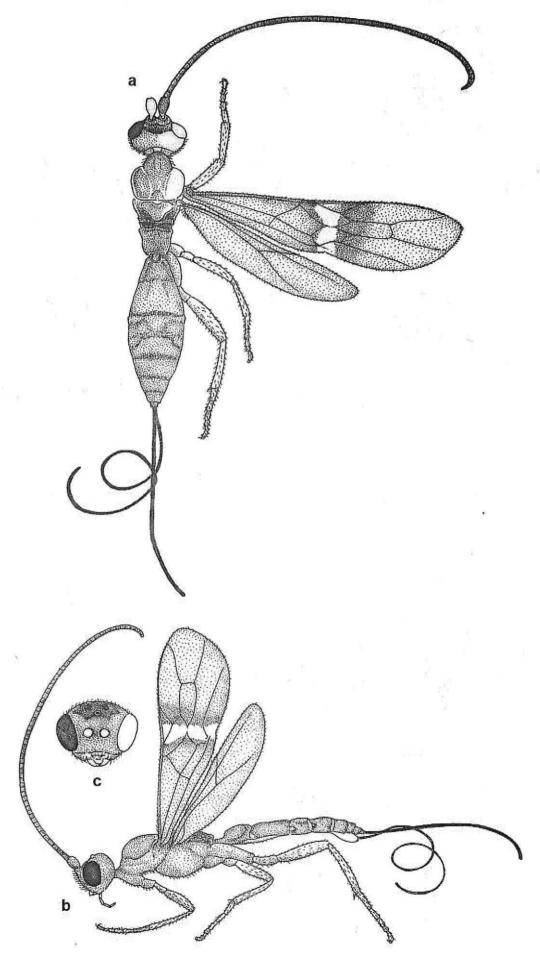


Figure 97 a-c. Dorsal (a) and lateral (b) views and head (c) of Stenobracon rufus Szepligèti

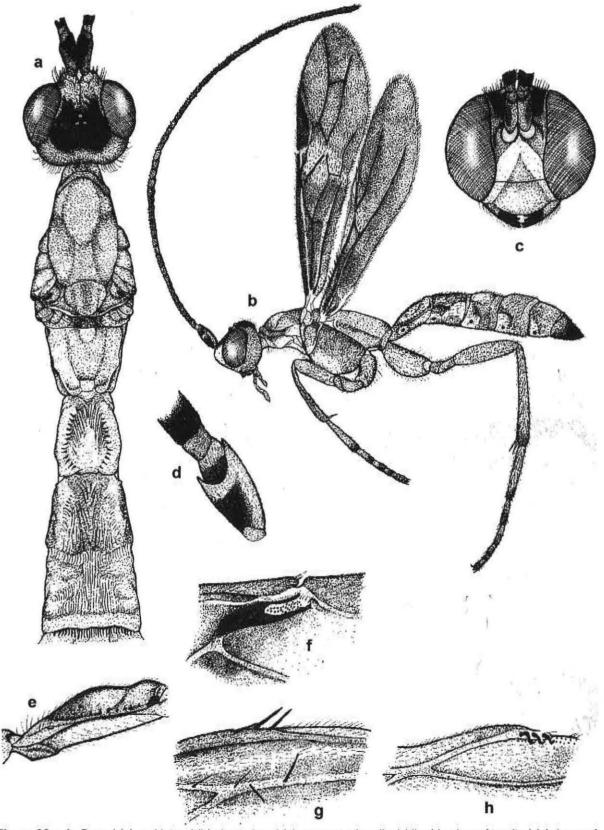


Figure 98 a—h. Dorsal (a) and lateral (b) views, head (c), scape and pedicel (d), side view of tergite I (e), base of stigma and inner part of submarginal cell (f), hairs on hindwing (g) and shape of hamuli (h) of Stenobracon sp.

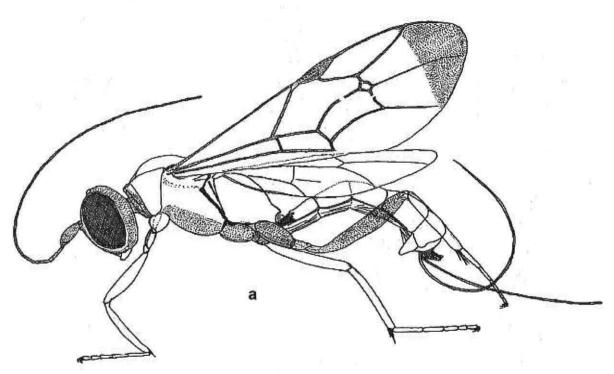


Figure 99 a. Lateral view of Syzeuctus sp.

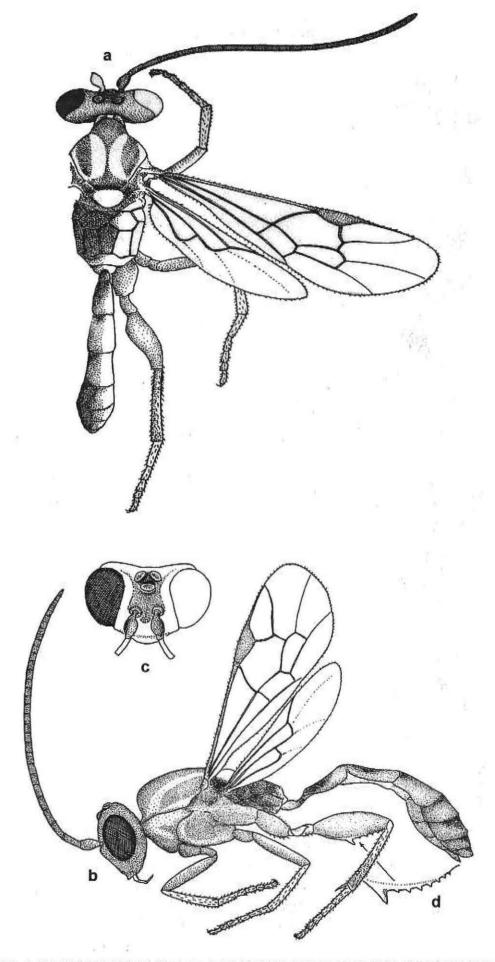


Figure 100 a-d. Dorsal (a) and lateral (b) views, head (c) and spines on femur III (d) of Pristomerus bullis Fitton

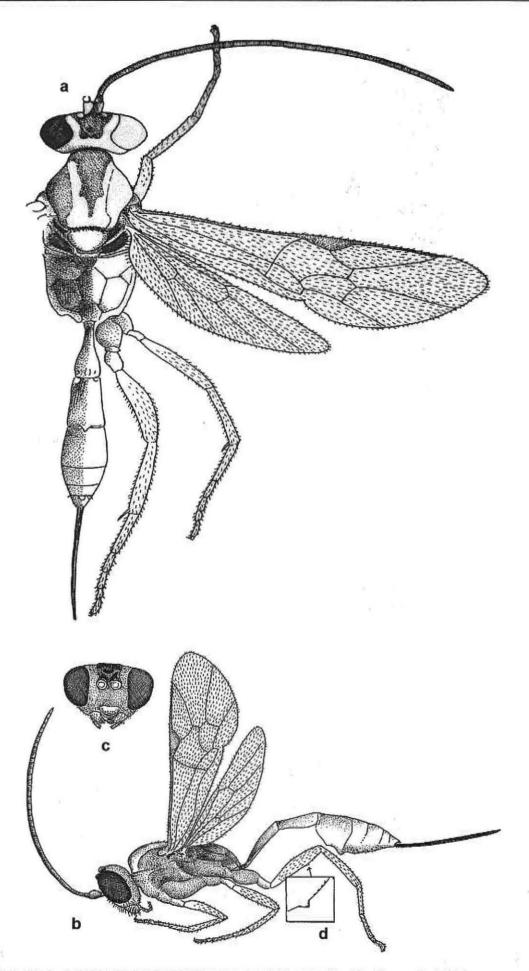


Figure 101 a-d. Dorsal (a) and lateral (b) views, head (c) and spine (d) of femur III of Pristomerus sp. A

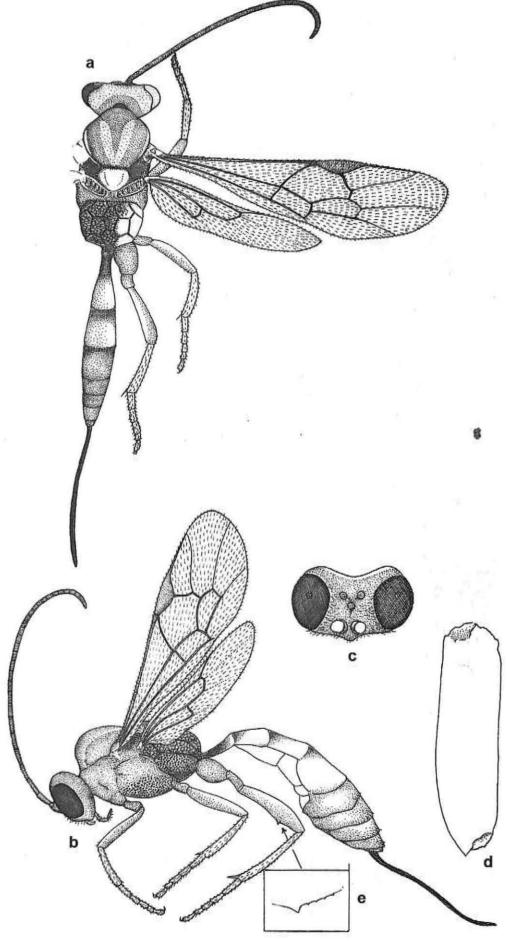


Figure 102 a-e. Dorsal (a) and lateral (b) views, head (c), pupal cocoon (d) and ventral spines on femur III (e) of *Pristomerus* sp. B

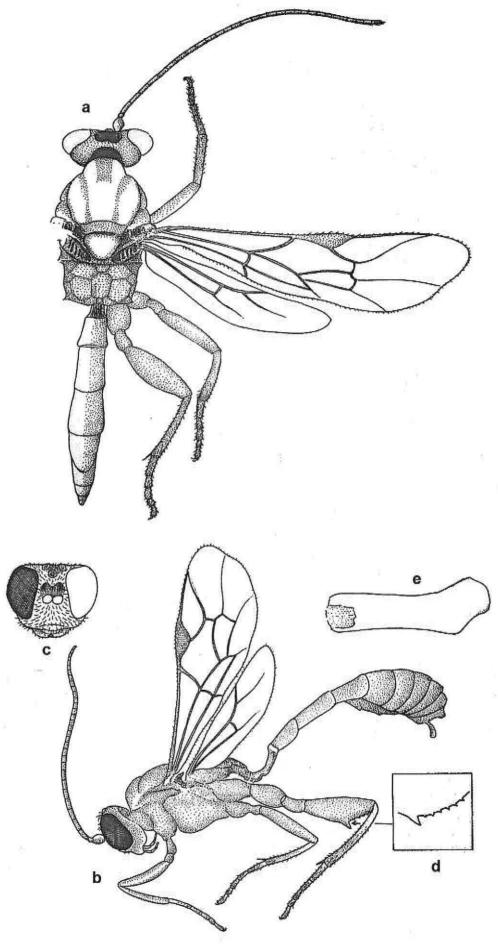


Figure 103 a-e. Dorsal (a) and lateral (b) views, head (c), ventral spines in femur III (d) and pupal cocoon (e) of *Pristomerus* sp. C

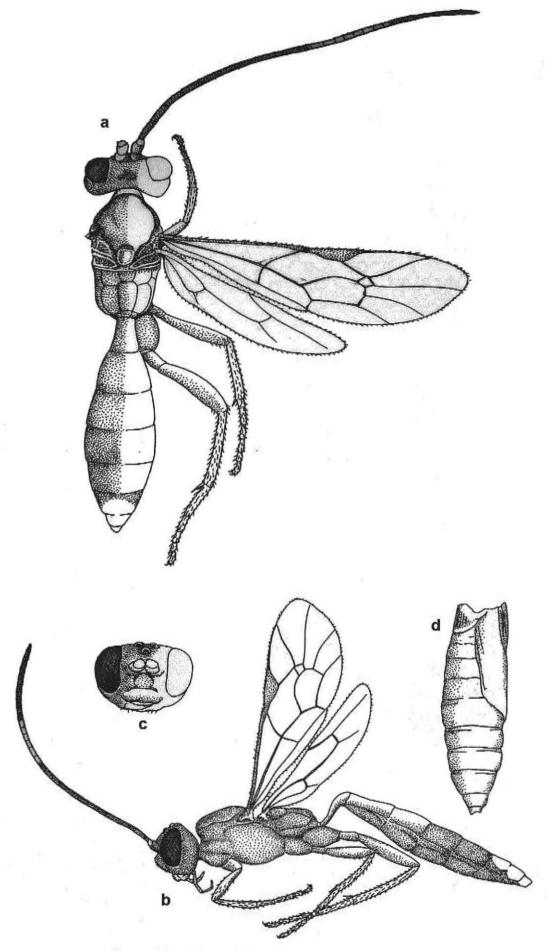


Figure 104 a-d. Dorsal (a) and lateral (b) views and head (c) of Dentichasmias busseolae Heinrich, and pupa (d) of its host

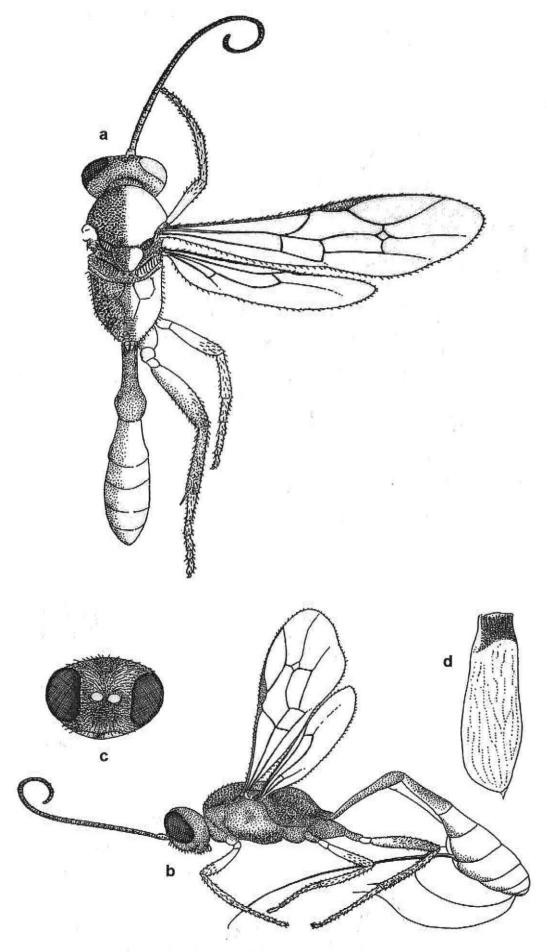


Figure 105 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Venturia sp. A

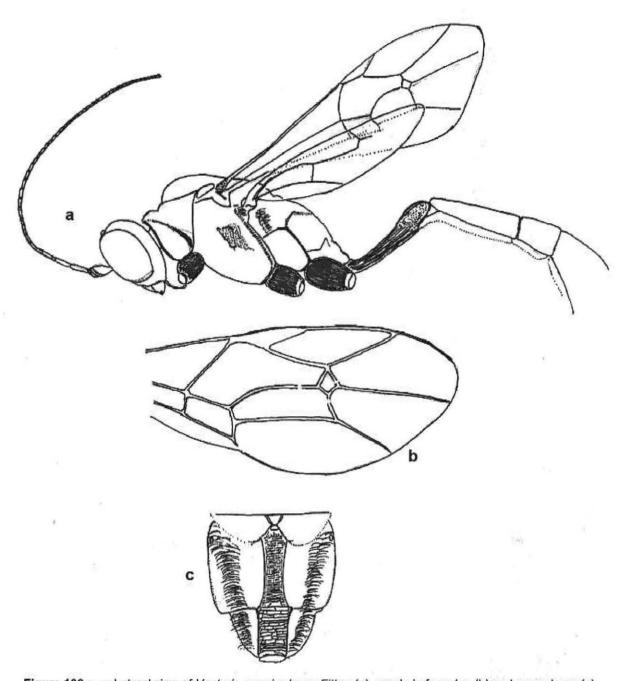


Figure 106 a-c. Lateral view of Venturia near jordanae Fitton (a), areole in forewing (b) and propodeum (c)

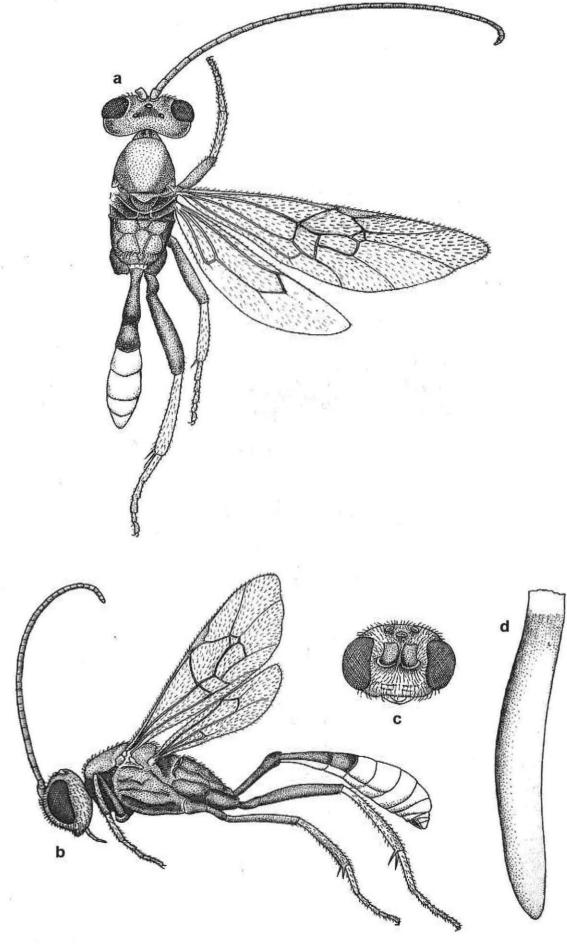


Figure 107 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Venturia sp. B

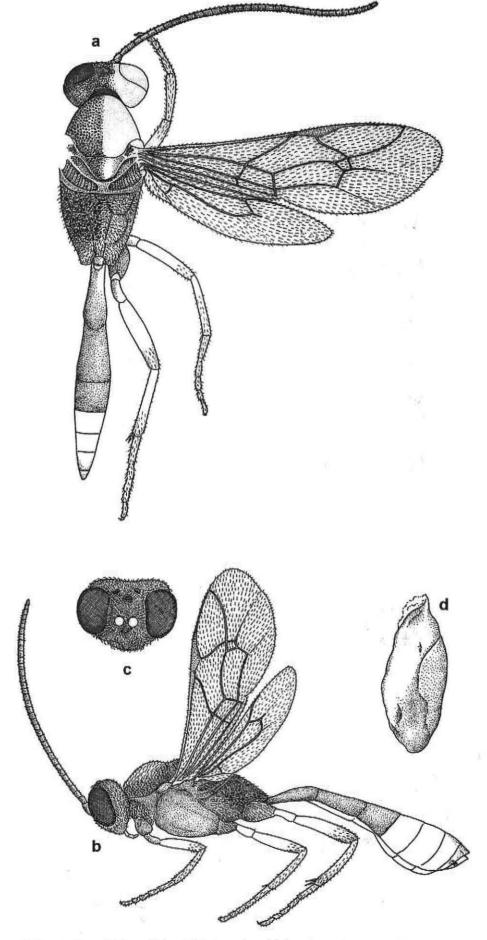


Figure 108 a-d. Dorsal (a) and lateral (b) views, head (c) and pupal cocoon (d) of Venturia sp. C

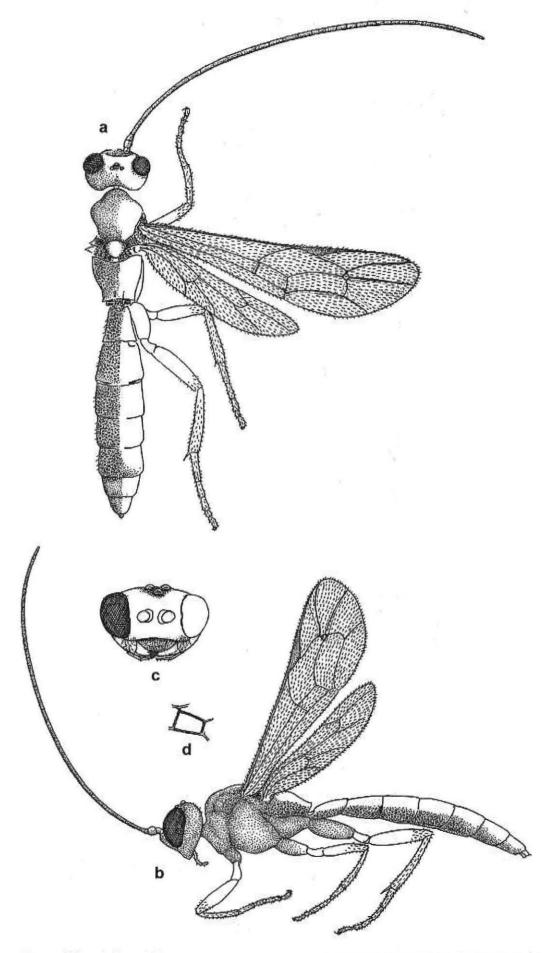


Figure 109 a-d. Dorsal (a) and lateral (b) views, head (c) and areole (d) of Holcopimpla sp. A

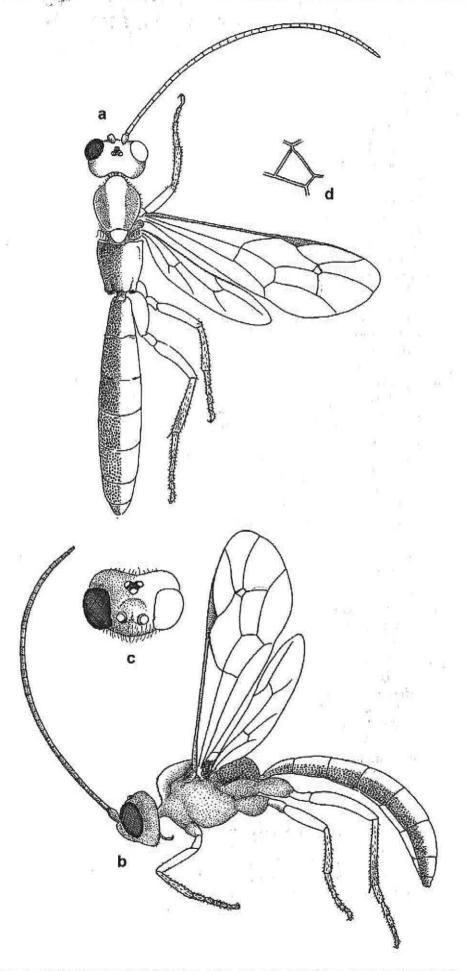


Figure 110 a-d. Dorsal (a) and lateral (b) views, head (c) and areole (d) of Holcopimpla concolor (Brullé)

APPENDIX—LIST OF STEMBORERS AND PARASITOIDS REARED FROM GRASSES

STEMBORERS

```
Order: Lepidoptera
     Family: Cossidae
          Phragmataecia boisduvalii (Figure 14 a-l)
     Family: Gelechiidae
          Stegasta sp. (Figure 4 a-l)
     Family: Noctuidae
          Busseola fusca (Plates 19, 20, 28, 29; Figure 13 a-m)
          Busseola obliquifascia (Plates 19, 20, 28)
          Busseola phaia (Plates 19, 20, 28)
          Manga nubifera (Plates 21, 22, 27; Figure 12 a-i)
          Sciomesa piscator (Plates 10, 27)
          Sciomesa cf. sp. nov. (Plate 18)
          Sesamia calamistis (Plates 11, 13, 14, 26, 29; Figure 11 a-q)
          Sesamia nonagrioides botanephaga (Plates 11, 15, 26; Figure 10 a-f)
          Sesamia penniseti (Plates 11, 16, 26)
          Sesamia poephaga (Plates 11, 14, 17, 26)
          Sesamia sp. nov. (Plates 11, 12, 26)
     Family: Pyralidae
          Chilo incertus (Plate 2)
          Chilo orichalcociliellus (Figure 7a-g)
          Chilo partellus (Plates 1, 3, 23, 29; Figure 6 a-u)
Chilo thyrsis (Plates 1, 3, 23; Figure 8 a-o)
          Eldana saccharina (Plates 1, 2, 23; Figure 5 a-r)
          Ematheudes sp. nov. (Plates 7, 8, 25)
          Ematheudes straminella (Plates 7, 8, 25)
          Maliarpha concinnella (Plates 6, 24)
          Peoriinae taxon A (Plates 4, 5, 24)
          Peoriinae taxon B (Plates 9, 24)
          Saluria léntistrigella (Plates 10, 25)
    Family: Tortricidae
          Thaumatotibia leucotreta (Figure 9 a-f)
Order: Coleoptera
    Family: Anthribidae
          Phloebius sp. A (Figure 16 a-c)
          Phloebius sp. B (Figure 17 a-c)
    Family: Cerambycidae
          Hyllisia near vittata (Figure 34 a-d)
          Hypamazso pauli (Figures 29 a-w, 30 a-i)
          Hypamazso sp. B (Figure 31 a-o)
          Hypamazso sp. C (Figure 32 a-k)
          Oheneopsis sp. (Figure 33 a-y)
    Family: Curculionidae
          ?Tanymecus sp. (Figure 23 a-i)
          ?Odoiporus sp. (Figure 18 a-g)
          Lixus near germaini (Figure 20 a-1)
          Lixus sp. A (Figure 19 a-i)
          Tanymecus near dilaticollis (Figure 21 a-c)
          Tanymecus sp. A (Figure 22 a-o)
```

Unknown genus sp. A (Figure 24 a–z4) Unknown genus sp. B (Figure 25 a–p) Unknown genus sp. C (Figure 26 a–i) Unknown genus sp. D (Figure 27 a-e) Unknown genus sp. E (Figure 28 a-j)

Family: Languriidae

Barbaropus near olseni (Figure 44 a-m)

Barbaropus sp. B (Figure 46 a-h)

Barbaropus sp. C (Figure 45 a-h)

Barbaropus sp. D (Figure 47 a-c)

Barbaropus sp. E (Figure 48 a-h)

Promecolanguria rufocephala (Figure 49 a-i)

Stenolanguria caudata (Figure 42 a-n)

Stenolanguria sp. (Figure 43 a-1)

Family: Mordellidae

Stenalia near occidentalis (Figure 35 a-q)

Stenalia sp. B (Figure 36 a-f)

Stenalia sp. C (Figure 37 a-g)

Stenalia sp. D (Figure 38 a-e)

Stenalia sp. E (Figure 39 a-e)

Family: Tenebrionidae

Paramarygmus sp. (Figure 40 a-i)

Zophodes sp. (Figure 41 a-g)

Order: Diptera (Figure 50 a-d)

Family: Chloropidae

Elachiptereicus abessynicus (Figure 54 a-d)

Mepachymerus sp. (Figure 53 a-d)

Pachylophus sp. (Figure 55 a-d)

Family: Diopsidae

Diopsina sp. (Figure 51 a-d)

Diopsis near lindneri (Figure 52 a-c)

Diopsis sp. B (Figure 52 d)

Family: Muscidae

Atherigona soccata (Figure 57 a-h)

Family: Tephritidae

Bistrispinaria fortis (Figure 56 a-b)

Bistrispinaria magniceps

Bistrispinaria woodi

PARASITOIDS

Order: Diptera

Family: Tachinidae

Descampsina sesamiae (Figure 62 a-e)

Leskia sp. (Figure 63 a-g)

Lydella nr sesamiae (Figure 61 a-g)

Siphona sp. A (Figure 59 a-e)

Siphona sp. B (Figure 60 a-f)

Order: Hymenoptera (Figure 58 a-f)

Family: Bethylidae

Goniozus indicus (Figure 79 a)

Family: Braconidae

Bracon sp. (Figure 96 a-d)

Chelonus curvimaculatus (Figure 89 a-b)

Cotesia flavipes (Figure 81 a-c)

Cotesia nr sesamiae (Figure 83 a-c)

Cotesia sesamiae (Figure 82 a-c)

Dolichogenidea polaszeki (Figure 84 a-d)

Dolichogenidea sp. A (Figure 86 a-c)
Dolichogenidea sp. B (Figure 87 a-c)
Dolichogenidea sp. C (Figure 85 a-d)
Habrobracon sp. A (Figure 93 a-h)
Habrobracon sp. B (Figure 94 a-h)
Habrobracon sp. C (Figure 95 a-d)
Phanerotoma sp. (Figure 88 a-c)
Rhaconotus sp. A (Figure 90 a-h)
Rhaconotus sp. B (Figure 91 a-c)
Rhaconotus sp. C (Figure 92 a-d)
Stenobracon rufus (Figure 97 a-c)
Stenobracon sp. (Figure 98 a-h)
Triaspis sp. (Figure 80 a-c)

Family: Chalcididae

Brachymeria kassalensis (Figure 66 a-c) Hockeria sp. (Figure 67 a-d)

Family: Eulophidae

Pediobius furvus (Figure 71 a–c)
Pediobius homoeus (Figure 70 a–c)
Tetrastichus sp. (Figure 69 a–c)

Family Eupelmidae

Macroneura sp. (Figure 77 a-c)

Family: Eurytomidae

Eurytoma oryzivora (Figure 73 a-c)
Eurytoma sp. A (Figure 74 a-c)
Eurytoma sp. B
? Eurytoma sp. C (Figure 75 a-c)
Eurytoma sp. D (Figure 76 a-d)
Sycophila sp. (Figure 72 a-c)

Family: Ichneumonidae

Dentichasmias busseolae (Figure 104 a-d)
Holcopimpla concolor (Figure 110 a-d)
Holcopimpla sp. A (Figure 109 a-d)
Pristomerus bullis (Figure 100 a-d)
Pristomerus sp. A (Figure 101 a-d)
Pristomerus sp. B (Figure 102 a-e)
Pristomerus sp. C (Figure 103 a-e)
Syzeuctus sp. (Figure 99 a)
Venturia near jordanae (Figure 106 a-c)
Venturia sp. A (Figure 105 a-d)
Venturia sp. B (Figure 107 a-d)
Venturia sp. C (Figure 108 a-d)

Family: Mymaridae

Gonatocerus sp. (Figure 68 a-j)

Family: Platygasteridae

Inostemma sp. (Figure 65 a-c)

Family: Pteromalidae

Norbanus sp. (Figure 78 a-c)

Family: Scelionidae

Scelio sp. (Figure 64 a-c)

GLOSSARY

Aedeagus: The male copulatory organ.

Ampulla: Of the male genitalia of some moths, a process, arising from the inner face of

the valve; of coleopteran larvae, vesicular balloon or blister-like processes on

the body surface.

Annellus(i): Of moths; short, ring-like antennal segment between the pedicel and the

funicle.

Annular: Ring-shaped.

Anterior: Front of the body or body part (opposite of posterior).

Apical: Toward the end (apex) of a structure (opposite of basal).

Apophysis(es): Of the genitalia of female moths; rod-like support structures.

Area superomedia: Of Ichneumonidae; one of the areolae (spaces) on the propodeum; these spaces defined by bordering carinae. According to Heinrich, for ichneumonine ichneumonids, the area superomedia is the large median areola, set off by a transverse carina dorsally from the area basalis and from another transverse

carina ventrally from the area posteromedia.

Areola: A space defined by bordering carinae (see area superomedia).

Areole: Of moths; the extra wing cell distal to discal cell.

Areolet: Of Ichneumonidae (Hymenoptera); the small cell in the middle of the wing.

Arista: Bristle-like structure originating on the antennal flagellum of some flies.

Basal: Toward the origin (base) of a structure (opposite of apical).

Bifid: Forked or cleft into two parts. Biordinal: Of crochets, arranged in two ranks.

Bipectinate: With feather-like setae on both sides of the antenna.

Bulla(e): Sac-like evagination(s).

Bursa copulatrix: Of female moths, the anterior-most part of the genitalia; a sack-like structure

into which sperm packet (spermatophore) is deposited.

Carina: A ridge.

Chaetosema(ta): Of moths, raised cuticular area(s) on the head associated with sensory hairs

or setae, found near the compound eyes.

Chorda: Of the moth wing; the stem of veins R₄₊₅.

Ciliate: With small hairs.

Clasper: Of male moths, the valve or a specialised grasping structure on the inner

surface of the valve.

Clavate: Club-shaped.

Clypeus: The part of the insect head just above the mouthparts.

Coarctate: Of a pupa, where the developing adult is hidden inside the 'case' formed

from the last larval skin (e.g. in the higher Diptera).

Coremata: Of male moths; secondary structures typically involved in the dissemination

of pheromone; can be as simple as dense patches of specialised scales or setae

to complicated eversible balloon-like organs.

Corneous: Horn-like.

Cornutus(i): Sclerotised projection(s) on the vesica of the aedeagus of some moths.

Coronal suture: Longitudinal suture along midline of the head, from the epicranial notch to

the apex of the clypeus.

Costa: (a) The vein, when present, along the edge of the wing, usually complete along the anterior margin; along the rest of the wing its presence and strength

highly variable among different insect groups;

(b) Of male moths, the dorsal part of the valve (= harpe).

Coxa: The most basal of the five leg segments of insects.

Cremaster: An attachment organ at the posterior of a moth pupa, comprising a group of

small hooked sclerites.

Crochets: Small, hook-like sclerotisations on the caterpillar (larva) prolegs.

Cubitus: The 5th longitudinal vein of the wing.

Cucullus: The distal portion of the male valva or clasper.

Denticle(s): Small tooth-like process(es).

Digitate: Finger-like.

Distal: Far away from (opposite of proximal).

Dorsal: The upper surface (opposite of ventral).

Ductus bursae: The duct in female moths connecting the ostium to the bursa copulatrix.

Eclosion: Emergence of the adult from the pupa.

Elytron(a): The sclerotised forewings of Coleoptera (beetles).

Epicranial stem: (= Coronal suture) longitudinal suture on the midline of the head, reaching

from the epicranial notch to the apex of the clypeus.

Epicranial suture: A U-, V-, or Y-shaped suture on the dorsal part of the head.

Epiphysis: Lobe-like process on the fore tibia of some moths.

Exarate: Of a pupa, where the appendages of the developing adult are free to move

(e.g. in Coleoptera and Hymenoptera).

Face: That part of the insect head between the clypeus and the antennal insertions.

Femur: The third leg segment, usually the most robust.

Filiform: Thin, like a wire.

Flagellum: Segments of the flagellum of an antenna.

Flagellum: The part of the antenna distal to the pedicel.

Frenulum: One or more spines on the forward edge of the hindwing of a moth that

engage the forewing, synchronising wing action during flight.

Frons: That part of the insect head between the face and the vertex (top).

Funicle: The group of antennal segments between the annellus (see above) and the

club.

Funicular: Pertaining to the funicle.

Gaster: Of Hymenoptera; the portion of the abdomen behind the petiole.

Gena: The side of the head below and continuing behind the eyes.

Geniculate: Bent like an elbow.

Gula: Sclerite forming the central part of the ventral (under) side of the head.

Gular: Pertaining to the gula.

Gnathos: Paired sclerotised processes of the vinculum.

Harpe: See valve. Hyaline: Translucent.

Hypognathous: Head oriented more or less vertically, mouthparts pointing down.

Infuscate: Dark, smoky brown.

Juxta: Plate-like structure on which the aedeagal manica rests.

Longitudinal: Running lengthwise along the body or structure.

Medial: Towards or near the middle of a structure.

Notauli: Of Hymenoptera; a more or less straight depression or furrow on the

mesoscutum, sometimes complete, often punctuate, usually arranged

obliquely.

Obtect: Of a pupa, where the appendages adhere to the body (e.g. in moths)

Occiput: Posterior-most part of the head, behind and below the vertex.

Ocellar triangle: More or less defined area of frons on which ocelli are found.

Ocellus(i): Simple eye; single facet, light sensitive structure on the head.

Ostium: Of Lepidoptera, the external opening of the female genitalia.

Ostium bursae: Of the female genitalia of moths, the opening connecting the bursa copulatrix

to the ductus bursae.

Ovipositor: Egg-laying organ.

Patagium: Pad-like structure with hairs or scales located on the thorax of moths.

Pecten: In some moths, elongate scales on ventral surface of the scape.

Pectinate: Comb-like.

Pedicel: The 2nd antennal segment.
Penellipse: An incomplete circle or ellipse.

Penultimate: Next to last.

Petiole: In wasps, the more or less constricted portion of the abdomen that connects

it to the propodeum (see below), the apparent last segment of the thorax.

Pinaculum(a): Usually small, variable raised processes on the integument of the larva that

bear the primary seta(e).

Plica(e): Ridge(s).

Porrect: Palps projected forward.

Posterior: End of the body or body part (opposite of anterior).

Prolegs: Fleshy leg-like protuberances on the caterpillar abdomen.

Prognathous: Head oriented more or less horizontally, mouthparts pointing forward.

Propodeum: In Hymenoptera, the apparent part of the thorax just above insertion of the

abdomen; in reality, the first abdominal segment.

Prothoracic shield: The variably sclerotised dorsum of the prothorax of the larva.

Proximal: Close to (opposite of distal).

Punctate: Having more or less tiny depressions or holes.

Punctation: The pattern of the tiny depressions or holes.

Reclinate: Directed backwards.

Retinaculum: Of Lepidoptera, hook(s) or specialised scales on the posterior, ventral area of

forewing which engage the frenulum.

Rostrum: Of some beetles, the produced, snout-like part of the head.

Rugose: Wrinkled, rough.

Sacculus: Of male genitalia, the ventral part of the valve.

Saccus: Anteriorly projecting process from the vinculum or 9th abdominal segment.

Scape: The 1st antennal segment.

Sclerite: A discrete section of hardened exoskeleton.

Sclerotised: Hardened.

Scobinate: Corrugated, wavy.

Scrobe: In weevils, the groove at the side of the rostrum into which the antennae fit.

Scutellum: The sclerotised dorsal surface immediately posterior to the thorax.

Scutum: The dorsal surface of the mesothorax.

Serrate: Saw-like, with teeth.

Seta(e): Bristle(s).

Signum(a): Sclerotised structure(s) on the wall of the corpus bursae of some female

moths.

Sinuate: Long and wavy.

Spermatheca(e): In the female, a receptacle for storing sperm, usually sclerotised.

Spiracle: Opening in the insect cuticle through which gases diffuse.

Spur: Sword-like structure on the adult leg, usually at the tip of a segment. Sternite: The ventral sclerite of a body segment (e.g. an abdominal sternite).

Stigma: Of the wing; a spot-like, usually darkened area on the anterior costal

margin.

Stria(e): Fine, longitudinal line(s) or minute ridge(s).

Strigula(e): Fine, short transverse mark or line.
Suture: The fixed seam between two sclerites.

Tarsal formula: The number of segments (tarsomeres) each of the three tarsi is divided into,

starting from front to back (i.e. fore, mid and hind tarsus). For example, a tarsal formula of 5-5-4 indicates that the fore and mid tarsi each comprise five segments, while the hind tarsus has only four segments.

Tarsomere: One of the segments making up the tarsus.

Tarsus: The most apical leg segment (usually subdivided into 3-5 parts).

Tegula: A small thoracic sclerite that covers the base of the wing.

Tegumen: Part of the male genitalia of moths; the roughly triangular sclerite derived

from the ninth tergite; dorsal to, and articulating with, the vinculum.

Tergite: A dorsal sclerite of a body segment (e.g. an abdominal tergite).

Termen: The outer margin of the wing, between the apex and the posterior angle.

Tibia: The penultimate leg segment.

Torulus(i): At the base of the antenna, the socket joint allowing it to move.

Transverse: Running across the body or structure from side to side.

Triordinal: Of crochets, arranged in three ranks.

Trochanter: The second, usually small, leg segment, basal to the femur.

Tubercle: A small rounded projecting part of the insect cuticle.

Tympanum: The auditory organ of Lepidoptera, more or less round.

Tympanal organ: Membranous surface structures on the abdomen or thorax.

Uncus: Of the male genitalia of moths, the dorsal posterior projection from the

tegumen (vinculum).

Valve: Paired structures of the male genitalia, used to clasp the female during

mating.

Ventral: The lower surface (opposite of dorsal).

Vermiform: Thin and wormlike. Verrucae: Wart-like projections.

Vesica: Membrane surrounding the aedeagus of male moths.

Vinculum: Of moths, the sclerotised ring of the male genitalia formed of the combined

9th and 10th fused tergites and sternites; sometimes referring to the usually

U- or V-shaped latero-ventral portion.

Some definitions from: http://www.ndsu.edu/ndsu/ndmoths/glossary_h.htm.

Published by the Department of Entomology, North Dakota State University, and some adapted from Gordh and Headrick (2001).

SELECTED REFERENCES

Barrion A. T. and Khan Z. R. (2004) How to Handle Grass Stemborer Larvae. icipe Science Press, Nairobi. 37 pp.

Barrion A. T., Copeland R. S. and Khan Z. R. (2007) Comment mener à terme un élevage de

foreurs? icipe Science press, Nairobi. 30 pp.

Bezzi M. (1924) Further notes on the Ethiopian fruitflies with keys to all known genera and species. Bulletin of Entomological Research 15, 73-188. Bleszynski S. (1962) Studies on the Crambidae. Part 36. Notes on some species of the genus Chilo

Zincken. Acta Zoologica Cravov. 7, 107-135.

Bleszynski S. (1970) A revision of the world species of Chilo Zincken (Lepidoptera: Pyralidae). Bulletin of the British Museum (Natural History): Entomology 25, 99-195.

Booth R. G., Cox M. L. and Madge R. B. (1990) International Institute of Entomology. Guides to Insects of Importance to Man. 3. Coleoptera. IIE and The Natural History Museum, The University Press, Cambridge, 384 pp.

Borror D. J., Triplehorn C. A. and Johnson N. F. (1989) An Introduction to the Study of Insects. 6th

edition. Saunders College Publishing. 875 pp.

Boucek Z. (1988) Australasian Chalcidoidea (Hymenoptera). A Biosystematic Revision of Genera of Fourteen Families, with a Reclassification of Species. CAB International Institute of Entomology, Wallingford, UK. 832 pp.

Bowden J. (1956) New species of African stem-boring Agrotidae. Bulletin of Entomological Research

47, 415-428.

- Cook M. (1997) Revision of the genus Maliarpha (Lepidoptera: Pyralidae) based on adult morphology with description of three new species. Bulletin of Entomological Research 87, 25-
- Crosskey R. W. (1984) Annotated keys to the genera of Tachinidae (Diptera) found in tropical and southern Africa. Annals of Natal Museum 26, 189-337.
- CSIRO (1979) The Insects of Australia: A Textbook for Students and Research Workers. Melbourne Press, Australia. 1029 pp.

Davies J. C. and Seshu Reddy K. V. (1981) Shootfly species and their graminaceous hosts in Andhra Pradesh, India. Insect Science and Its Application 2, 33-37.

Deeming J. C. (1973) A review of the taxonomy of African shoot flies of sorghum, pp. 3-26. In Control of Sorghum Shoot Fly (Edited by M. G. Jotwani and W. R. Young). Oxford and IBH Publishing Co., New Delhi, Bombay and Calcutta, 324 pp.

Delvare G. (1988) Revision des Eurytoma (Hymenoptera: Eurytomidae) d'Afrique occidentale

decrits par Risbec. Annales de la Societe Entomologique de France (NS) 24, 117-149.

- Eady R. D. (1974) The present state of nomenclature of wing venation in the Braconidae (Hymenoptera): Its origins and comparison with related groups. Journal of Entomology (B) 43,
- Feijen H. R. (1978) Diopsidae (Diptera: Acalyptratae) from Togo and Zaire. Stuttgarter Museum fur Naturkunde in Stuttgart (ser A) Biologie 318, 1-5.

Gahan C. J. (1890) Longicornia from Africa and Madagascar. Transactions Entomology Nyasaland Society London 1890, 326-327.

Gauld I. D. and Bolton B. (1988) The Hymenoptera. British Museum (Natural History) London. Oxford University Press. 332 pp.

Gordh G. and Headrick D. (2001) A Dictionary of Entomology. CAB International, Oxon, UK. 1032

Gordh G. and Moczar L. (1990) A catalog of the world Bethylidae (Hymenoptera: Aculeata).

Memoirs of the American Entomological Institute 46, 1–364.

- Graham M. W. R. de V. (1991) A reclassification of the European Tetrastichinae (Hymenoptera: Eulophidae): Revision of the remaining genera. Memoirs of the American Entomological Institute 49, 1-322.
- Gupta T. S. and Crowson R. A. (1971) A review of the classification of the family Languriidae and the place of Languriidae in the natural system of Clavicornia. Memoirs of Indian Museum
- Hancock D. L. (1999) Grass-breeding fruit flies and their allies of Africa and Asia (Diptera: Tephritidae: Ceratitidinae). Journal of Natural History 33, 911–948.
- Harris K. M. (1962) Lepidopterous stem borers of cereals in Nigeria. Bulletin of Entomological Research 53, 139-171.

Holloway J. D., Bradley J. D. and Carter D. J. (1987) CIE Guides to Insects of Importance to Man. 1. Lepidoptera. CAB International Institute of Entomology and The British Museum of Natural History, Wallingford. UK. 262 pp.

 $Horstmann\,K. (1990)\,Die\,west palaarktichen\,Arten\,gattung\,\textit{Pristomerus}\,Curtis\,1836\,(Hymenoptera:$

Ichneumonidae). Entomofauna Zeitschrift für Entomologie 11, 9-44.

Ingram W. R. (1958) The lepidopterous stalk borers associated with Gramineae in Uganda. Bulletin of Entomological Research 49, 367–383.

Kalshoven L. G. E. (1981) *Pests of Crops in Indonesia*. Revised and translated by P.A. van der Laan with the assistance of G. H. L. Rothschild. P.T. Ichtiar Baru-Van Hoeve, Jakarta. 701 pp.

Khan Z. R., Pickett J. A., Wadhams L. and Muyekho F. (2001) Habitat management for the control of cereal stem borers in maize in Kenya. *Insect Science and Its Application* 21, 375–380.

Marsh P. M. (1979) Family Braconidae, pp. 144–295. In Catalog of Hymenoptera in America North of Mexico, Volume 1 (Edited by K. V. Krombein, P. D. Hurd Jr., D. R. Smith and B. D. Burks). Smithsonian Institution Press, Washington, DC.

Marsh P. M., Shaw S. R. and Wharton R. A. (1987) An identification manual for the North American genera of the family Braconidae (Hymenoptera). The Entomological Society of Washington 13,

1 - 98.

Moyal P. and Le Ru B. (2006) From population to species: Morphological and molecular diversity in East African stem borer species of the genus *Manga* Bowden 1956 (Lepidoptera: Noctuidae). *Annales de la Société Entomologique de France* (n.s.) 42, 293–307.

Nye I. W. B. (1960) The insect pests of graminaceous crops in East Africa. Colonial Research Studies

Ohara J. E. (1989) Systematics of the genus group taxa of the Siphonini. *Quaestiones Entomologicae* 25, 1–229.

Pinhey C. G. (1975) Moths of Southern Africa. Descriptions and Colour Illustrations of 1183 Species (XII). Balkema, Rotterdam. 273 pp.

Polaszek A. (1992) Cereal stem borers and their parasitoids of Africa. Proceedings of the Section. Experimental and Applied Entomology of the Netherlands Entomological Society 3, 70–71.

Polaszek A. (1998) (Ed.) African Cereal Stem Borers: Economic Importance, Taxonomy, Natural Enemies and Control. CAB International, Oxon, UK. 530 pp.

Pont A. C. and Magpayo F. R. (1995) Muscid shoot-flies of the Philippine Islands (Diptera: Muscidae, genus Atherigona Rondani). Bulletin of Entomological Research, Supplement (Series) 3, 1–123.

Prinsloo G. L. (2000) The Families of Parasitoids Associated with Cereal Stem Borers in Southern Africa—A Workshop Manual. ARC—Plant Protection Research Institute. 57 pp.

Schmutterer H. (1969) Pests of Crops in Northeast and Central Africa. Gustav Fischer Verlag. Stuttgart. 296 pp.

Scholtz C. H. and Holm E. (1985) Insects of Southern Africa. Butterworth Publishers (Pty) Ltd., Durban. 502 pp.

Shaffer J. C. (1968) A revision of the Peoriinae and Anerastiinae of America North of Mexico (Lepidoptera: Pyralidae). Bulletin of the United States National Museum 280, 1–124.

Shaffer J. C. (1997) A preliminary revision and cladistic analysis of the Ethiopian species of Ematheudes (Lepidoptera: Pyralidae: Phycitinae: Peoriini). Entomologica Scandinavica 28, 403– 444.

Shillito J. F. (1971) The genera of Diopsidae. Zoological Journal of the Linnean Society 50, 287–295.
Steyskal G. C. (1972) A catalogue of species and key to the genera of family Diopsidae. Stuttgarter Beiträge Naturkunde 234, 1–20.

Tams W. H. T. and Bowden J. (1953) A revision of the African species of Sesamia Guenée and related genera (Agrotidae: Lepidoptera). Bulletin of Entomological Research 43, 645–678.

Townes H. and Townes M. (1973) A catalogue and reclassification of the Ethiopian Ichneumonidae. Memoirs of the American Entomological Society 19, 1–416.

Ullyett G. C. (1935) Notes on Apanteles sesamiae Cameron, a parasite of the maize stalk borer (Busseola fusca Fuller) in South Africa. Bulletin of Entomological Research 26, 253–262.

Villiers A. (1961) Revision des coleopteres languriides africains. Annales du Musee Royal d'Afrique Centrale 98, 1–385.

Villiers A. (1962) Coleopteres languriides africains. Bulletin de l'Institute Français de l'Afrique Noire (ser A) 24, 360–394.

Woodley N. E. (1994) A new species of *Lydella* from Mexico with a discussion of the definition of the genus. *Bulletin of Entomological Research* 84, 131–136.

About this book

Stemborers of cereal crops (such as maize, sorghum and millet) are responsible for serious losses in food production among smallholder farms in Africa. Accurate identification of insect pests of cereal crops is essential for describing the scope and importance of regional and local infestations. The most important of these pests, the moths, are also the most difficult to identify. Adults are relatively fragile, exhibit considerable variation in scale pattern, and the scales are easily rubbed off. Identification is often impossible without pristine specimens. For this reason, examination of the highly conserved characters of moth genitalia is accepted as the 'gold standard' for accurate identification of pests and their close relatives.

This manual provides high magnification photographic images of male and female genitalia of moth stemborers reared from cereal and wild grasses in Kenya. Line drawings are also provided for the moths, and for other stem-boring insects (Coleoptera, Diptera) that were reared from grasses, as well as for their parasitoids. Used together with the accompanying keys, we hope this book fills a gap by providing comprehensive tools for the identification of the insect borers of grasses and the parasitoids associated with them.

All photographic plates by R. S. Copeland except the following by Ms Karie Darrow:

Plate 1: Top to bottom, male genitalia of Chilo partellus, C. thyrsis and Eldana saccharina

Plate 2: Far right, female genitalia of E. saccharina

Plate 3: Far right, female genitalia of C. thyrsis

Plate 13: Top, male genitalia and aedeagus of Sesamia calamistis

Plate 14: Left, female genitalia of S. calamistis

Plate 19: Top and middle, male genitalia and aedeagus of Busseola fusca and B. phaia

Plate 20: Far left and second from left, female genitalia of B. fusca and B. phaia

Plate 23: Top to bottom, adults of E. saccharina, C. partellus and C. thyrsis

Plate 28: Top, adult B. fusca.





